



The Open Agriculture Journal

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REVIEW ARTICLE

In vitro Methodology to Assess Quantitative Resistance in Plant-fungus Pathosystems

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Abstract:

Background:

Quantitative resistance (QR) to fungal pathogens remains a primary focus of all major host breeding programs. Field screenings for resistant plants, although accurate and effective, demand significant time and a sizable workforce to accomplish. Moreover, weather conditions in the field may not always be favorable for uniform disease spread, which eventually may lead to the failure of the overall experiment. The development of a more efficient and reliable bioassay to screen for resistance to fungal pathogens would be advantageous for any breeding program working on disease resistance; however, only if it correlates with field screening trials.

Objective:

The objective of this review is to combine novel findings related to rapid screening methods to evaluate QR, which are needed to accelerate the progress in developing fungal disease resistance in cultivars. Insights into the *in vitro* quantitative components of the host-pathogen interaction, factors affecting *in vitro* evaluation in young plant materials, as well as molecular pathways for the association between the *in vitro* and adult plant responses to fungal infection are also reviewed.

Conclusion:

An *in vitro* method was found to be efficient and successful in terms of inoculum volume, plant samples, and working space. The main advantage of this method is its predictive ability for adult plant disease. In addition, it provides reproducible results and is found to be a simple and reliable method. The *in vitro* assay allows rapid and early determination of resistance/susceptibility to fungal pathogens, which can be incorporated into a breeding program for identifying resistant plants.

Keywords: Breeding program, Field screening, *In vitro* quantitative resistance components, Young plant materials, Pathogens.

Article History

Received: May 24, 2022

Revised: August 30, 2022

Accepted: September 20, 2022

1. INTRODUCTION

When infection is possible, interactions between fungal pathogens and plants can be characterized by several quantitative traits related to disease development [1]. Overall, the values arising from these quantitative traits result from the host effect (quantitative resistance [QR]), the pathogen effect (quantitative pathogenicity, often called aggressiveness in the plant pathology literature), and their interaction. QR in agricultural crops against fungal pathogens is characterized by a continuous variation ranging from very low to moderate levels of resistance [2]. QR is characterized by a reduced rate of epidemic development in the host population by altering spore infection efficiency, time from infection to sporulation,

lesion development rate, and the abundance of spores produced, and therefore the severity of disease (DS) [3]. These quantitative disease resistance components follow directly from the differential-difference equation proposed by Van der Plank [4] as a model for epidemic development. QR is generally thought to be more durable than qualitative disease resistance and is therefore being used to an increasing extent in crop protection [5]. Qualitative resistance, determined by the presence of R genes, only reduces the initial inoculum by removing avirulent spores [4]. QR has, until recently, been used less in breeding programs than qualitative resistance, mainly because the latter has been easier to incorporate into desirable agronomical genotypes [6].

Disease resistance is a sustainable solution to plant disease control due to environmental and economic factors [2, 5]. Among several methods used to assess the level of resistance

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among plants or genotypes, field screening through natural or artificial infection is preferred as it matches the situation that a producer confronts while growing crops [4]. However, field screening has limitations since it depends on proper environmental conditions, such as it can often be carried out only once a year, is time-consuming and expensive [7]. In addition, field screening demands substantial time, area and workforce to maintain the number of plants required for an experiment [8]. Moreover, uncontrollable environmental conditions, such as humidity, temperature, and the simultaneous presence of other pathogens in the field, may not always be favorable for disease propagation, which may lead to the failure of the overall experiment [9]. The whole plant inoculation method in the greenhouse has some major logistical disadvantages in terms of resources and destructive phenotyping [10]. Only a limited number of cultivars can be evaluated because of space limitations, and the risk of cross-contamination also reduces the number of assays that can be performed at a given time. More often, such limitations restrict quick progress in breeding plants for disease resistance [11 - 13].

The development of a more efficient, simple and reliable bioassay for resistance screening of facultative and obligate parasites without the limitations posed by field and greenhouse trials would be advantageous for any breeding program working on disease resistance [14]. An *in vitro* assay, being a non-destructive (for the whole plant) disease evaluation method, can facilitate mass and rapid germplasm screening [15 - 18], one of the key elements in plant-breeding programs [19]. It would require much less space and inoculum and allows a specified plant to be evaluated with multiple pathogens [20]. In several plant-fungal pathosystems, *in vitro* assays appear more precise and reproducible than field screenings; thus, the results are not prone to fluctuations in environmental conditions [21 - 23]. It allows the screening of a single plant against multiple pathogens at a time. Moreover, this method can be utilized while screening a segregating population where each plant has unique genetics; therefore, seeds from those plants can be saved for further breeding purposes. *In vitro* assays have been used to differentiate the level of resistance in several plants, ranging from herbaceous crops to large tree species, with a high success rate [24 - 26]. *In vitro* assays are reliable and efficient alternatives to field and greenhouse screening methods and may be used for mass screening of host plants to facilitate fungal resistance breeding [27 - 29]. Furthermore, cultivars identified as resistant based on these rapid screening methods must be evaluated under field conditions as the final confirmatory test [30]. The detached leaf disease phenotyping technique has been extensively used and reported as a rapid and alternative tool for screening several plant species for resistance against numerous biotrophic, hemibiotrophic and necrotrophic fungal pathogens [7 - 10, 30 - 33]. The objective of this review is to combine novel findings related to quick screening methodologies to assess QR, which are requested to speed up the progress in developing resistant cultivars to fungal infection. Insights into the *in vitro* quantitative components of the host-pathogen interaction, factors affecting *in vitro* evaluation in young plant materials, as well as molecular pathways for the association between the *in vitro* and adult

plant responses to fungal infection are also reviewed.

2. INSIGHTS INTO THE *IN VITRO* METHODOLOGIES USED IN THE HOST-FUNGAL PATHOGEN PATHOSYSTEMS

Resistance against fungal pathogens remains a primary focus of all major host breeding programs [2]. The development of genetically resistant cultivars is a cost-effective and desirable option to reduce yield losses caused by fungal pathogens [5]. Although QR to fungal diseases has been identified, it is considerably more challenging to assess than qualitative resistance [3, 5]. In the process of resistance breeding, screening for fungal resistance needs to be performed with accuracy, which is conducted either in a greenhouse or in a field by applying the whole plant assay technique [4]. However, both involve tedious procedures and have challenges [34 - 41]. Experimental efficacy for the fungal-plant host pathosystem could be improved considerably by reducing inoculum production time, increasing the reliability of inoculum production, and increasing the number of cultivars that can be evaluated in each experiment, especially when large numbers of fungal isolates are to be screened for comparisons of pathogenicity [42 - 49].

The advanced *in vitro* assay allows better control over humidity, temperature and a large number of replicates with less inoculum and may therefore be an option to eliminate the limitations [50 - 57]. *In vitro* techniques have the advantage of being more efficient in terms of the time of execution and the number of revealed components of QR, such as the disease intensity, latent period, and incubation period [7 - 10]. In addition, the inoculation of various living plant parts, such as seeds, fruits/heads, stems, leaves, and roots, has been used successfully in many plant-fungal systems. The required host material for each *in vitro* assay is small compared to the whole plant disease evaluation method (Fig. 1).

The advantageous features of this method reside in its predictive ability of adult plant disease; the results from the *in vitro* assay are considerably correlated and comparable with the results of screening under greenhouse and field conditions in many fungal-plant intersections [7 - 10]. Using ranking studies, the relative resistance levels of host cultivars under *in vitro* experiments appear comparable to resistance levels in the field or greenhouse, indicating that *in vitro* method predicts resistance occurring at the earliest and latest host development stages during disease infection [7 - 10]. The *in vitro* assay can prevent unnecessary interactions between pathogens during host evaluation for resistance to different pathogens or different isolates/strains of the same fungus [34]. With this method of screening, a host plant can be screened repeatedly because inoculations can be done at any growth stage without affecting the growth and seed production of the rest of the plant. This method also saves time in breeding for resistance and works well on newly expanding leaves because they appear to have less contamination [34]. The *in vitro* method can be used during the simultaneous evaluation of progenies for multiple resistance as a single plant can be evaluated several times at any growth stage without affecting the growth of the plant, thus, facilitating a selection program at a reduced cost [36].

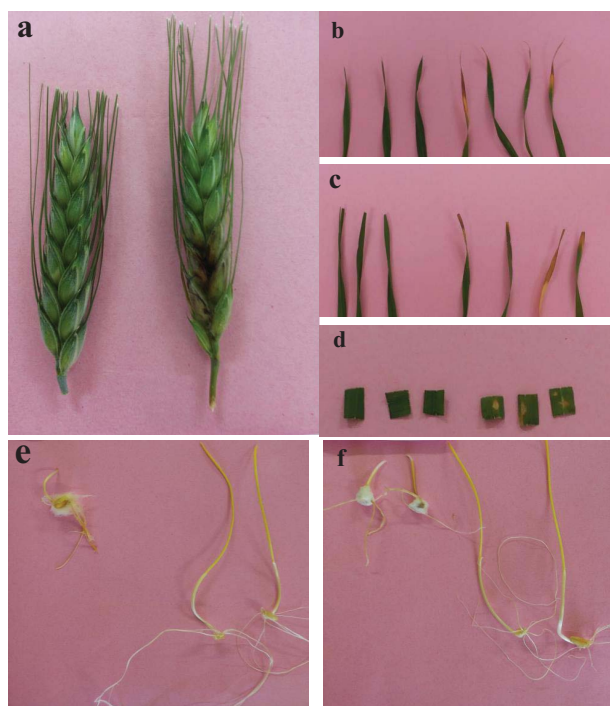


Fig. (1). Whole plant and *in vitro* inoculation techniques used to assess Fusarium head blight resistance on plant materials of durum wheat infected with *F. culmorum*, (a) floret inoculation assay, (b) foliar-spraying assay, (c) clip-dipping assay, (d) detached-leaf assay, (e) pin-point test assay (f) modified Petri-dish assay. Adapted from Sakr [11].

3. MEASURING *IN VITRO* QUANTITATIVE COMPONENTS OF THE HOST-PATHOGEN INTERACTION

QR can be selected in breeding programs either based on overall symptom expression or by prior identification of resistance mechanisms or correlated variables, followed by selection for these mechanisms. In the latter, the first step towards efficient selection for QR is the determination of *in vitro* resistance components. No single measure is suitable, and each plant–fungus interaction is likely to be different [58]. The three *in vitro* components include reduced infection frequency or density, reduced lesion size or concentration, and reduced propagule production per unit of host tissue over a period [59]. In most plant–fungus interactions involving necrotrophs, hemibiotrophs and biotrophs, the major epidemiological components of QR are infection efficiency (IE), latent period (LP), sporulation rate (SR), and lesion length (LL) [60]. Some *in vitro* components are more important than others in characterizing QR, and it is possible for some host–fungus interactions to recognize one component of QR, or a set of subcomponents, that adequately represent the QR in the field. For example, in *Puccinia hordei*/barley leaf rust and *Phytophthora infestans*/potato late blight, LP and LL were the most suitable *in vitro* components to assess field QR, respectively [61, 62]. The identification of single components may be especially useful in material that has been selected for QR over a long period but less in recently accessed material [63]. In populations that have not been strongly selected for QR, no or negative correlation of components is expected.

3.1. Infection Efficiency

IE is defined as the probability that a spore deposited on a receptive host surface produces a lesion in the absence of competitive interactions. It is usually measured as a percentage of successful infections resulting from a controlled number of deposited spores [64]. For practical reasons, IE may be indirectly measured by the observed numbers of chlorotic flecks or lesions per unit of leaf area [65]. This trait is difficult to precisely estimate because it depends on the number of spores deposited as well as on microclimatic conditions, which are difficult to control [66].

3.2. Latent Period

LP is the time interval between infection and the onset of sporulation from that infection. It determines the duration of epidemic cycles and thus largely controls the rate of epidemic development. The definition of LP is clear when applied to a single lesion. In most experimental studies, however, artificial inoculations result in many infections per leaf, and the variation in observed LP among infection sites on a leaf may be considerable [67]. To cope with this difficulty, several criteria are used to estimate LP, such as the time from inoculation to first sporulation [68] or the time needed for half of the final number of lesions to sporulate [69] or to show apparent sporulation structures [70]. The most precise method for estimating the time needed for half of the final number of lesions to sporulate was proposed by Shaner [67] and is based on an adjustment of the dynamics of lesion emergence to a sigmoid curve. Since LP is highly dependent on temperature, it is recommended to express the time in degree-days to allow comparisons between different experiments [71]. It has been observed that using different methods to measure LP (i.e., the time needed for half of the final number of lesions to sporulate vs. the time to first sporulation) could lead to differences in its estimation as observed for *Puccinia triticina*/wheat [65] and *Phytophthora infestans*/potato [69]. Such differences might, of course, result from uncontrolled environmental effects. A more interesting alternative is that variability in LP among fungal isolates could reveal heterogeneity in both the time at which the first sporulation occurs and the dynamics of lesion maturation [72].

3.3. Sporulation Rate

SR is the number of spores produced per lesion and per unit of time [73, 74]. In practice, spores are either weighed [75] or counted [76]. Sporulation is sometimes expressed in spore production per unit area of diseased leaf [75] or relative to lesion size [76]. Spore production per lesion is highly density-dependent [73]. It can be useful to consider the spore production per unit area of sporulating tissue [76], which is considerably less density-dependent [77]. The number of spores produced by a diseased plant will determine the quantity of inoculum that can infect neighboring plants but also indicates the interaction between the host plant and the parasite since the quantity of spores produced depends on the aggressiveness of the parasite and the level of host QR [4]. It also reflects the capacity of the fungus to invade host tissues during the incubation period.

The infectious period is the time from the beginning to the end of sporulation. This component is difficult to precisely estimate since sporulation often shows an early peak, followed by an asymptotic decrease [77]; however, more irregular patterns may be obtained [78]. For cereal rusts, sporulation can last for more than 40 days under controlled conditions on adult plants [77].

Since many pathogen species have two or more forms of propagule related to sexual or asexual reproduction [79], infection efficiency, sporulation rate, latent period, and infectious period may, in principle, be measured for each type of spore [80]. Nevertheless, a given parameter may not have the same meaning when measured on sexual and asexual spores. For example, the latent period associated with sexual spores is very different from that of asexual spores because it depends on the fortuitous encounter and merger of two sexually compatible lesions. Therefore, while there seems to be room for adaptive adjustment of asexual latency, sexual latency is expected to be highly dependent on environmental stochasticity [1]. Moreover, organs resulting from sexual reproduction often ensure inter-season survival, as in *Blumeria graminis* or *Leptosphaeria maculans*, and the latent period, as defined above, has no meaning in such cases [1].

3.4. Lesion Length

LL is another quantitative trait that is measured as a QR component [81]. It is generally defined as the surface area that produces spores. For some pathogens, such as *Puccinia triticina*/leaf rust, lesion size remains limited, but it can dramatically increase in some species, such as *Puccinia striiformis* var. *striiformis*/stripe rust or *Phytophthora infestans*/late blight, for which lesion growth is semisystemic [82]. In this case, LL accounts for a large part of the quantitative development of epidemics and lesion growth rate is a key factor in pathogen competition for available host tissue. LL is not easy to precisely determine for pathogens, such as *Mycosphaerella graminicola*/Septoria blotch, that induce necrosis on the host leaf [83]. Moreover, such pathogens often indirectly cause apical necrosis on the leaves that can be confused with a diseased area.

3.5. Dwarfing

Dwarfing is a symptom characteristic of plants systemically infected by fungal pathogens and is explained by a decrease in the concentration of growth hormone in infected tissue [4]. This decrease in size can be observed at a very early stage.

3.6. Disease Severity

QR is sometimes estimated through disease severity, measured as the percentage of the infected plant organ (root, leaf or fruit/spike) covered by pathogen lesions [84]. DS is a composite variable resulting from the integrated effect of infection efficiency and lesion size, but also, when assessed at the crop scale, sporulation and dispersal.

In any case, it is essential to measure quantitative traits that can be clearly related to disease dynamics and pathogen evolution. For instance, a few *in vitro* traits have been

identified that may be useful for the characterization of quantitative resistance by making comparisons with two cultivars that are susceptible to the pathogen in the field. The collective effects of each or some of the *in vitro* traits are particularly important.

4. EXAMPLES OF *IN VITRO* STUDIES ON DIFFERENT CROPS AND FUNGI

Under *in vitro* tests, commonly used parameters for resistance assessment are lesion growth rate, i.e., the rate of necrosis extension, infection efficiency, i.e., the percentage of successful infections, lesion size, latent period, and spore density [7 - 57]. These traits are well adapted to the description of the epidemic phase of polycyclic pathogens that belong to the fungi and oomycete groups [1]. Moreover, the *in vitro* traits mentioned above can easily be applied to pathogens with specific biological features [1]. *In vitro* assays have been used as a tool for investigating QR in the whole plant for a wide range of important fungal diseases, including rust, powdery mildew, downy mildew, Fusarium head blight, etc. (Table 1). Table 1 presents *in vitro* applications used to assess quantitative resistance in several fungal-plant pathosystems.

4.1. A Necrotrophic Fungus

A detached leaf assay was used to determine the reaction of five barley genotypes to spot blotch caused by *Cochliobolus sativus* [25]. The estimation of the infected leaf area enabled discrimination among barley genotypes differing in their susceptibility to the pathogen. Significant correlations were found ($P=0.001$) between *in vitro* values in both seedling ($r=0.89$) and adult ($r=0.95$) plants. The established *in vitro* assay enabled a fast assessment of the susceptibility of barley to spot blotch and should be useful for many types of studies on this disease [25].

4.2. A Hemibiotrophic Fungus

The detached leaf assay was successful in the identification of an important component of the resistance of *Fusarium* species, causing head blight (FHB) in the European wheat germplasm. This *in vitro* method may be a useful mechanism to discriminate amongst different types of resistance in a breeding program [8]. *Microdochium majus* is used in the detached leaf assays to detect leaf symptoms for observing QR components (incubation period, latent period and lesion length) [12]. Lesion length is one of the components of QR measured as an indicator of fungal pathogenicity and aggressiveness. QR components detected in the *M. majus* detached leaf assay have been correlated to FHB resistance in wheat inoculated with *F. culmorum* and *F. graminearum* [8, 12]. FHB resistance in a seed germination assay was highly correlated in *F. graminearum*, *F. avenaceum*, *F. culmorum*, *M. majus* and *Microdochium nivale*, indicating common resistance between *M. Majus* and other *Fusarium* spp. in the *in vitro* assay [12]. Browne and Cooke [13] reported that QR components (incubation periods, longer latent periods and shorter lesion lengths) in the detached leaf assay and higher germination rates in the seed germination assay were related to greater FHB resistance (Type II). However, the exotic wheat germplasms, which provide highly effective resistances to FHB resistance,

do not appear to be detected in the detached leaf or seed germination assays [8, 12].

Table 1. Fungal-plant pathosystems on which *in vitro* applications were used to assess quantitative resistance.

Host Plant	Fungal Pathogen/ Disease	Targeted Young Plant Materials	Analyzed <i>in vitro</i> Components	Reliability of <i>in vitro</i> Test to Predict Resistance at Adult Plant Stage	References
Wheat	Several Fusarium and <i>Microdochium</i> species/ Fusarium head blight	Detached leaves Seedlings Detached heads	Latent period (LP), incubation period (IP) and lesion length (LL) Area under disease progress Curve (AUDPC), coleoptile length reduction and LL Disease incidence and disease severity (DS)	Yes Yes Yes	[8, 12, 13] [11, 14 - 16] [17]
	<i>Septoria tritici</i> /blotch	Detached leaves	The percentage leaf area covered by lesions bearing pycnidia	Yes	[18]
	<i>Septoria nodorum</i> /blotch	Detached seedling leaves	DS	Yes	[19]
	<i>Puccinia triticina</i> /leaf rust	Detached leaves	Disease symptoms and the rate of pustule development Incubation and latent period	Yes Yes	[20] [21]
Barley	Several Fusarium and <i>Microdochium</i> species/Fusarium head blight	Detached leaves Detached leaves Seedlings Seedlings Detached heads	LP and IP LP, LL, AUDPC and percentage of infected seedlings Root weight, coleoptile length, coleoptile weight and seed germination reductions Disease incidence and severity	Yes Yes Yes Yes	[13] [22] [23] [10]
	<i>Cochliobolus sativus</i> /common root rot	Seedlings	Percentage infected area of the sub-crown underdone	Yes	[24]
	<i>Cochliobolus sativus</i> /spot blotch	Detached leaves	Infected leaf area	Yes	[25]
	<i>Puccinia striiformis</i> f.sp. <i>hordei</i> / stripe rust	Detached leaves	DS	Not analyzed	[26]
Oat	<i>Microdochium nivale</i> /Fusarium head blight	Detached leaves	LP	Yes	[13]
Maize	<i>Puccinia sorghi</i> /rust	Detached leaves	DS	Not analyzed	[27]
	<i>Bipolaris maydis</i> /southern corn leaf blight	Detached leaves	Percentage leaf area	Yes	[28]
Rice	<i>Magnaporthe grisea</i> /blast	Detached leaves	DS	Not analyzed	[29]
Banana	<i>Mycosphaerella fijiensis</i> /black leaf streak	Detached leaves	IP and leaf areas infected	Yes	[30]
St. Augustine grass	<i>Magnaporthe oryzae</i> /grey leaf spot	Detached stolon and detached leaves	LL and DS	Yes	[31]
Faba beans	<i>Uromyces viciae-fabae</i> /rust	Detached leaves	Percentage leaf area	Yes	[32]
	<i>Ascochyta fabae</i> /ascochyta blight	Detached leaves and stem segments with attached leaves	Number and size of lesions and the number of lesions bearing pycnidia	Yes	[33]
	<i>Colletotrichum lindemuthianum</i> / anthracnose	Detached leaves	DS	Not analyzed	[34]
	<i>Botrytis fabae</i> /chocolate spot	Detached leaves	DS	Not analyzed	[35]

(Table 1) contd.....

Host Plant	Fungal Pathogen/ Disease	Targeted Young Plant Materials	Analyzed <i>in vitro</i> Components	Reliability of <i>in vitro</i> Test to Predict Resistance at Adult Plant Stage	References
Pea	<i>Erysiphe pisi</i> /powdery mildew	Leaflet Detached leaves	Infection efficiency and spore production Percentage of leaf area affected with disease	Yes Yes	[6, 36]
Chickpea	<i>Ascochyta rabiei</i> /ascochyta blight	Detached leaves	DS	Yes	[37]
Alfalfa	<i>Sclerotinia trifoliorum</i> /clover rot	Excised leaf tissues	Rate and extent of necrosis	Yes	[38]
Soybean	<i>Phakopsora pachyrhizi</i> /rust	Detached leaves	Lesion size and number of spores	Yes.	[9]
Rose	<i>Diplocarpon rosae</i> /black spot, <i>Podospaera pannosa</i> / powdery mildew and <i>Peronospora Sparsa</i> /downy mildew	Detached leaves	DS	Yes	[39]
Sweet cherry	<i>Podospaera clandestine</i> / powdery mildew	Detached leaves	DS	Yes	[40]
Grapevine	<i>Erysiphe necator</i> /powdery mildew	Detached leaves	Percentage of inoculating spots leading to a colony and the mean diameter of colonies	Not analyzed	[41]
	<i>Plasmopara viticola</i> /downy mildew	Detached leaves	DS	Yes	[10]
Tomato	<i>Phytophthora infestans</i> /late blight	Detached leaves	LL	Yes	[41]
	<i>Alternaria solani</i> /early blight	Detached leaflets	Lesion radius, rate of lesion expansion, and final DS	Yes	[43]
Tobacco	<i>Phytophthora parasitica</i> <i>var. nicotianae</i> /black shank	Detached leaves	Number of lesions/leaf and percentage of leaf area infected	Yes	[44]
Potato	<i>Phytophthora infestans</i> /late blight	detached leaves and intact plants	DS	Yes	[45]
Cucumber	<i>Sphaerotheca fuliginea</i> / powdery mildew	Leaf blades	DS	Not analyzed	[46]
	<i>Didymella bryoniae</i> /gummy stem blight	Detached leaves	DS	No	[7]
Sunflower	<i>Plasmopara halstedii</i> /downy mildew	Seedlings	Percentage infection, LP, sporulation density and reduction of hypocotyls length	Yes	[47]
Spinach	<i>Peronospora effuse</i> /downy mildew	Detached leaves and cotyledons	DS	Yes	[48]
Quinoa	<i>Peronospora farinose</i> /downy mildew	Detached leaves	AUDPC, IP and LP	Yes	[49]
Lucerne	<i>Phytophthora medicaginis</i> /Phytophthora root rot	Detached leaves and intact cotyledons	DS	Cotyledon test predicted resistance at the adult plant stage	[50]
White lupin	<i>Diaporthe toxica</i> /phomopsis leaf blight	Detached leaves	DS	Yes	[51]
Cocoa	<i>Phytophthora palmivora</i> and <i>Phytophthora megakarya</i> / black pod	Detached leaves	DS	Yes	[52]
Fenugreek	<i>Cercospora traversiana</i> / Cercospora leaf spot	Detached leaves	The number of lesions and LL	Yes	[53]
Oak	<i>Phytophthora ramorum</i> /sudden oak death	Detached leaves	LL	Yes	[54]
Spindle tree	<i>Colletotrichum gloeosporioides</i> / leaf anthracnose	Detached leaves	Chlorotic spots	Not analyzed	[55]
Taro	<i>Phytophthora colocasiae</i> /leaf blight	Detached leaves	LL	Yes	[56]
Durian	<i>Phytophthora palmivora</i> /root rot	Detached leaves	DS	Yes	[57]

4.3. A Biotrophic Fungus

In vitro components of quantitative resistance in sunflower (*Helianthus annuus*) to *Plasmopara halstedii* [47], i.e., percentage infection, latent period, sporulation density, and reduction of hypocotyl length, were compared between two sunflower cultivars and showed different levels of quantitative resistance in the field. The moderately susceptible inbred line showed a higher percentage of infection, a higher sporulation density, a shorter latent period and less reduced hypocotyl length than the moderately resistant inbred line. The percentage infection of FU was 1.4% less than BT, the latent period of BT was 12.4% less than FU, the sporulation density of FU was 22.3% less than BT, and the reduced hypocotyl length of BT was 15.3% less than FU. It seems that the criteria, such as latent period, sporulation density and reduction of hypocotyl length, may be used to measure quantitative resistance in sunflowers to *P. halstedii* [47].

The above-mentioned studies have generally concluded that *in vitro* traits can be applied for measuring QR in several plant species infected with biotrophic, hemibiotrophic, and necrotrophic fungal pathogens.

5. FACTORS AFFECTING *IN VITRO* EVALUATION IN YOUNG PLANT MATERIALS

Several factors varying in importance affect *in vitro* evaluation in young plant materials, which include growth stage, senescence of plant parts, amending growth medium with plant hormones, incubation conditions, and the origin of genetic host and fungal materials.

5.1. Growth Stage

The level of expression of QR is known to vary with plant development [3, 4]. For example, QR is greater in late expanding leaves against ascochyta blight in chickpeas and faba beans [33, 37], powdery mildew and crown rust in oat [85, 86], powdery mildew in sweet cherries and rusts in grapes [40, 41], rust and Fusarium head blight wheat [87, 88], powdery mildew in leaf blight in taro [56] and southern corn leaf blight in maize [28]. The effect of leaf position was considerably more pronounced in the powdery mildew-grapevine interactions; the magnitude of differences between susceptible and resistant cultivars increased with leaves at positions 3-5. Data suggested that leaves at position 3 must be inoculated to clearly distinguish resistant from susceptible cultivars [41]. Senescing grapevine leaves became more resistant to grapevine powdery mildew, and this effect was evident in both the success of infection and the diameter of the colony [41]. Conversely, older cucumber leaves were more susceptible to gummy stem blight than younger leaves in the field, greenhouse, and detached-leaf tests [7]. Arraiano *et al.* [18], using a detached leaf assay in wheat, found that secondary leaves were more susceptible to *Septoria tritici* than first-expanding leaves. Important factors for downy mildew/*Plasmopara viticola* resistance screens include leaf age; the fourth fully expanded leaf corresponds well with vineyard ratings [89].

5.2. Senescence of Plant Parts

A critical aspect of the *in vitro* assay is the prevention of senescence of leaf pieces before the duration required to express symptom stages and disease severity levels necessary to differentiate among cultivar responses. The leaf senescence may have had an inhibiting effect on sporulation for a period before sporulation occurred profusely over leaf stomata [41]. The youngest tissues can be infected, but the development of the fungus is generally arrested during the aging of tissues before sporulation occurs [41]. Senescence of leaf tissue before symptom development altered the accuracy with which QR resistance was determined, which has implications for using detached leaf assays to determine resistance to other diseases [88]. Using whole plants, which prevents the senescence associated with detached leaves, Singh and Huerta-Espino [87] found that cultivar differences were most clearly observed in later-forming leaves, which showed greater resistance. In a detached leaf assay, Arraiano *et al.* [18] inoculated seedling leaves with an *S. tritici*, which showed a relatively long period between inoculation and symptom development prior to detaching the leaves and mounting them on water agar.

5.3. Amending Growth Medium With Plant Hormones

Amending agar medium with plant hormones aided the retention of green leaf color in detached leaves [9, 12, 26, 90]. Benzimidazole and cytokinin have been used in the incubation medium to prevent chlorophyll degradation in detached leaves. Detached wheat and barley leaves were kept green for 14 to 34 days on benzimidazole agar [26], detached wheat leaves were kept green for at least 14 days using kinetin [12], and detached oat leaves were kept green for up to 10 days using kinetin and 6-benzylaminopurine treatments [86], indicating that maintaining green detached leaves for much longer periods is essential to propagate fungi. Soybean leaf pieces in a medium containing kinetin at 10 mg/liter had 5% chlorosis 18 days after plating compared to leaf pieces in media amended with all other plant hormones, which had higher levels of chlorosis [9]. Longer periods of healthy leaf tissue are important for the short-term use of rust isolates [90].

5.4. Incubation Conditions

The incubation conditions of detached potato leaves in closed trays rather than detachment itself appeared to affect the late blight resistance expression, concluding that the lower expression of resistance in the detached leaf tests was due to differences in environmental conditions [45]. The constant, highly favorable environment a pathogen finds in the closed trays apparently enhances infection by the *Phytophthora infestans* zoospores. Incubating detached leaves in closed trays appears to decrease resistance expression. Therefore, a suitable experimental condition must be chosen depending on the aim of the experiment. When the expression of resistance is to be examined on detached leaves, the reduced level of resistance should be measured against the low infection frequency inherent to intact plants. Inoculation of intact plants is preferred, but in most cases, the inoculation of detached leaves incubated in covered trays appears to be an adequate alternative [45]. Relative humidity during incubation in closed containers was 85% and may have proved lethal to the more

fragile *Phytophthora colocasiae* zoospores (taro leaf blight) without an agar drop to protect them [56].

5.5. Origin Of Genetic Host And Fungal Materials

The origin of genetic host and fungal materials plays a critical role in evaluating *in vitro* resistance reactions. Brown and Cooke [12] did not find a relationship between the detached leaf assay and whole plant FHB resistance in International Maize and Wheat Improvement Center (CIMMYT) genotypes containing more diverse sources of resistant germplasms in their pedigree. The detached leaf assay did not appear to detect the whole plant resistance of Sumai 3, a Chinese wheat cultivar, because it displayed shorter incubation and latent periods and longer lesion lengths relative to European cultivars. Moreover, the detached leaf technique did not detect the greater FHB resistance of Frontana wheat cultivar because this genotype had a comparable incubation period, latent period, and lesion length to the most resistant Irish and UK cultivars possessed only moderate FHB resistance [12]. Although considerable differences in mean *Plasmopara viticola*/downy mildew severity separated *Vitis vinifera*, *Vitis* hybrid, *V. riparia*, and *V. labrusca* during *in vitro* inoculations (from susceptible to resistant), notable intra-species variation was identified for all well-represented species [89]. The effect of the pathogen source on resistance ratings could reflect genetic variation in *P. viticola*; the lack of disease severity was much more likely due to resistance against the single isolates screened [89].

Certain precautions are required while conducting detached-leaf assays. Leaves should be free from mites because they forage on rust spores, which in turn, reduce the efficiency of infection. Surface sterilization of leaves with 0.1% NaOCl or 70% ethanol should also be avoided because it causes patches of necrosis that interfere with pathogen development [9].

6. MOLECULAR PATHWAYS FOR THE CORRELATION BETWEEN THE *IN VITRO* AND WHOLE PLANT REACTIONS TO FUNGAL INFECTION

Molecular pathways for the relationship between the *in vitro* and whole plant responses to fungal infection have been reviewed in the FHB-wheat pathosystem [15]. The genetic basis of FHB resistance is highly complex but is beginning to be understood. Quantitative trait loci (QTLs) for resistance have been linked with passive factors, such as plant height, spike architecture and flowering date [91], and with variability in specific genes. Some specific compounds of wheat anthers (choline, betaine) were thought to be responsible for fungal growth stimulation [92], but in a later study, a substantial role of floral structures in resistance to *Fusarium graminearum* was denied [93]. Multiple signaling pathways involved in spike response to *F. graminearum* infection [94] have been found to be similarly regulated in inoculated seedlings [95]. Moreover, some up-regulated genes during infection affect plant growth and stem cell division [95]. It is possible that lipid transfer and deoxynivalenol resistance genes associated with 5A (*Qfhs.ifa-5A*) and 3B (*Qfhs.ndsu-3BS*) QTLs in common wheat [96] are also components of resistance at the seedling stage. In

addition, the observed correlations between parameters of coleoptiles and roots within each experiment could reflect concurrent gene expression in different tissues/organs. An improved understanding of these putative parallel mechanisms is expected as comparative gene expression studies across developmental stages increase in number.

The above-mentioned studies have generally concluded that the biological explanation for an association between the seedling and adult plant responses to infection remains unknown, but it can be hypothesized that similar genetic pathways become activated at both developmental stages.

7. OTHER APPLICATIONS OF *IN VITRO* TECHNIQUES IN THE HOST-FUNGAL PATHOGEN PATHOSYSTEMS

Reliable *in vitro* tests have been extensively used for studying plant-fungal interactions at the physiological or molecular levels, highlighting that the use of *in vitro* techniques is not only limited to evaluating QR components but extends to several fungal-plant life history traits.

7.1. Studying Plant-Fungal Interactions At The Physiological Level

Production of single-strain cultures of obligate pathogens for use in genetic studies is difficult. By using *in vitro* techniques, the ability to handle and propagate pathogens on the young plant materials helped to generate pure cultures and avoid cross-contamination for *Podosphaera fusca*, causing cucumber powdery mildew, *Pseudoperonospora cubensis* causing cucumber downy mildew, *Plasmopara halstedii* causing sunflower downy mildew, *Puccinia coronata* causing crown rust, *Phakopsora pachyrhizi* causing soybean rust, and *Peronospora effuse* causing spinach downy mildew [48, 86, 90, 97, 98]. As a pre-screening test, the effect of potential resistance-inducing chemicals, DL-3-aminobutyric acid, Bion (benzo-(1,2,3) thiadiazole-7-carbothioic acid S-methyl ester), and a foliar fertilizer containing potassium phosphite, on development of *Microdochium majus* causing Fusarium head blight was studied in detached leaves [99]. Reduced disease development of *M. majus* was also only observed in detached leaves pre-treated with the foliar fertilizer, indicating that the effect on disease development was at least partly due to a fungistatic effect [99]. The *in vitro* technique also performed well in testing the pathogenic behavior of *Erysiphe necator* on grapevine [41], *Plasmopara halstedii* on sunflower [100, 101], *Mycosphaerella fijiensis* on bananas [30], *Phytophthora infestans* on potato [102], *Fusarium* and *Microdochium* species causing a head blight on wheat and barley [16, 17, 103 - 105] and *Phytophthora cactorum*, *P. citrophthora*, *Pythium dissotocum* complex, *Py. aphanidermatum*, *Globisporangium heterothallicum*, *G. ultimum*, *Phytophythium vexans*, *Phy. mercurial*, and *Phy. litorale* on strawberries [106]. The *in vitro* screening method was effective in determining the pathotype of *Pseudocercospora griseola* isolates on a set of angular leaf spot differential common bean cultivars [107] and host-specialization in *E. necator-V. vinifera* interactions [41] and four *Fusarium* species causing a head blight on wheat and barley [108 - 110]. The *in vitro* laboratory assay was used efficiently to determine the sensitivity of *P. infestans* and two

rust fungi to fungicides [111, 112]. Phytotoxins as selective agents under *in vitro* selection were utilized for improved disease resistance [113].

7.2. Studying Plant-Fungal Interactions At The Molecular Level

The detached leaf assay provided a means to enable fundamental studies on the defense mechanisms of Arabidopsis in response to *F. graminearum*/Fusarium head blight [114]. *In vitro* infection of detached leaves *Puccinia substriata*, the causal agent of rust disease, and *Sclerospora graminicola*, the causal agent of downy mildew, resulted in a significant reduction of disease symptoms in transgenic pearl millet cultivars in comparison to wild-type control plants; the disease resistance of pearl millet was increased by up to 90% when infected with two diverse, economically important pathogens [115]. On detached leaves, virus-induced gene silencing was adopted to dissect gene functions in cotton resistant to Verticillium wilt [116]. Using *in vitro* methods, transgenic lines of tomato harboring rice chitinase gene were evaluated for resistance to *Fusarium oxysporum* f. sp. *lycopersici* causing Fusarium wilt and *Alternaria solani* causing early blight. *In vitro* studies concluded that introducing a chitinase gene in a susceptible cultivar of tomato not only enhanced the resistance but was stably inherited in transgenic lines [117]. Using detached leaf assay, two novel powdery mildew resistance loci, *Ren6* and *Ren7*, were identified from the wild Chinese grape species *Vitis piasezkii* [118]. Phenotyping of transformed potato cultivars developed through small interfering RNA (siRNA) and artificial micro RNA (amiRNA) techniques for late blight resistance was undertaken using *in vitro* assays. The siRNA and amiRNA-derived cultivars exhibited less lesion area as well as spore production and were categorized as moderately resistant [41].

7.3. Using *In Vitro* Techniques To Explore Defense Responses In Plants To Fungal Infection

Wheat endophytes, *Phoma glomerata*, *Aureobasidium proteae* and *Sarocladium kiliense*, were screened as biological control agents against Fusarium head blight using two different *in vitro* tests [119]. This study pointed out that the test on detached wheat spikelets provided information about the potential pathogenicity, growth capacity, and efficacy of the endophyte strains on the targeted plant before testing them on whole plants [119]. Detached leaves of the Arabidopsis lines were inoculated with *Verticillium dahliae* to evaluate the defense response under drought stress; leaves from transgenic plants showed increased callose deposition and reduced mycelia growth [120]. The effect of chemical treatments affecting the ethylene pathway was studied in detached head assays, and the results showed that the ethylene signaling could mediate wheat Fusarium head blight resistance to *Fusarium graminearum* [121]. A notable difference was found between non-amended and amended treatments with silicon on young plant parts in potato detached leaves infected with *Phytophthora infestans* (late blight) [122] and wheat detached leaves and seedlings challenged with four Fusarium head blight pathogens [123]. By *in vitro* techniques, the direct fungitoxicity was investigated for Fusarium head blight pathogens and

Cochliobolus sativus causing spot blotch and common root rot [124]. Recently, Sakr [125, 126] explored how the deployment of QR in wheat and barley affects aggressiveness under *in vitro* conditions, leading to potential resistance erosion.

7.4. Using *In Vitro* Techniques In Plant Breeding

In breeding programs and during gene pyramiding, it will be challenging to evaluate plant symptoms using only conventional screening methods; however, this alternative rapid detached leaf method is proposed for marker-assisted gene pyramiding in combination with molecular markers to aid the selection of resistant progenies during each backcross generation [107]. In addition, the proposed method can help when evaluating progenies for different pathotypes using leaves from the same plant, which could be difficult with the conventional evaluation method. Thus, this alternative screening technique could provide a great opportunity for breeders working in gene pyramiding and marker-assisted back-crossing programs to evaluate common bean genotypes and progenies for multiple disease resistance [107].

CONCLUSION

Besides tolerance to abiotic stresses like salinity and drought, plant breeders should also take into account the resistance to fungal pathogens. Therefore, further studies should be carried out on the pathogens, their life cycle, QR resistance components, and the environment by host-fungal interactions to develop new cultivars that resist the damage caused by fungal pathogens. The *in vitro* technique developed, optimized, and validated during several studies can be used to rapidly detect plant genotypes with superior resistance to fungal infection. The *in vitro* screening technique provides the plant breeders' community with highly informative parameters and allows them to establish diversified host germplasm and enhance breeding program efficiency. The *in vitro* assay can be conducted in a relatively short period of time, making it a convenient protocol for the evaluation of a large number of progeny at early breeding stages. Hence, evaluation of fungal resistance under greenhouse or field conditions could be restricted to a narrower sample of selected genotypes with the best performance in the seedling assay. The *in vitro* assay has a low-cost approach and can be carried out off-season. This assay may also constitute a model system for biochemical and genetic studies on plant defense responses.

LIST OF ABBREVIATIONS

QR	=	Quantitative Resistance
IE	=	Infection Efficiency
LP	=	Period
SR	=	Sporulation Rate
LL	=	Lesion Length
FHB	=	Fusarium Head Blight
QTLs	=	Quantitative Trait Loci

CONSENT FOR PUBLICATION

Not applicable.

FUNDING

This study is financially supported by the Atomic Energy Commission of Syria.

CONFLICT OF INTEREST

The author declares no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

The author would like to thank the Atomic Energy Commission of Syria for providing assistance with this research. The author would also like to thank the reviewers for their constructive comments on this manuscript.

REFERENCES

- [1] Lannou C. Variation and selection of quantitative traits in plant pathogens. *Annu Rev Phytopathol* 2012; 50(1): 319-38. [http://dx.doi.org/10.1146/annurev-phyto-081211-173031] [PMID: 22702351]
- [2] Mundt CC. Durable resistance: A key to sustainable management of pathogens and pests. *Infect Genet Evol* 2014; 27: 446-55. [http://dx.doi.org/10.1016/j.meegid.2014.01.011] [PMID: 24486735]
- [3] Geiger HH, Heun M. Genetics of quantitative resistance to fungal diseases. *Annu Rev Phytopathol* 1989; 27(1): 317-41. [http://dx.doi.org/10.1146/annurev.py.27.090189.001533]
- [4] Van Der Plank JE. *Disease Resistance in Plants*. 1st ed. New York, USA: Academic Press 1968; p. 208.
- [5] Johnson R. A critical analysis of durable resistance. *Annu Rev Phytopathol* 1984; 22(1): 309-30. [http://dx.doi.org/10.1146/annurev.py.22.090184.001521]
- [6] Viljanen RSLH, Gaunt RE, Frampton CMA, Falloon RE, McNeil DL. Components of quantitative resistance to powdery mildew (*Erysiphe pisi*) in pea (*Pisum sativum*). *Plant Pathol* 1998; 47(2): 137-47. [http://dx.doi.org/10.1046/j.1365-3059.1998.00217.x]
- [7] Amand PCS, Wehner TC. Greenhouse, detached-leaf, and field testing methods to determine cucumber resistance to gummy stem blight. *J Am Soc Hortic Sci* 1995; 120(4): 673-80. [http://dx.doi.org/10.21273/JASHS.120.4.673]
- [8] Browne RA, Murphy JP, Cooke BM, et al. Evaluation of components of Fusarium head blight resistance in soft red winter wheat germ plasm using a detached leaf assay. *Plant Dis* 2005; 89(4): 404-11. [http://dx.doi.org/10.1094/PD-89-0404] [PMID: 30795457]
- [9] Twizeyimana M, Ojiambo PS, Ikotun T, Paul C, Hartman GL, Bandyopadhyay R. Comparison of field, greenhouse, and detached-leaf evaluations of soybean germplasm for resistance to *Phakopsora pachyrhizi*. *Plant Dis* 2007; 91(9): 1161-9. [http://dx.doi.org/10.1094/PDIS-91-9-1161] [PMID: 30780658]
- [10] Prajongjai T, Poolsawat O, Pornbungkerd P, Wongkaew S, Tantasawat PA. Evaluation of grapevines for resistance to downy mildew (*Plasmopara viticola*) under laboratory and field Conditions. *S Afr J Enol Vitic* 2014; 35: 43-50.
- [11] Sakr N. *In vitro* quantitative resistance components in wheat plants to Fusarium head blight. *Open Agric J* 2019; 13(1): 9-18. [http://dx.doi.org/10.2174/1874331501913010009]
- [12] Browne RA, Cooke BM. Development and evaluation of an *in vitro* detached leaf assay for pre-screening resistance to Fusarium head blight in wheat. *Eur J Plant Pathol* 2004; 110(1): 91-102. [http://dx.doi.org/10.1023/B:EJPP.0000010143.20226.21] [http://dx.doi.org/10.1007/s10658-005-2077-z]
- [13] Shin S, Kim KH, Kang CS, et al. Simple method for the assessment of Fusarium head blight resistance in Korean wheat seedlings inoculated with *Fusarium graminearum*. *Plant Pathol J* 2014; 30(1): 25-32. [http://dx.doi.org/10.5423/PPJ.OA.06.2013.0059] [PMID: 25288982]
- [14] Soresi D, Zappacosta D, Garayalde A, Miranda R, Carrera A. An *in vitro* assay for pre-screening resistance to Fusarium head blight in durum wheat. *Phytopathol Mediterr* 2015; 54: 253-64.
- [15] Sakr N. The use of *in vitro* area under disease progress curve to predict quantitative traits in the Fusarium head blight-small grain cereal pathosystem. *Pak J Phytopathol* 2021; 33(2): 303-12. [http://dx.doi.org/10.33866/phytopathol.033.02.0694]
- [16] Sakr N, Al-Attar J. Screening for wheat resistance and pathogenicity of Fusarium head blight species using an *in vitro* detached head test. *Arch Phytopathol Pflanzenschutz* 2021; 54(19-20): 2423-39. [http://dx.doi.org/10.1080/03235408.2021.1984747]
- [17] Arraiano LS, Brading PA, Brown JKM. A detached seedling leaf technique to study resistance to *Mycosphaerella graminicola* (anamorph *Septoria tritici*) in wheat. *Plant Pathol* 2001; 50(3): 339-46. [http://dx.doi.org/10.1046/j.1365-3059.2001.00570.x]
- [18] Benedikz PW, Mappedoram CJ, Scott PR. A laboratory technique for screening cereals for resistance to *Septoria nodorum* using detached seedling leaves. *Trans Br Mycol Soc* 1981; 77(3): 667-9. [http://dx.doi.org/10.1016/S0007-1536(81)80126-X]
- [19] Boydom A, Dawit W. Evaluation of detached leaf assay for assessing leaf rust (*Puccinia triticina* Eriks.) resistance in wheat. *J Plant Pathol Microbiol* 2013; 4: 176.
- [20] Patial M, Kumar J, Pal D. Detached leaf assay for evaluating resistance to leaf rust *Pst.* 104-2 in wheat (*Triticum aestivum* L.). *Indian J Exp Biol* 2017; 55: 789-94.
- [21] Sakr N. Components of quantitative resistance in barley plants to Fusarium head blight infection determined using three *in vitro* assays. *J Plant Prot Res* 2018; 58: 176-83.
- [22] Sakr N. An efficient *in vitro* assay to predict resistance and pathogenicity in the Fusarium head blight-*Hordeum vulgare* pathosystem. *Open Agric J* 2020; 13: 9-18. [http://dx.doi.org/10.2174/1874331501913010009]
- [23] Arabi MI, Jawhar M. A rapid technique for determining common root rot reaction in barley. *Plant Breed* 1999; 118(3): 278-80. [http://dx.doi.org/10.1046/j.1439-0523.1999.118003278.x]
- [24] Arabi MIE, Jawhar M. Barley reaction to *Cochliobolus sativus* based on detached first leaf. *Australas Plant Pathol* 2007; 36(5): 475-7. [http://dx.doi.org/10.1071/AP07053]
- [25] Osman GN. The uses and limitations of detached leaf culture for the assessment of reaction type in yellow rust of barley. *Cereal Rusts Bull* 1982; 10: 16-21.
- [26] Kushalappa AC, Hegde RK. Studies on maize rust *Puccinia sorghi* in mysore state I. effect of temperature on urediospores germination on water agar and detached host leaf. *Indian Phytopathol* 1971; 24: 759-64.
- [27] Aregbesola E, Ortega BA, Falade T, Jonathan G, Hearne S, Bandyopadhyay R. A detached leaf assay to rapidly screen for resistance of maize to *Bipolaris maydis*, the causal agent of southern corn leaf blight. *Eur J Plant Pathol* 2020; 156(1): 133-45. [http://dx.doi.org/10.1007/s10658-019-01870-4]
- [28] Jia Y, Valent B, Lee FN. Determination of host responses to *Magnaporthe oryzae* on detached rice leaves using a spot inoculation method. *Plant Dis* 2003; 87(2): 129-33. [http://dx.doi.org/10.1094/PDIS.2003.87.2.129] [PMID: 30812916]
- [29] Twizeyimana M, Ojiambo PS, Tenkouano A, Ikotun T, Bandyopadhyay R. Rapid screening of *Musa* species for resistance to black leaf streak using *in vitro* plantlets in tubes and detached leaves. *Plant Dis* 2007; 91(3): 308-14. [http://dx.doi.org/10.1094/PDIS-91-3-0308] [PMID: 30780565]
- [30] Metz SP, Chandra A, Genovesi AD, Campos CF. Comparison of screening methods to evaluate the response of St. Augustine grass to *Magnaporthe oryzae*. *Plant Pathol* 2012; 61(2): 255-61. [http://dx.doi.org/10.1111/j.1365-3059.2011.02521.x]
- [31] Herath IHMB, Stoddard FL, Marshall DR. Evaluating faba beans for rust resistance using detached leaves. *Euphytica* 2001; 117(1): 47-57. [http://dx.doi.org/10.1023/A:1004071916288]
- [32] Kohpina S, Knight R, Stoddard FL. Evaluating faba beans for resistance to ascochyta blight using detached organs. *Aust J Exp Agric* 2000; 40(5): 707-13. [http://dx.doi.org/10.1071/EA99168]
- [33] Tu JC. A detached leaf technique for screening beans (*Phaseolus vulgaris* L.) *in vitro* against anthracnose (*Colletotrichum lindemuthianum*). *Can J Plant Sci* 1986; 66(3): 805-9. [http://dx.doi.org/10.4141/cjps86-100]
- [34] El-Sherbeeney MH, Mohamed H. A Detached leaf technique for infection of faba bean plants (*Vicia faba* L.) with *Botrytis fabae*. *FABIS Newsl* 1980; 2: 44-5.
- [35] Azmat MA, Khan AA, Cheema HMN, Ashraf M, Niaz S. Detached leaf assay coupled with microscopic conidial quantification: An efficient screening method for powdery mildew resistance in pea. *Int J*

- Agric Biol 2013; 15: 957-62.
- [37] Dolar FS, Tenuta A, Higgins VJ. Detached leaf assay for screening chickpea for resistance to ascochyta blight. *Can J Plant Pathol* 1994; 16(3): 215-20. [http://dx.doi.org/10.1080/07060669409500756]
- [38] Pratt R. Screening for resistance to *Sclerotinia trifoliorum* in alfalfa by inoculation of excised leaf tissue. *Phytopathology* 1996; 86(9): 923-8. [http://dx.doi.org/10.1094/Phyto-86-923]
- [39] Schulz DF, Linde M, Bleichert O, Debener T. Evaluation of genus *Rosa* germplasm for resistance to black spot, downy mildew and powdery mildew. *Eur J Hort Sci* 2009; 74: 82000938.
- [40] Olmstead JW, Lang GA, Grove GG. A leaf disk assay for screening sweet cherry genotypes for susceptibility to powdery mildew. *HortScience* 2000; 35(2): 274-7. [http://dx.doi.org/10.21273/HORTSCI.35.2.274]
- [41] Peros JP, Nguyen TH, Troulet C, Michel RC, Notteghem JL. Assessment of powdery mildew resistance of grape and *Erysiphe necator* pathogenicity using a laboratory assay. *Vitis* 2006; 45: 29-36.
- [42] Goth RW, Keane J. A detached-leaf method to evaluate late blight resistance in potato and tomato. *Am Potato J* 1997; 74(5): 347-52. [http://dx.doi.org/10.1007/BF02851579]
- [43] Foolad MR, Ntahimpera N, Christ BJ, Lin GY. Comparison of field, greenhouse, and detached-leaflet evaluations of tomato germ plasm for early blight resistance. *Plant Dis* 2000; 84(9): 967-72. [http://dx.doi.org/10.1094/PDIS.2000.84.9.967] [PMID: 30832028]
- [44] Tedford EC, Miller TL, Nielsen MT. A detached-leaf technique for detecting resistance to *Phytophthora parasitica* var. *nicotianae* in tobacco. *Plant Dis* 1990; 74(4): 313-6. [http://dx.doi.org/10.1094/PD-74-0313]
- [45] Vleeshouwers VGAA, van Dooijewert W, Paul Keizer LC, Sijpkens L, Govers F, Colon LT. A laboratory assay for *Phytophthora infestans* resistance in various *Solanum* species reflects the field situation. *Eur J Plant Pathol* 1999; 105(3): 241-50. [http://dx.doi.org/10.1023/A:1008710700363]
- [46] Fanourakis NE. Screening procedures for powdery mildew resistance in the cucumber. *Acta Hort* 1991; (287): 147-54. [http://dx.doi.org/10.17660/ActaHortic.1991.287.14]
- [47] Sakr N. Components of quantitative resistance to downy mildew (*Plasmopara halstedii*) in sunflower (*Helianthus annuus*). *J Plant Prot Res* 2009; 49(3): 297-301. [http://dx.doi.org/10.2478/s10045-009-0048-7]
- [48] Bhattarai G, Feng C, Dhillon B, et al. Detached leaf inoculation assay for evaluating resistance to the spinach downy mildew pathogen. *Eur J Plant Pathol* 2020; 158(2): 511-20. [http://dx.doi.org/10.1007/s10658-020-02096-5]
- [49] Mhada M, Ezzahiri B, Benlhabib Q. Assessment of downy mildew resistance (*Peronospora farinosa*) in a quinoa (*Chenopodium quinoa* Willd.) germplasm. *Int J Agric Biol Eng* 2014; 8: 277-80.
- [50] Irwin JAG, Musial JM, Mackie JM, Basford KE. Utility of cotyledon and detached leaf assays for assessing root reactions of lucerne to *Phytophthora* root rot caused by *Phytophthora medicaginis*. *Australas Plant Pathol* 2003; 32(2): 263-8. [http://dx.doi.org/10.1071/AP03006]
- [51] Cowley RB, Luckett DJ, Harper JDI, Ash GJ. Development of a reliable and rapid detached leaf assay to detect resistance to the fungal disease phomopsis leaf blight, caused by *Diaporthe toxica*, in *Lupinus albus*. *Can J Plant Pathol* 2012; 34(3): 401-9. [http://dx.doi.org/10.1080/07060661.2012.705327]
- [52] Nyassé S, Cilas C, Herail C, Blaha G. Leaf inoculation as an early screening test for cocoa (*Theobroma cacao* L.) resistance to *Phytophthora* black pod disease. *Crop Prot* 1995; 14(8): 657-63. [http://dx.doi.org/10.1016/0261-2194(95)00054-2]
- [53] Subedi U, Acharya S, Chatterton S, Thomas J, Friebe D. Techniques for screening *Cercospora* leaf spot resistant fenugreek genotypes. *Can J Plant Sci* 2019; 99(3): 324-37. [http://dx.doi.org/10.1139/cjps-2018-0247]
- [54] Hansen EM, Parke JL, Sutton W. Susceptibility of Oregon forest trees and shrubs to *Phytophthora ramorum*: A comparison of artificial inoculation and natural infection. *Plant Dis* 2005; 89(1): 63-70. [http://dx.doi.org/10.1094/PD-89-0063] [PMID: 30795286]
- [55] Huang L, Li QC, Zhang Y, Li DW, Ye JR. *Colletotrichum gloeosporioides* sensu stricto is a pathogen of leaf anthracnose on evergreen spindle tree (*Euonymus japonicus*). *Plant Dis* 2016; 100(4): 672-8. [http://dx.doi.org/10.1094/PDIS-07-15-0740-RE] [PMID: 30688606]
- [56] Brooks FE. Detached-leaf bioassay for evaluating taro resistance to *Phytophthora colocasiae*. *Plant Dis* 2008; 92(1): 126-31. [http://dx.doi.org/10.1094/PDIS-92-1-0126] [PMID: 30786368]
- [57] Vawdrey LL, Martin TM, De Faveri J. A detached leaf bioassay to screen Durian cultivars for susceptibility to *Phytophthora palmivora*. *Australas Plant Pathol* 2005; 34(2): 251-3. [http://dx.doi.org/10.1071/AP05005]
- [58] Johnson R. Past, present and future opportunities in breeding for disease resistance, with examples from wheat. *Euphytica* 1992; 63(1-2): 3-22. [http://dx.doi.org/10.1007/BF00023908]
- [59] Parlevliet JE. Selecting components of partial resistance. In: Stalker HT, Murphy JP, Eds. *Plant Breeding in the 1990s*. Wallingford: CAB International 1992; Vol. 1: pp. 281-302.
- [60] Parlevliet JE. Identification and evaluation of quantitative resistance. In: Leonard KJ, Fry WE, Eds. *Plant Disease Epidemiology*. London: McGraw-Hill Publishing Company 1989; Vol. 2: pp. 215-48.
- [61] Neervoort WJ, Parlevliet JE. Partial resistance of barley to leaf rust, *Puccinia hordei*. V. Analysis of the components of partial resistance in eight barley cultivars. *Euphytica* 1978; 27(1): 33-9. [http://dx.doi.org/10.1007/BF00039117]
- [62] Birhman RK, Singh BP. Path-coefficient analyses and genetic parameters of the components of field resistance of potatoes to late blight. *Ann Appl Biol* 1995; 127(2): 353-62. [http://dx.doi.org/10.1111/j.1744-7348.1995.tb06679.x]
- [63] Simmonds NW. Genetics of horizontal resistance to diseases of crops. *Biol Rev Camb Philos Soc* 1991; 66(2): 189-241. [http://dx.doi.org/10.1111/j.1469-185X.1991.tb01140.x]
- [64] Sache I. Effect of density and age of lesions on sporulation capacity and infection efficiency in wheat leaf rust (*Puccinia recondita* f.sp. *tritici*). *Plant Pathol* 1997; 46(4): 581-9. [http://dx.doi.org/10.1046/j.1365-3059.1997.d0133.x]
- [65] Knott EA, Mundt CC. Latent period and infection efficiency of *Puccinia recondita* f.sp. *tritici* populations isolated from different wheat cultivars. *Phytopathology* 1991; 81(4): 435-9. [http://dx.doi.org/10.1094/Phyto-81-435]
- [66] Lehman JS, Shaner G. Heritability of latent period estimated from wild-type and selected populations of *Puccinia triticina*. *Phytopathology* 2007; 97(8): 1022-9. [http://dx.doi.org/10.1094/PHYTO-97-8-1022] [PMID: 18943643]
- [67] Shaner G. Probits for analyzing latent period data in studies of slow rusting resistance. *Phytopathology* 1980; 70(12): 1179-82. [http://dx.doi.org/10.1094/Phyto-70-1179]
- [68] Miller JS, Johnson DA, Hamm PB. Aggressiveness of isolates of *Phytophthora infestans* from the Columbia Basin of Washington and Oregon. *Phytopathology* 1998; 88(3): 190-7. [http://dx.doi.org/10.1094/PHYTO.1998.88.3.190] [PMID: 18944964]
- [69] Flier WG, Turkensteen LJ. Foliar aggressiveness of *Phytophthora infestans* in three potato growing regions in the Netherlands. *Eur J Plant Pathol* 1999; 105(4): 381-8. [http://dx.doi.org/10.1023/A:1008737222830]
- [70] Tomerlin JR, Eversmeyer MG, Kramer CL, Browder LE. Temperature and host effects on latent and infectious periods and on urediniospore production of *Puccinia recondita* f.sp. *tritici*. *Phytopathology* 1983; 73(3): 414-9. [http://dx.doi.org/10.1094/Phyto-73-414]
- [71] Lovell DJ, Hunter T, Powers SJ, Parker SR, Van den Bosch F. Effect of temperature on latent period of septoria leaf blotch on winter wheat under outdoor conditions. *Plant Pathol* 2004; 53(2): 170-81. [http://dx.doi.org/10.1111/j.0032-0862.2004.00983.x]
- [72] Shaw MW. Effects of temperature, leaf wetness and cultivar on the latent period of *Mycosphaerella graminicola* on winter wheat. *Plant Pathol* 1990; 39(2): 255-68. [http://dx.doi.org/10.1111/j.1365-3059.1990.tb02501.x]
- [73] Kardin MK, Groth JV. Density-dependent fitness interactions in the bean rust fungus. *Phytopathology* 1989; 79(4): 409-12. [http://dx.doi.org/10.1094/Phyto-79-409]
- [74] Rouse DI, Nelson RR, MacKenzie DR, Armitage CR. Components of rate-reducing resistance in seedlings of four wheat cultivars and parasitic fitness in six isolates of *Erysiphe graminis* f.sp. *tritici*. *Phytopathology* 1980; 70(11): 1097-100. [http://dx.doi.org/10.1094/Phyto-70-1097]
- [75] Clifford BC, Clothier RB. Physiologic specialization of *Puccinia hordei* on barley hosts with non-hypersensitive resistance. *Trans Br Mycol Soc* 1974; 63(3): 421-30. [http://dx.doi.org/10.1016/S0007-1536(74)80088-4]
- [76] Dowkiw A, Husson C, Frey P, Pinon J, Bastien C. Partial resistance to *Melampsora larici-populina* leaf rust in hybrid poplars: Genetic variability in inoculated excised leaf disk bioassay and relationship

- with complete resistance. *Phytopathology* 2003; 93(4): 421-7. [http://dx.doi.org/10.1094/PHTO.2003.93.4.421] [PMID: 18944356]
- [77] Robert C, Bancal MO, Lannou C. Wheat leaf rust uredospore production on adult plants: Influence of leaf nitrogen content and *Septoria tritici* blotch. *Phytopathology* 2004; 94(7): 712-21. [http://dx.doi.org/10.1094/PHTO.2004.94.7.712] [PMID: 18943903]
- [78] Imhoff MW, Leonard KJ, Main CE. Pattern of bean rust lesion size increase and spore production. *Phytopathology* 1982; 72(4): 441-6. [http://dx.doi.org/10.1094/Phyto-72-441]
- [79] Pringle A, Taylor JW. The fitness of filamentous fungi. *Trends Microbiol* 2002; 10(10): 474-81. [http://dx.doi.org/10.1016/S0966-842X(02)02447-2] [PMID: 12377558]
- [80] Karolewski Z, Evans N, Fitt BDL, Todd AD, Baierl A. Sporulation of *Pyrenopeziza brassicae* (light leaf spot) on oilseed rape (*Brassica napus*) leaves inoculated with ascospores or conidia at different temperatures and wetness durations. *Plant Pathol* 2002; 51(5): 654-65. [http://dx.doi.org/10.1046/j.1365-3059.2002.00746.x]
- [81] Mundt CC, Nieva LP, Vera CCM. Variation for aggressiveness within and between lineages of *Xanthomonas oryzae* pv. *oryzae*. *Plant Pathol* 2002; 51(2): 163-8. [http://dx.doi.org/10.1046/j.1365-3059.2002.00674.x]
- [82] Emge RG, Kingsolver CH, Johnson DR. Growth of the sporulating zone of *Puccinia striiformis* and its relationship to stripe rust epiphytology. *Phytopathology* 1975; 65(6): 679-81. [http://dx.doi.org/10.1094/Phyto-65-679]
- [83] Cowger C, Hoffer ME, Mundt CC. Specific adaptation by *Mycosphaerella graminicola* to a resistant wheat cultivar. *Plant Pathol* 2000; 49(4): 445-51. [http://dx.doi.org/10.1046/j.1365-3059.2000.00472.x]
- [84] Cumagun CJR, Miedaner T. Aggressiveness of 42 isolates of *Gibberella zeae* (*Fusarium graminearum*) in wheat under field and greenhouse conditions. *J Plant Dis Prot* 2003; 110: 554-9.
- [85] Roderick HW, Clifford BC. Variation in adult plant resistance to powdery mildew in spring oats under field and laboratory conditions. *Plant Pathol* 1995; 44(2): 366-73. [http://dx.doi.org/10.1111/j.1365-3059.1995.tb02789.x]
- [86] Jackson EW, Obert DE, Chong J, Avant JB, Bonman JM. Detached-leaf method for propagating *Puccinia coronata* and assessing crown rust resistance in oat. *Plant Dis* 2008; 92(10): 1400-6. [http://dx.doi.org/10.1094/PDIS-92-10-1400] [PMID: 30769569]
- [87] Singh RP, Huerta EJ. Effect of leaf rust resistance gene Lr34 on components of slow rusting at seven growth stages in wheat. *Euphytica* 2003; 129(3): 371-6. [http://dx.doi.org/10.1023/A:1022216327934]
- [88] Browne RA, Mascher F, Golebiowska G, Hofgaard IS. Components of partial disease resistance in wheat detected in a detached leaf assay inoculated with *Microdochium majus* using first, second and third expanding seedling leaves. *J Phytopathol* 2006; 154(4): 204-8. [http://dx.doi.org/10.1111/j.1439-0434.2006.01084.x]
- [89] Cadle DL. Variation within and between *Vitis* spp. for foliar resistance to the downy mildew pathogen *Plasmopara viticola*. *Plant Dis* 2008; 92(11): 1577-84. [http://dx.doi.org/10.1094/PDIS-92-11-1577] [PMID: 30764435]
- [90] Twizeyimana M, Hartman GL. Culturing *Phakopsora pachyrhizi* on detached leaves and urediniospore survival at different temperatures and relative humidities. *Plant Dis* 2010; 94(12): 1453-60. [http://dx.doi.org/10.1094/PDIS-02-10-0131] [PMID: 30743399]
- [91] Buerstmayr M, Hube K, Heckmann J. Mapping of QTL for *Fusarium* head blight resistance and morphological and developmental traits in three backcross populations derived from *Triticum dicoccum* x *Triticum durum*. *Theor Appl Genet* 2012; 125: 1751-65. [http://dx.doi.org/10.1007/s00122-012-1951-2] [PMID: 22926291]
- [92] Strange RN, Majer JR, Smith H. The isolation and identification of choline and betaine as the two major components in anthers and wheat germ that stimulate *Fusarium graminearum* *in vitro*. *Physiol Plant Pathol* 1974; 4(2): 277-90. [http://dx.doi.org/10.1016/0048-4059(74)90015-0]
- [93] Engle JS, Lippes PE, Graham TL, Boehm MJ. Effects of choline, betaine, and wheat floral extracts on growth of *Fusarium graminearum*. *Plant Dis* 2004; 88(2): 175-80. [http://dx.doi.org/10.1094/PDIS.2004.88.2.175] [PMID: 30812425]
- [94] Ding L, Xu H, Yi H, *et al.* Resistance to hemi-biotrophic *F. graminearum* infection is associated with coordinated and ordered expression of diverse defense signaling pathways. *PLoS One* 2011; 6(4): e19008. [http://dx.doi.org/10.1371/journal.pone.0019008] [PMID: 21533105]
- [95] Cho SH, Lee J, Jung KH, Lee YW, Park JC, Paek NC. Genome-wide analysis of genes induced by *Fusarium graminearum* infection in resistant and susceptible wheat cultivars. *J Plant Biol* 2012; 55(1): 64-72. [http://dx.doi.org/10.1007/s12374-011-9190-7]
- [96] Schweiger W, Steiner B, Ametz C, *et al.* Transcriptomic characterization of two major *F. usarium* resistance Quantitative Trait Loci (QTLs), *F. hbl* and *Q.fhs.ifa* 5A, identifies novel candidate genes. *Mol Plant Pathol* 2013; 14(8): 772-85. [http://dx.doi.org/10.1111/mpp.12048] [PMID: 23738863]
- [97] Ishii H, Fraaije BA, Sugiyama T, *et al.* Occurrence and molecular characterization of strobilurin resistance in cucumber powdery mildew and downy mildew. *Phytopathology* 2001; 91(12): 1166-71. [http://dx.doi.org/10.1094/PHTO.2001.91.12.1166] [PMID: 18943331]
- [98] Sakr N, Ducher M, Tourvieille J, Walser P, De Labrouhe TD. A new method to obtain monozygospore isolates of sunflower downy mildew (*Plasmopara halstedii*). *Cryptogam, Mycol* 2007; 28: 123-31.
- [99] Hofgaard IS, Ergon Å, Henriksen B, Tronsmo AM. The effect of potential resistance inducers on development of *Microdochium majus* and *Fusarium culmorum* in winter wheat. *Eur J Plant Pathol* 2010; 128(2): 269-81. [http://dx.doi.org/10.1007/s10658-010-9662-5]
- [100] Sakr N. Phenotypical characterization in "*Plasmopara halstedii*" (sunflower downy mildew) isolates of several races. *Bot Complut* 2015; 39: 7-18. [http://dx.doi.org/10.5209/rev_BOCM.2015.v39.49129]
- [101] Sakr N. *Pl2* resistance gene differentiates the pathogenicity in four "*Plasmopara halstedii*" (sunflower downy mildew) races 304, 704, 314 and 714. *Bot Complut* 2015; 39: 19-29. [http://dx.doi.org/10.5209/rev_BOCM.2015.v39.49130]
- [102] Sanju S, Thakur A, Siddappa S, *et al.* *In-vitro* detached leaf assay of host-mediated RNAi lines carrying *Phytophthora infestans AVR3a* effector gene for late blight resistance. *Potato J* 2016; 43: 30-7.
- [103] Ren R, Yang X, Ray RV. Comparative aggressiveness of *Microdochium nivale* and *M. majus* and evaluation of screening methods for *Fusarium* seedling blight resistance in wheat cultivars. *Eur J Plant Pathol* 2015; 141(2): 281-94. [http://dx.doi.org/10.1007/s10658-014-0541-3]
- [104] Sakr N. Variation in aggressiveness of *Fusarium* head blight species towards barley plants determined using three *in vitro* assays. *Pak J Phytopathol* 2019; 31(1): 19-33. [http://dx.doi.org/10.33866/phytopathol.031.01.0478]
- [105] Sakr N. Aggressiveness of *Fusarium* species causing head blight on wheat plants determined in detached leaf and seedling *in vitro* assays. *Indian Phytopathol* 2020; 73(3): 483-91. [http://dx.doi.org/10.1007/s42360-020-00234-x]
- [106] Pánek M, Strížková I. A comparison of the virulence of selected *Pythium*, *Globisporangium*, *Phytophthora* and *Phytophthora* species against strawberry plants. *J Plant Dis Prot* 2021; 128(6): 1447-58. [http://dx.doi.org/10.1007/s41348-021-00531-1]
- [107] Yayas R, Kassahun T, Clare M, Esther A, Paul G. Simple and rapid detached leaf technique for screening common beans (*Phaseolus vulgaris* L.) *in vitro* against angular leaf spot (*Pseudocercospora griseola*) disease. *Afr J Biotechnol* 2018; 17(35): 1076-81. [http://dx.doi.org/10.5897/AJB2018.16584]
- [108] Sakr N. Aggressiveness of *Fusarium* species obtained from different head parts of wheat under *in vitro* conditions. *Indian Phytopathol* 2022; 75(1): 93-9. [http://dx.doi.org/10.1007/s42360-021-00439-8]
- [109] Sakr N. *In vitro* variation in pathogenicity of *Fusarium* species causing head blight in relation to their isolation from barley head. *Acta Phytopathol Entomol Hung* 2021; 56(2): 133-42. [http://dx.doi.org/10.1556/038.2021.00118]
- [110] Sakr N, Al-Attar J. Screening for wheat resistance and pathogenicity of *Fusarium* head blight species using an *in vitro* detached head test. *Acta Phytopathol Entomol Hung* [http://dx.doi.org/10.1556/038.2022.00138]
- [111] Sedegui M, Carroll RB, Morehart AL, Hamlen RA, Power RJ. Comparison of assays for measuring sensitivity of *Phytophthora infestans* isolates to fungicides. *Plant Dis* 1999; 83(12): 1167-9. [http://dx.doi.org/10.1094/PDIS.1999.83.12.1167] [PMID: 30841144]
- [112] Chaulagain B, Dufault N, Raid RN, Rott P. Sensitivity of two sugarcane rust fungi to fungicides in urediniospore germination and detached leaf bioassays. *Crop Prot* 2019; 117: 86-93. [http://dx.doi.org/10.1016/j.cropro.2018.11.014]
- [113] Slavov S. Phytotoxins and *in vitro* screening for improved disease

- resistant plants. *Biotechnol Biotechnol Equip* 2005; 19(sup3): 48-55. [http://dx.doi.org/10.1080/13102818.2005.10817285]
- [114] Chen X, Steed A, Harden C, Nicholson P. Characterization of *Arabidopsis thaliana*?*Fusarium graminearum* interactions and identification of variation in resistance among ecotypes. *Mol Plant Pathol* 2006; 7(5): 391-403. [http://dx.doi.org/10.1111/j.1364-3703.2006.00349.x] [PMID: 20507455]
- [115] Girgi M, Breese WA, Lörz H, Oldach KH. Rust and downy mildew resistance in pearl millet (*Pennisetum glaucum*) mediated by heterologous expression of the *afp* gene from *Aspergillus giganteus*. *Transgenic Res* 2006; 15(3): 313-24. [http://dx.doi.org/10.1007/s11248-006-0001-8] [PMID: 16779647]
- [116] Gao X, Li F, Li M, *et al.* The effect of potential resistance inducers on development of *Microdochium majus* and *Fusarium culmorum* in winter wheat. *J Integr Plant Biol* 2013; 55: 586-96. [http://dx.doi.org/10.1111/jipb.12064] [PMID: 23675706]
- [117] Jabeen N, Chaudhary Z, Gulfranz M, Rashid H, Mirza B. Expression of rice chitinase gene in genetically engineered tomato confers enhanced resistance to fusarium wilt and early blight. *Plant Pathol J* 2015; 31(3): 252-8. [http://dx.doi.org/10.5423/PPJ.OA.03.2015.0026] [PMID: 26361473]
- [118] Pap D, Riaz S, Dry IB, *et al.* Identification of two novel powdery mildew resistance loci, Ren6 and Ren7, from the wild Chinese grape species *Vitis piasezkii*. *BMC Plant Biol* 2016; 16(1): 170. [http://dx.doi.org/10.1186/s12870-016-0855-8] [PMID: 27473850]
- [119] Comby M, Gacoin M, Robineau M, *et al.* Screening of wheat endophytes as biological control agents against *Fusarium* head blight using two different *in vitro* tests. *Microbiol Res* 2017; 202: 11-20. [http://dx.doi.org/10.1016/j.micres.2017.04.014] [PMID: 28647118]
- [120] Wang P, Sun Y, Pei Y, *et al.* GhSNAP33, a t-SNARE protein from *Gossypium hirsutum*, mediates resistance to *Verticillium dahliae* infection and tolerance to drought stress. *Front Plant Sci* 2018; 9: 896. [http://dx.doi.org/10.3389/fpls.2018.00896] [PMID: 30018623]
- [121] Foroud NA, Pordel R, Goyal RK, *et al.* Chemical activation of the ethylene signaling pathway promotes *Fusarium graminearum* resistance in detached wheat heads. *Phytopathology* 2019; 109(5): 796-803. [http://dx.doi.org/10.1094/PHYTO-08-18-0286-R] [PMID: 30540553]
- [122] Xue X, Geng T, Liu H, *et al.* Foliar application of silicon enhances resistance against *Phytophthora infestans* through the ET/JA- and NPR1- dependent signaling pathways in potato. *Front Plant Sci* 2021; 12: 609870. [http://dx.doi.org/10.3389/fpls.2021.609870] [PMID: 33584769]
- [123] Sakr N. Silicon reduces the severity of *Fusarium* infection on young wheat parts *in vitro*. *Open Agric J* 2022; 16(1): e187433152207260. [http://dx.doi.org/10.2174/18743315-v16-e2207260]
- [124] Sakr N. pH variation inhibits the mycelial growth of *Fusarium* Spp. and *Cochliobolus sativus*. *Pak J Phytopathol* 2021; 33(2): 265-73. [http://dx.doi.org/10.33866/phytopathol.033.02.0637]
- [125] Sakr N. Adaptation of phytopathogenic fungi to quantitative host resistance: *In vitro* selection for greater in *Fusarium* head blight species on wheat. *Cytol Genet* 2022; 56(3): 261-72. [http://dx.doi.org/10.3103/S0095452722030112]
- [126] Sakr N. Evidence for increased aggressiveness in *Fusarium* species causing head blight detected using serial passage assays through barley cultivars of contrasted quantitative resistance levels *in vitro*. *Pak J Phytopathol* 2022; 34(1): 93-104. [http://dx.doi.org/10.33866/phytopathol.034.01.0753]