



The Open Agriculture Journal

Content list available at: <https://openagriculturejournal.com>



RESEARCH ARTICLE

Productivity of Quinoa (*Chenopodium quinoa* L.) Genotypes Across Different Agro-Ecological Regions of Oman

Saif Ali AlKhamisi^{1,*}, Saleem K. Nadaf², Nadiya Mohammed Al-Jabri¹, Khalid Said Al-Hashmi³, Asma Ismail Al-Shirawi⁴, Rashad Rasool Khan⁵, Haitham Abdullah Al-Sulaimi¹ and Masoud S. Al-Azri¹

¹Plant Production Research Center, Directorate General of Agriculture and Livestock Research, Ministry of Agriculture and Fisheries, Muscat, Oman

²Oman Animal and Plant Genetic Resources Center, Ministry of Higher Education, Research and Innovation. Sultan Qaboos University Campus, Alkhod. PO Box 92; PC123. Muscat, Oman

³Department of Agriculture Research, A'Sharqiya (Alkamil), Directorate General of Agriculture and Livestock Research, Ministry of Agriculture and Fisheries, Muscat, Oman

⁴Department of Agriculture Research, North Batinah (Sohar), Directorate General of Agriculture and Livestock Research, Ministry of Agriculture and Fisheries, Muscat, Oman

⁵Plant Protection Research Center, Directorate General of Agriculture and Livestock Research, Ministry of Agriculture and Fisheries, Muscat, Oman

Abstract:

Background:

Quinoa has the great potentiality of its expansion in world agriculture owing to its resilient traits of productivity under marginal conditions across varying altitudes, soil and climatic conditions with extreme levels of abiotic factors like moisture stress, high temperature and salinity as influenced by climatic change. It offers a crop of food security in several countries that have begun the trials of evaluation since the early 1980s leading to the rapid expansion of its cultivation in other countries after the United Nation's declaration for 2013 as "Year of Quinoa" to promote its production as a grain crop. In this view, five quinoa genotypes were evaluated for their yield, agronomic performance and adaptability at three locations across different agro-ecological regions of Oman.

Methods:

Five quinoa genotypes Amarilla Marangani, Amarilla Sacaca, Blanca de junin, Kancolla and Salcedo INIA, received from the Oman FAO office in 2016, were evaluated for three consecutive years from 2016/2017 to 2018/2019 during the winter season (October to March) at national agriculture research stations located at Rumais in South Batinah governorate, AlKamil in South Sharqia governorate and Sohar in North Batinah governorate. The experiments were conducted on the sites containing sandy loam soil under Randomized Complete Block Design (RCBD) involving these five genotypes with four replications under a drip-irrigation system with varying levels of irrigation water salinity. The irrigation and fertilizers were applied as per national recommendations. The characters *viz.* chlorophyll content, plant height (cm), number of branches, inflorescence length (cm), days to maturity and grain yield (ton/ha) were recorded at appropriate times of growth of crops at all locations. Grain samples were subjected to proximate analysis. The data on yield and yield contributing characters were subjected to multi-factor analysis of variance (ANOVA) using GenStat Statistical Package v12.

Results:

The results indicated that among the effects of main factors *viz.* genotypes, locations and years, all the effects were highly significant ($p < 0.01$) in respect of plant height and grain yield, whereas for a number of branches, all the main effects except genotypes were highly significant ($p < 0.01$). In respect of chlorophyll content (SPAD value), effects of genotype and years were found significant ($p < 0.05$) and whereas for inflorescence length, effects of locations and years were highly significant ($p < 0.05$). Genotype Amarillia Sacaca had the highest plant height (135 cm), followed by genotype Salcedo INIA (117 cm) during the winter season of 2016/2017. The highest grain yield of 5.40 t ha⁻¹ obtained was harvested at Agriculture Research Station, Rumais where Amarillia Sacaca produced the highest (8.86 t ha⁻¹). The protein content was highest (17.49%) in the genotype Amarillia Marangani. Irrespective of genotypes, locations and years, the quinoa crop had acceptable performance in terms of 107.47 cm plant height, 18 number of branches, 44.47 SPDA value (chlorophyll) and 31.96 cm inflorescence length with an average yield of 3.83 t ha⁻¹.

Conclusion:

Quinoa has been found to be highly adaptable to Northern agro-ecological regions (South and North Batinah Governorates) of Oman, and genotypes evaluated can be successfully introduced for general cultivation in Northern Oman.

Keywords: *Chenopodium quinoa*, Grain yield, Chlorophyll, Proximate analysis, Agro-climatic regions, Genotypes.

1. INTRODUCTION

The quinoa (*Chenopodium quinoa* L.), once neglected or underutilized in the past, has now attained a status of quality food in the world only recently owing to its balanced mineral contents and high protein contents with quality amino acids profile, and very high antioxidants in its grains [1 - 4] and rapid expansion of its cultivation from the Andean region in South America where it was domesticated to 8 countries in 1980 to 95 countries in 2015 [5, 6]. This is due to its versatility in adaptation to all ranges of agro-ecological extremes (soils, rainfall, temperature, and altitude) [7] and tolerance to frost up to -8 °C for four hours [8], drought/moisture stress up to 200-220 mm for a cropping season [9] and salinity levels from 15 to 75 dSm⁻¹ [10], which confer high levels of genetic diversity in its varieties/ ecotypes/ accessions [11, 12]. Globally, quinoa has been distinguished as one of the strategic crops for food security and nutrition because of its resilience and high nutritional quality of its grains [13 - 15].

Quinoa is ranked high for its nutritive value as it contains more protein than other plant-foods [7]. It is chiefly cultivated in the Andean countries, where it is famous as 'the golden grain of the Andes' [7, 16 - 18]. Quinoa is light, tasty, easy to digest, and often described as nutty with a delicate taste. Its texture adds flavor to almost any recipe. In addition, it is also recognized for its nutritional and dietary properties, genetic multiplicity, adaptableness to diverse agro-environmental situations, as well as the socio-economic and cultural benefits on the indigenous environment [1, 19, 20].

Quinoa is adapted to cold environment and soil salinity; however, some current researches have revealed that it can be grown in warmer latitudes (Mediterranean region) where combinations of soil salinity and high temperature can prevail. Its ability to adjust the plant canopy and hence photosynthesis optimization makes it appropriate for cultivation in counties with adverse environmental confines, for example, in the Middle East [21]. Quinoa production worldwide has been increased from 80,069 metric tons in 2010 to 161 415 metric tons in 2019 [22]. This crop has shown tremendous potential to cope with certain global challenges *viz.*, climate change, desalinization, phytoremediation and food security [18, 23, 24]. In the marginal environments of the Middle East and North Africa (MENA) and other regions of Central Asia, quinoa offers an excellent replacement to ensure food and nutrition security to the growing population [25].

Since the declaration of the United Nations for 2013 as the "Year of Quinoa" in recognition of its significance in food security, there aroused global thrust to increase its production among several countries by way of initiating field evaluation trials for expansion of its area [1, 2, 18, 26, 27]. On similar lines, the trials were initiated on evaluating the performance of introduced varieties of quinoa in 2016 in the Sultanate of

Oman, an arid country in the Arabian Peninsula, because of its potential of expansion of cultivation in all its agriculture areas facing challenges of water stress and salinity [28]. The present investigations were to evaluate diverse quinoa genotypes for plant growth and grain yield performance in different agro-ecological locations of Oman to introduce quinoa in the present production systems of Oman.

2. MATERIALS AND METHODS

2.1. Experimental Material

Five quinoa genotypes Amarilla Marangani, Amarilla Sacaca, Blanca de junin, Kancolla and Salcedo INIA, were received from the Oman FAO office in 2016 for evaluation of their productivity in Oman. These genotypes reported to be originated in the high altitudes (2000-4000 m) of the Andean valley are characterized by a long growth period, high Saponin content and colored seed [29].

2.2. Details of Conducting Experiments

These genotypes were evaluated on three experimental sites located in different agro-ecological regions of Oman *viz.* Agriculture Research Station, Rumais in South Batinah (23°40'57.00"N, 58° 0'37.08"E), Agriculture Research Station, Alkamil in Sharqia (22°14'18.31"N, 59°10'50.17"E) and Agriculture Research Station, Wadi Hibi, Sohar, in North Batinah (24°28'15.40"N, 56°35'4.47"E) (Fig. 1).

The experiments were conducted consecutively for three years (2016/2017, 2017/2018 and 2018/2019) at layouts on sites composed of sandy loam soils during winter seasons from November to March. The electrical conductivity (EC) and pH of soil and water were recorded for each experimental site (Table 1) before the start of the experiments. The soil at Alkamil research station had the highest pH (8.1) as compared to the soil of Rumais (7.62) and Sohar (7.90) research sites. However, the EC of Sohar soil was the highest (2.5 ds m⁻¹). The irrigation water EC at Sohar ranged between 1.5 to 2.0 ds m⁻¹, whereas it was between 0.93 and 1.00 ds m⁻¹ at Alkamil and between 0.32 and 1.21 ds m⁻¹ at Rumais. Water pH ranged from 6.71 to 7.20 at Rumais and from 7.5 to 8.0 at Alkamil and Sohar (Table 2). The maximum, minimum and mean temperature conditions of the experiments at three locations *viz.* Rumais, Alkamil and Sohar during three cropping seasons from October to November are presented in Figs. (2a-2c), respectively.

The experiments were laid in Randomized Complete Block Design (RCBD) with four replications at each location. Seeds of each genotype were planted in six rows of a plot (3 × 4 m) within 2.5 cm soil depth and a planting distance of 35 cm was maintained along with 50 cm distance between the rows. The seed rate was 5 kg ha⁻¹ (53,000 seed per hectare given that the weight of seed is about 0.3 g). The fertilizers were applied at the rate of 170, 100 and 10 kg N, P and K ha⁻¹ using urea, triple superphosphate and potassium sulphate as sources, respectively. Half of nitrogen and whole of potassium and phosphorus were applied as basal doses at planting, while the

* Address correspondence to this author at the Plant Production Research Center, Directorate General of Agriculture and Livestock Research, Ministry of Agriculture and Fisheries Wealth & Water Resources, Muscat, Oman; E-mail: saifalkhamisi@hotmail.com

remaining nitrogen was applied after 30 days of emergence. The plants were irrigated for 15 minutes every day in the evening hours until germination (two weeks after planting). The irrigation time was increased by 5 minutes every 15 days until physiological maturity through a well-maintained drip irrigation system. All the crop husbandry practices were followed according to national guidelines to raise a successful crop [30].

The observations on leaf SPAD chlorophyll index were measured one month before harvest (during the flowering stage) in two locations only, whereas days to maturity were recorded whenever more than 95% maturity was attained by each genotype. Leaf chlorophyll was recorded in the field using Chlorophyll Meter SPAD-502 on three sample sites of a leaf

prior to the top leaf, which gives a value called SPAD value that corresponds to the amount of chlorophyll present in the leaf sample according to Ling *et al.* [31]. The remaining characters *viz.* plant height (cm), number of branches, inflorescence length (cm), and grain yield (ton/ha) were recorded at harvest (average of six plants) at all locations. The plants were harvested at grain maturity (after 30-35 days of flowering) by inspecting grains of five randomly sampled panicles in each plot, and panicles were threshed manually during March. In respect of proximate analysis, grain samples from each genotype were analyzed for moisture, protein, fat and fiber contents according to ISO 712:2014, ISO 1871:2015, ISO 11085:2015 and ISO 5498:2014, respectively [32] by outsourcing with Oman Flour Mill.

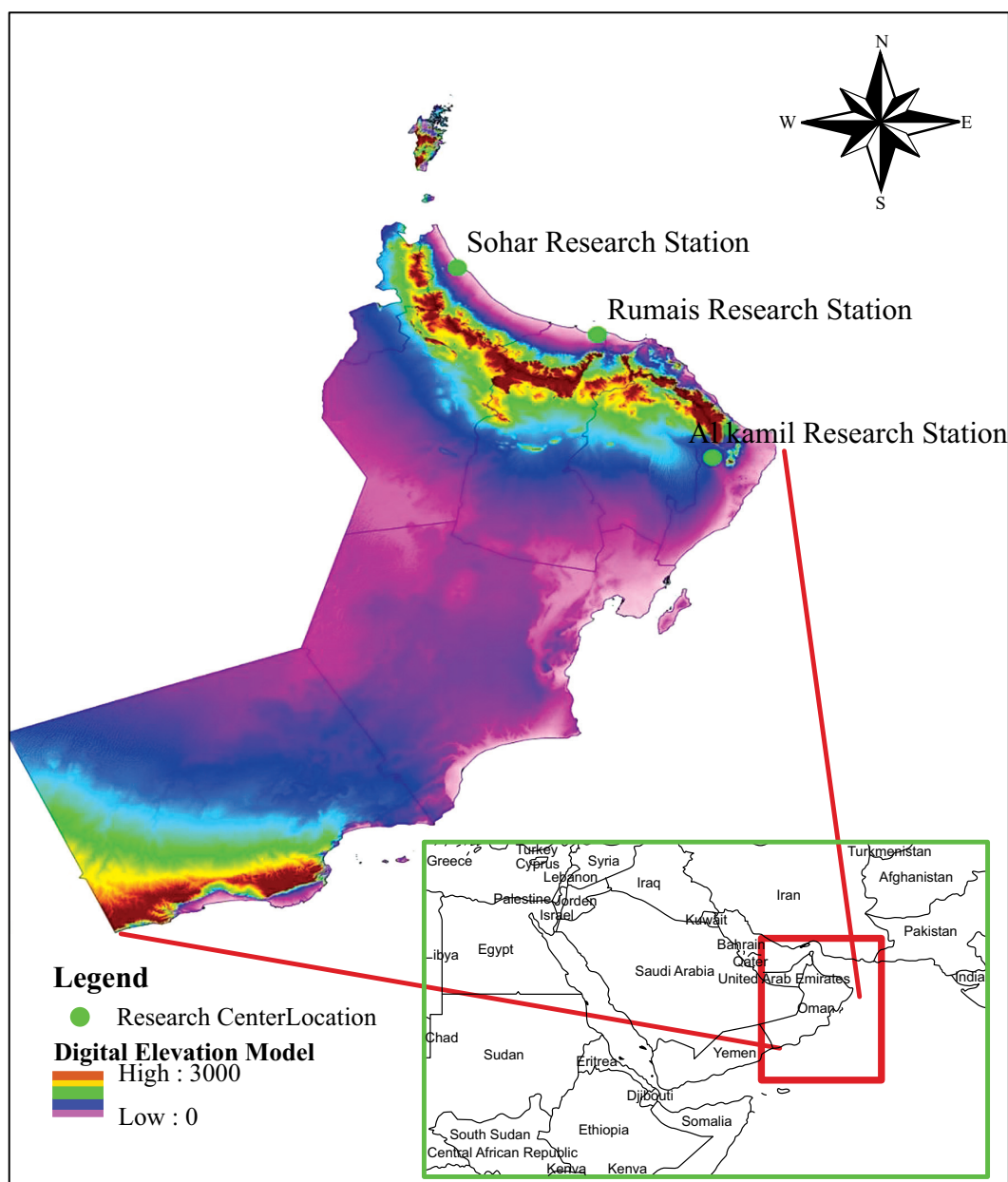


Fig. (1). Map of Oman showing three experimental locations (Rumais, Alkamil and Sohar Research Stations).

Table 1. pH and electrical conductivity (EC, dS m⁻¹) of experimental soil and water at three locations during experimentation.

Soil & Water Properties	Rumais Research Station		Alkamil Research Station		Sohar Research Station	
	Soil	Water	Soil	Water	Soil	Water
pH	7.62	6.71 to 7.20	8.1	7.5 to 8.0	7.9	7.5 to 8.0
EC (ds m ⁻¹)	1.09	0.32 to 1.21	2.2	0.93 to 1.0	2.5	1.5 to 2.0

Table 2. Means of plant height (cm) of five quinoa genotypes at three locations during winter seasons of 2016-17, 2017-18 and 2018-19.

Genotypes	Rumais Research Station			Mean of Years at Rumais	Alkamil Research Station			Mean of Years at Alkamil	Sohar Research Station			Mean of Years at Sohar	Mean of Years			Grand Mean of Years	Means of Locations			Grand Mean of Locations	Grand Mean
	2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		Rumais	Alkamil	Sohar		
Amarllia Maranagani	66.00	52.50	84.00	67.50	129.75	113.75	116.50	120.00	110.50	120.25	107.25	112.67	102.08	95.50	102.58	100.06	67.50	120.00	112.67	100.06	100.06 ^c
Amarllia Sacaca	111.00	115.75	125.50	117.42	108.00	106.25	108.25	107.50	118.25	182.75	105.25	135.42	112.42	134.92	113.00	120.11	117.42	107.50	135.42	120.11	120.11 ^a
Blanca de junio	68.75	72.25	127.25	89.42	117.75	112.00	106.25	112.00	128.25	147.00	94.25	123.17	104.92	110.42	109.25	108.19	89.42	112.00	123.17	108.19	108.19 ^b
Kancolla	63.75	75.75	98.00	79.17	106.00	104.00	101.50	103.83	117.00	130.50	92.75	113.42	95.58	103.42	97.42	98.81	79.17	103.83	113.42	98.81	98.81 ^c
Salcedo INIA	108.00	83.75	91.75	94.50	109.25	102.50	101.50	104.42	134.75	160.00	100.25	131.67	117.33	115.42	97.83	110.19	94.50	104.42	131.67	110.19	110.19 ^b
Mean of Genotypes	83.50	80.00	105.30	89.60	114.15	107.70	106.80	109.55	121.75	148.10	99.95	123.27	106.47	111.93	104.02	107.47	89.60	109.55	123.27	107.47	
				Sig (P value)				LSD at p<0.05													
Genotype				<0.001				6.91													
Location				<0.001				5.35													
Year				0.013				5.35													
Genotype × Location				<0.001				11.97													
Genotype × Year				0.002				11.97													
Location × Year				<0.001				9.27													
Genotype × Location × Year				0.003				20.74													
Coefficient of Variation (%)								3.4													

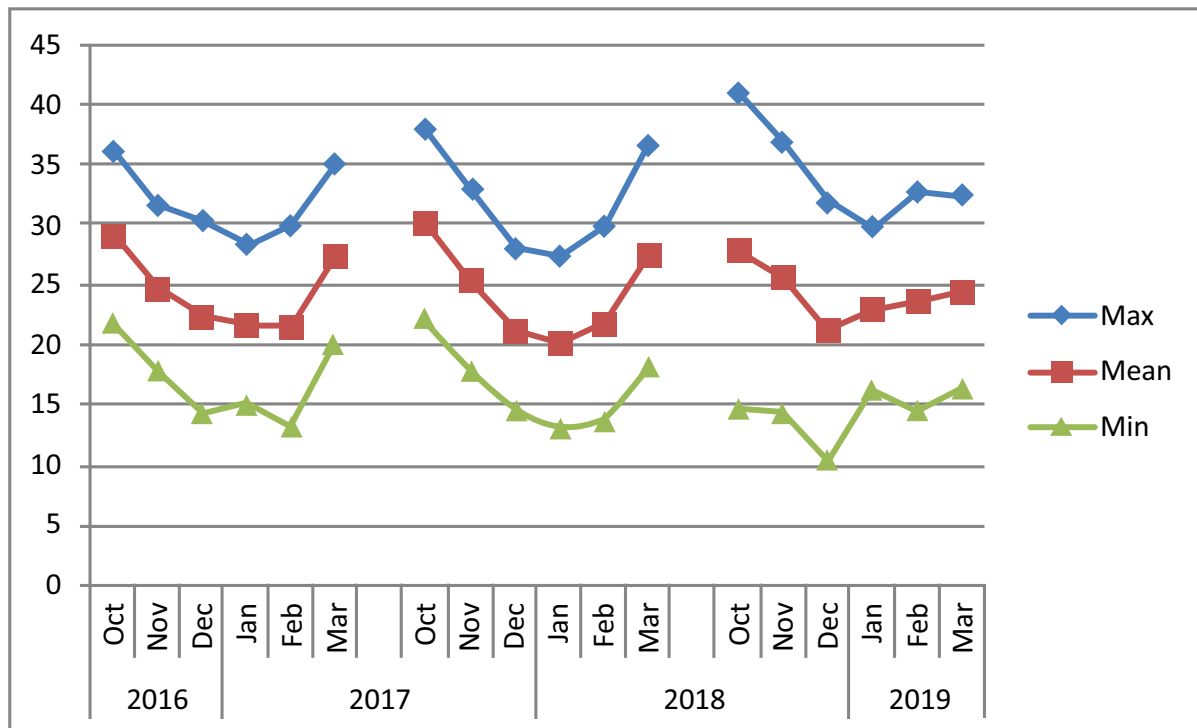


Fig. (2a). Maximum, mean and minimum temperatures in three cropping seasons during winter from October to March in 2016-2017, 2017-2018 and 2018-2019 at Rumais.

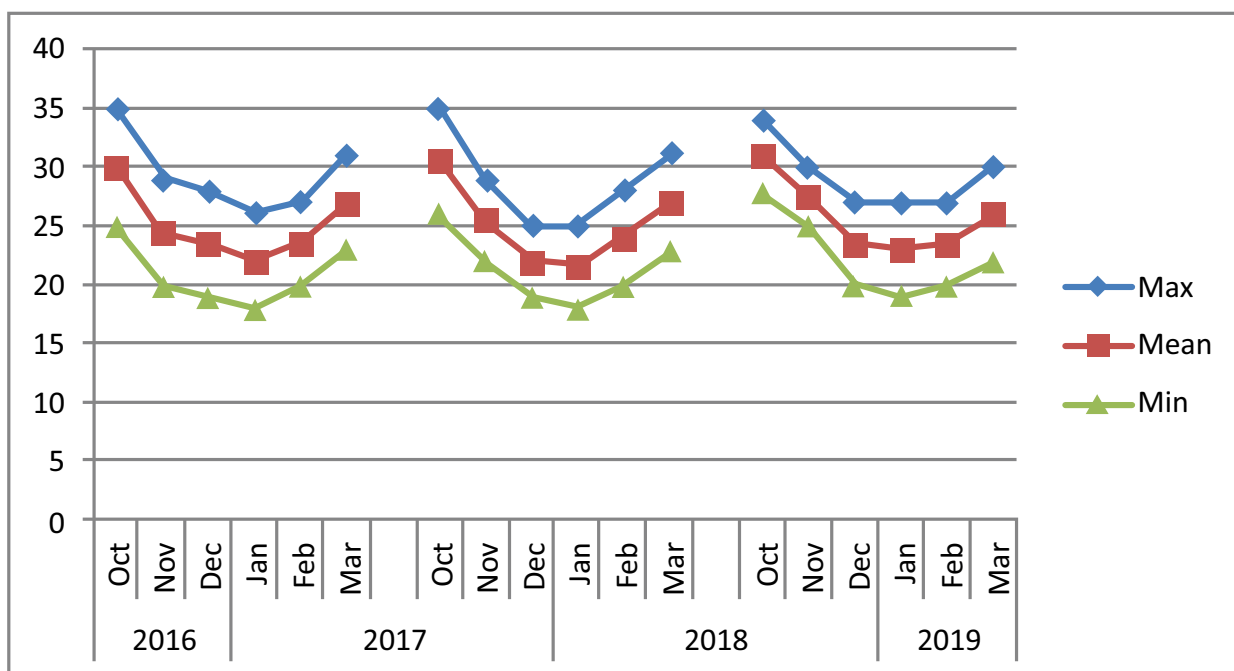


Fig. (2b). Maximum, mean and minimum temperatures in three cropping seasons during winter from October to March in 2016-2017, 2017-2018 and 2018-2019 at Alkamil.

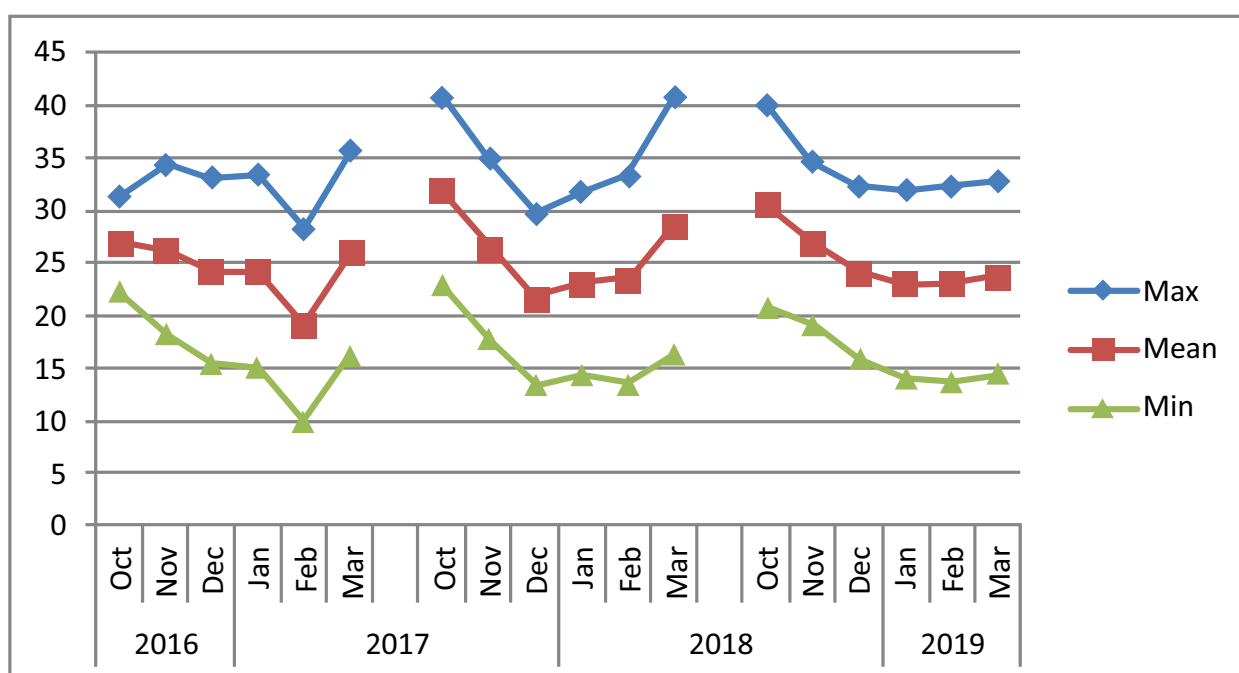


Fig. (2c). Maximum, mean and minimum temperatures in three cropping seasons during winter from October to March in 2016-2017, 2017-2018 and 2018-2019 at Wadi Hibi, Sohah.

2.3. Statistical Analysis

Data on yield and other characters were subjected to multi-factor analysis of variance (ANOVA) according to Gomez and Gomez [33], where genotype, location, and year were the factors used in the analysis using GenStat Statistical Package v12 [34].

3. RESULTS AND DISCUSSION

The results obtained from the analyzed data of the present investigations on the characters of performance of five introduced quinoa genotypes/ varieties are presented and discussed, hereunder, in light of contemporary results of the researches around the world. The results indicated that among

the effects of main factors viz. genotypes, locations and years, all the effects were highly significant ($p < 0.01$) in respect of plant height and grain yield, whereas for a number of branches, all the main effects except genotypes were highly significant ($p < 0.01$). In respect of chlorophyll content (SPAD value), effects of genotype and years were found significant ($p < 0.05$) and whereas for inflorescence length, effects of locations and years were highly significant ($p < 0.05$). However, among the interaction effects, all the effects were significant ($p < 0.05$) to highly significant ($p < 0.01$) for plant height and a number of branches. In the case of grain yield, all the interaction effects except genotypes x locations, were highly significant ($p < 0.01$). However, in respect of chlorophyll content and inflorescence length, only the effect of location and year was highly significant ($p < 0.01$). The significant effect of GxE interaction for yield was also reported earlier [7, 35, 36].

3.1. Plant Height (cm)

Uniform plant height plays a significant role in determining the quinoa suitability for mechanical harvesting [37]. Among the locations, Sohar Research Station (SRS) had the highest mean plant height (123.27 cm) in comparison with that at Rumais, which had the shortest quinoa plants (89.6 cm), whereas among the genotypes, Amarllia Sacaca genotype was the tallest (120.11 cm) followed by Salcedo INIA (110.19 cm), and Blanca de junin (108.19 cm) (Table 2). Winter season of 2017/2018 witnessed the highest mean plant height (148.10 cm) at Sohar Research Station and lowest at Rumais (80.00 cm) (Table 3). The shorter plant height at Rumais can be attributed to lower prevailing mean temperatures ranging from 20°C-29°C as compared to 22°C-31°C at Alkamil and 19°C-32°C at Sohar during the season (Fig. 2a). This is

attributed to slow enzymatic activity that causes slow and stunted plant growth [7, 37]. (Adams *et al.*, 2001; Maliro *et al.*, 2017). During the winter season of 2017/2018, Amarllia Sacaca was the highest in plant height (134.92 cm), followed by Salcedo INIA (117.33 cm) during the winter season of 2016/2017, whereas the lowest was obtained by Amarllia Maranagani (95.50 cm) during the winter season of 2017/2018 which did not significantly differ from Kancolla (95.58 cm) during the winter season of 2016/2017. (Table 3). The overall response of crop irrespective of varieties to the temperature climate in respect of plant height indicated quinoa attained the mean height of 107.49 cm with a range from 89.60 cm (Rumais) to 123.27 cm (Sohar). These observations are in line with the results of other researches across the contemporary world [39 - 42].

3.2. Number of Branches Per Plant

The results revealed highly significant differences among locations, year and their interaction ($P < 0.001$), whereas there were no significant ($p = 0.438$) differences found between genotypes in respect to a number of branches, in which all genotypes produced an average of 18 branches per plant except Amarllia Sacaca which produced highest of 20 branches (Table 4). Winter season of 2017/2018 was the highest in an average number of branches (22 branches per plant). However, the highest number of branches were produced by the plants at Rumais (26 branches per plant), which was significantly different from that produced in Sohar (16 branches per plant) and Alkamil (13 branches per plant). Irrespective of genotypes, years and locations, quinoa showed its ability to produce on an average of 18 number of branches. These observations are consistent with the results of earlier workers [41, 43 - 45].

Table 3. Means of no. of branches/ plant of five quinoa genotypes at three locations during winter seasons of 2016-17, 2017-18 and 2018-19.

Genotypes	Rumais Research Station			Mean of Years at Rumais	Alkamil Research Station			Mean of Years at Alkamil	Sohar Research Station			Mean of Years at Sohar	Mean of Years			Grand Mean of Years	Means of Locations			Grand Mean of Locations	Mean of genotypes
	2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		Rumais	Alkamil	Sohar		
Amarllia Maranagani	21	26	20	22	17	16	16	16	11	19	18	16	16	20	18	18	22	16	16	18	18
Amarllia Sacaca	19	38	34	30	12	12	12	12	11	24	15	17	14	25	20	20	30	12	17	20	20
Blanca de junin	13	30	34	26	13	13	12	13	12	20	19	17	13	21	22	18	26	13	17	18	18
Kancolla	14	34	22	23	16	14	14	15	12	21	17	17	14	23	18	18	23	15	17	18	18
Salcedo INIA	22	37	24	28	10	10	10	10	12	23	13	16	15	23	16	18	28	10	16	18	18
Mean of year x location	18	33	27	26	14	13	13	13	12	21	16	16	14	22	19	18	26	13	16	18	
					Sig (P value)				LSD at $p < 0.05$												
Genotype					0.438																
Location					<0.001				1.68												
Year					<0.001				1.68												
Genotype x Location					<0.001				3.75												
Genotype x Year					0.011				3.75												
Location x Year					<0.001				2.91												
Genotype x Location x Year					0.045				6.5												
Coefficient of Variation (%)									5.5												

Table 4. Means of chlorophyll content (SPAD value) of five quinoa genotypes at three locations during winter seasons of 2016-17, 2017-18 and 2018-19.

Genotypes	Rumais Research Station			Mean of years at Rumais	Sohar Research Station			Mean of years at Alkamil	Mean of years			Grand Mean of years	Means of locations		Grand Mean of locations	Grand mean	
	2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		Rumais	Sohar			
Amarllia Maranagani	47	47	43	45.67	52	42	37	43.67	49.50	44.50	40.00	44.67	45.67	43.67	44.67	4.04 ^a	
Amarllia Sacaca	45	51	43	46.33	56	45	39	46.67	50.50	48.00	41.00	46.50	46.33	46.67	46.50	3.95 ^a	
Blanca de junin	38	46	43	42.33	52	52	46	50.00	45.00	49.00	44.50	46.17	42.33	50.00	46.17	3.82 ^a	
Kancolla	40	46	40	42.00	52	43	35	43.33	46.00	44.50	37.50	42.67	42.00	43.33	42.67	3.34 ^b	
Salcedo INIA	40	51	33	41.33	49	41	40	43.33	44.50	46.00	36.50	42.33	41.33	43.33	42.33	3.99 ^a	
Mean of Genotypes	42.00	48.20	40.40	43.53	52.20	44.60	39.40	45.40	47.10	46.40	39.90	44.47	43.53	45.40	44.47		
				Sig (P value)				LSD (p<0.05)									
Genotype				0.026				3.3									
Location				0.085				-									
Year				<0.001				2.5									
Genotype × Location				0.076				-									
Genotype × Year				0.296				-									
Location × Year				<0.001				3.6									
Genotype × Location × Year				0.309				-									
Coefficient of Variation (%)									12.9								

3.3. Chlorophyll Content (SPAD)

In 2014, Riccardi and coworkers demonstrated the use of non-destructive evaluation of chlorophyll contents in the leaves of quinoa leaves using a chlorophyll meter that provides SPAD values which are directly proposal to chlorophyll contents [46]. The analysis of variance showed significant differences in genotypes ($p < 0.05$), and highly significant ($P < 0.001$) in the year and the interaction of year × location in respect of chlorophyll content, whereas there was the location ($p = 0.085$) and its' interaction with genotype ($p = 0.076$) was not significant (Table 4). Amarllia Sacaca and Blanca de junin genotypes, which had the highest Chlorophyll content (47 SPAD), were not significantly different, whereas Kancolla and Salcedo INIA contained the lowest Chlorophyll (43 SPAD). Quinoa chlorophyll at Sohar for 2016/2017 winter season was the highest (52 SPAD) followed by Rumais during 2017/2018 (48 SPAD), whereas the lowest was recorded at Sohar and Rumais during 2018/2019 (39 and 40 SPAD, respectively). The grand mean of SPAD over varieties, location and year, was found to be 44.47. The SPAD values reported in the present studies are in line with previous reports [8, 47, 48].

3.4. Inflorescence Length (cm)

The statistical analysis did not show any significant ($P > 0.05$) effect of genotypes and its' interaction with year and location in respect of inflorescence length. The location was significant ($p = 0.001$) in respect to inflorescence length (cm). The inflorescence length obtained in Rumais (41.87 cm) was the highest as compared to that in Sohar (32.38 cm), followed by that in Alkamil (21.63 cm), as displayed in Table 5. The

interaction between location and year was highly significant ($p < 0.001$). Inflorescence length during the winter season of 2018/2019 at Rumais was the highest (63.55 cm), followed by that obtained during 2017/2018 at Sohar (47.45 cm), whereas the lowest (14.85 cm) was obtained during the winter season of 2018/2019 at Sohar (Table 5). The grand mean of inflorescence length over genotypes, locations and years was found to be 31.96 cm. These results are similar to the reports of earlier researches that indicated inflorescence length in the range of 13.86 cm to 35.32 cm [7, 45], made by Maliro *et al.* 2017 (29.33 cm to 35.32 cm) and Biswas and Tanni, 2017 (13.86 cm to 29.62 cm) in their studies in quinoa.

3.5. Days to maturity

The results on days to maturity of five quinoa genotypes are presented in Fig. (3). Significant differences ($P < 0.05$) were found among genotypes in respect to days to maturity. Amerllia Maranagani genotypes showed the shortest period to maturity (84 days), which is significantly different from other genotypes (Fig. 3). The highest days to maturity were noted in Salcedo INIA (149 days), followed by Kancolla (124 days), Amerllia Sacaca (116 days) and Blanca de junin (103 days). The grand mean of days to maturity over the years for a location (Rumais) was found to be 115.2. These are in line with the findings of Spehar and Santos. 2005 (80-126 days) and Tan and Temel, 2018 (119- 141 days) among the genotypes studied. Belmonte *et al.* (2018) observed days to flowering ranging from 53.8 to 57.7 among the varieties investigated. It is reported that the physiological maturity of quinoa can be achieved within 70-90 days after flowering [49, 50].

Table 5. Means of Inflorescence length (cm) of five quinoa genotypes at three locations during winter seasons of 2016-17, 2017-18 and 2018-19.

Genotypes	Rumais Research Station			Mean of years at Rumais	Alkamil Research Station			Mean of years at Alkamil	Sohar Research Station			Mean of years at Sohar	Mean of years			Grand Mean of years	Means of locations			Grand Mean of locations	Grand mean
	2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		Rumais	Alkamil	Sohar		
Amarllia Maranagani	19.25	25.50	45.75	30.17	27.25	26.00	23.50	25.58	33.00	38.00	13.25	28.08	26.50	29.83	27.50	27.94	30.17	25.58	28.08	27.94	27.94
Amarllia Sacaca	31.75	37.75	49.25	39.58	21.00	21.25	19.25	20.50	33.75	56.25	14.00	34.67	28.83	38.42	27.50	31.58	39.58	20.50	34.67	31.58	31.58
Blanca de junin	35.00	29.25	47.00	37.08	25.50	24.00	22.50	24.00	34.00	48.75	17.25	33.33	31.50	34.00	28.92	31.47	37.08	24.00	33.33	31.47	31.47
Kancolla	44.00	31.50	135.75	70.42	21.50	21.50	19.25	20.75	36.50	35.25	14.50	28.75	34.00	29.42	56.50	39.97	70.42	20.75	28.75	39.97	39.97
Salcedo INIA	23.25	33.00	40.00	32.08	18.00	17.25	16.75	17.33	37.00	59.00	15.25	37.08	26.08	36.42	24.00	28.83	32.08	17.33	37.08	28.83	28.83
Mean of Genotypes	30.65	31.40	63.55	41.87	22.65	22.00	20.25	21.63	34.85	47.45	14.85	32.38	29.38	33.62	32.88	31.96	41.87	21.63	32.38	31.96	
				Sig (P value)				LSD at p<0.05													
Genotype				0.426				-													
Location				0.001				10.46													
Year				0.694				-													
Genotype × Location				0.125				-													
Genotype × Year				0.456				-													
Location × Year				<0.001				18.11													
Genotype × Location × Year				0.757				-													
Coefficient of Variation (%)										14.5											

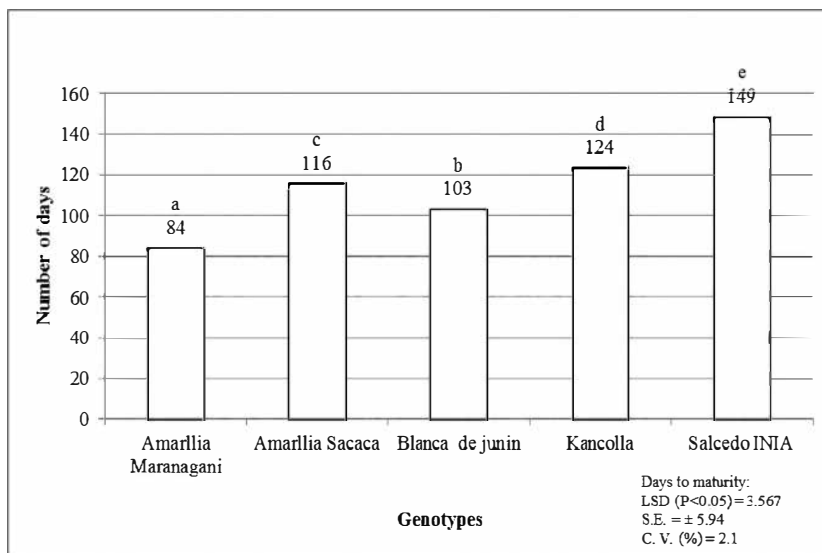


Fig. (3). Means of days to maturity for five quinoa genotypes.

3.6. Grain Yield (t ha⁻¹)

Genotype × Location × Year interaction was highly significant (p<0.001) for grain yield (t ha⁻¹). The genotypes Amarllia Maranagani, Salcedo INIA and Amarllia Sacaca produced higher grain yields of 4.04, 3.99 and 3.95 t ha⁻¹, respectively (Table 6). The grain yields obtained from these genotypes were within the range of mean grain yield (0.46 kg m⁻² equivalent to 4.6 t ha⁻¹) of five accessions evaluated at the International Center for Biosaline Agriculture (ICBA) during 2012 [38]. (Rao and Shahid, 2012). Locations had a highly significant effect (p<0.001) in which the highest grain yield was obtained at Rumais (5.40 t ha⁻¹) followed by Sohar (4.08 t ha⁻¹). Alkamil Research Station location was the lowest, with an average grain yield of 2.01 t ha⁻¹ (Table 6).

Winter season of 2017/2018 was the highest in grain yield (7.34 t ha⁻¹) at Rumais, which is significant from a season of 2018/2019 (5.95 t ha⁻¹) at the same location, whereas the lowest grain yield (1.62 t ha⁻¹) was found during winter 2018/2019 at Alkamil (Table 6). Among 25 genotypes tested in Brazilian Savannah in 2005, grain yield was recorded between 1.00 and 2.5 t ha⁻¹ [51]. Relatively, the grain yields were stable during the three seasons at Sohar (p>0.05), while they were found significantly different (p<0.05) in the other two locations, with higher preferential of Rumais and Sohar in comparison to Alkamil Station. In general, the grand mean yield of quinoa irrespective of genotypes, locations and years was found to be to the extent of 3.83 t ha⁻¹. This mean yield was similar to yield levels obtained in Egypt (3.87 t/ha) and Lebanon (4.5 t/ha) and other countries of the world [25, 52].

3.7. Proximate Analysis

The proximate analysis of quinoa genotypes are illustrated in Figs. (4-E). The moisture contents of the genotypes varied significantly between 8.75% (Salcedo INIA) to 9.48% (Amarilla Sacaca) with mean moisture of 9.17% (Fig. (4A)), which are similar to earlier findings [53]. This clearly indicates that these genotypes have about 88% and higher dry matter content indicating their higher storage ability in marketing [4, 54]. In respect of protein content, only two genotypes Amarilla Marangani (17.49%) and Salcedo INIA (17.07%), had significantly higher protein content than the remaining three genotypes (15.53% to 16.30%) (Fig. 4B). The mean protein content of the genotypes was found to be 16.5%. These figures are in line with the protein % reported by earlier workers in the range from 14 to 20% [53 - 56]. In respect of fat content, the present studies indicated an average fat content of 4.0% in

quinoa genotypes with a range from 3.57 to 4.55% (Fig. 4C). These are within the range of fat content (2% to 10%) reported earlier by Valencia-Chamorro in 2003 [49]. However, recent research reports indicated a higher range of fat contents from 5.3% to 7.8% [56 - 59]. In respect of fiber contents, the genotypes varied from 2.83 to 3.66%, with an average of 3.4% (Fig. 4D). These figures are in line with the fiber contents 4.2% reported in FAO-INFOODS Data base [59]. In the present study, the highest ash contents were found in a range of 2.87% (Amarilla Sacaca) to 3.74% (Amarilla Marangani) (Fig. 4E) with mean ash content of 3.4%. These results in the range of ash contents from 2.3% reported earlier [57, 58]. The ash content determines the amount and type of minerals in food; and has its own significance as the amount of minerals can determine the physiochemical properties of foods like quinoa [53, 60].

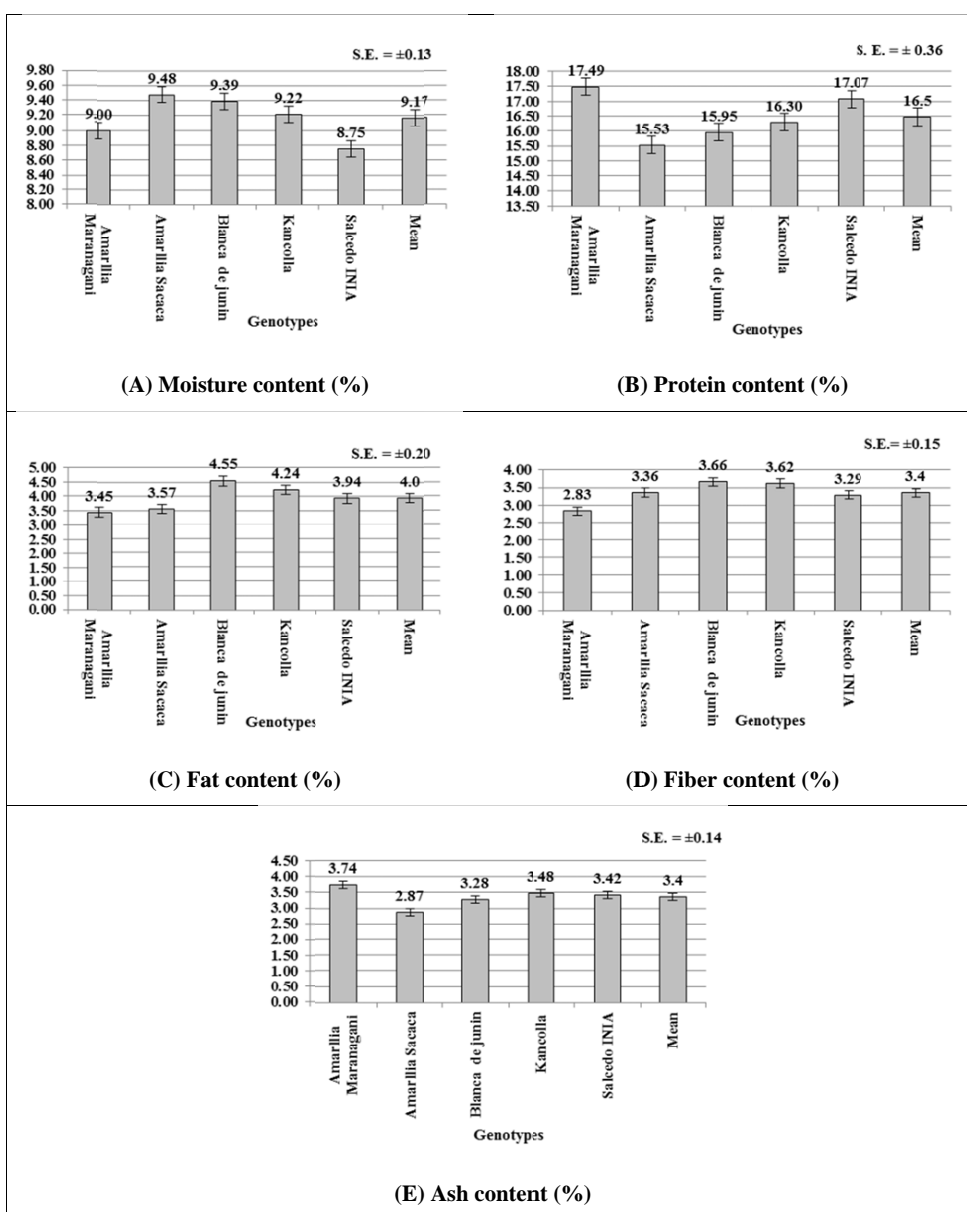


Fig. (4). Proximate analysis of five quinoa genotypes.

Table 6. Means of grain yield (t/ha) of five quinoa genotypes at three locations during winter seasons of 2016-17, 2017-18 and 2018-19.

Genotypes	Rumais Research Station			Mean of years at Rumais	Alkamil Research Station			Mean of years at Alkamil	Sohar Research Station			Mean of years at Sohar	Mean of years			Grand Mean of years	Means of locations			Grand Mean of locations	Grand mean
	2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		2016/17	2017/18	2018/19		Rumais	Alkamil	Sohar		
Amarllia Maranagani	5.47	6.08	4.93	5.49	3	2.06	1.88	2.31	4.01	4.63	4.32	4.32	4.16	4.26	3.71	4.04	5.49	2.31	4.32	4.04	4.04 ^a
Amarllia Sacaca	2.81	8.87	5.78	5.82	2.55	1.73	1.65	1.98	4.11	4	4.06	4.06	3.16	4.87	3.83	3.95	5.82	1.98	4.06	3.95	3.95 ^a
Blanca de junin	2.43	7.28	6.55	5.42	3.2	1.59	1.55	2.11	3.81	4.03	3.92	3.92	3.15	4.30	4.01	3.82	5.42	2.11	3.92	3.82	3.82 ^a
Kancolla	2.11	6.37	4.95	4.48	2.7	1.55	1.52	1.92	4.14	3.13	3.63	3.63	2.98	3.68	3.37	3.34	4.48	1.92	3.63	3.34	3.34 ^b
Salcedo INIA	1.81	8.08	7.53	5.81	2.18	1.43	1.52	1.71	4.17	4.73	4.45	4.45	2.72	4.75	4.50	3.99	5.81	1.71	4.45	3.99	3.99 ^a
Mean of Genotypes	2.93	7.34	5.95	5.40	2.73	1.67	1.62	2.01	4.05	4.10	4.08	4.08	3.23	4.37	3.88	3.83	5.40	2.01	4.08	3.83	
				Sig (P value)				LSD (p<0.05)													
Genotype				0.003				0.38													
Location				<0.001				0.30													
Year				<0.001				0.30													
Genotype × Location				0.061				-													
Genotype × Year				<0.001				0.66													
Location × Year				<0.001				0.51													
Genotype × Location × Year				<0.001				1.15													
Coefficient of Variation (%)				5.9																	

CONCLUSION

In recent years, quinoa has gained increasing interest on a global scale. The results of the present study clearly indicated that quinoa genotypes had outstanding performance reflected on mean grain productivity over seasons at two of three diverse locations located in the coastal regions, namely Rumais (5.4 t/ha) and Sohar (4.08 t/ha) with their optimum nutritional quality features in terms of protein (15.53 – 17.49%), fat (3.45 – 4.55%) and fiber (2.83 – 3.61%) contents. These results clearly revealed that quinoa genotypes tested could be conveniently introduced for general cultivation under the cropping systems existing not only in northern governorates of Oman, which are suffering from high temperature and higher soil and water salinity but also in arid areas of the Arabian Peninsula because of their adaptability to adverse agro-ecological conditions.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Not applicable.

HUMAN AND ANIMAL RIGHTS

Not applicable.

CONSENT FOR PUBLICATION

Not applicable.

AVAILABILITY OF DATA AND MATERIALS

All the experimental data are available with the corresponding author and shall be provided on request.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

FUNDING

The authors gratefully acknowledge the Directorate General of Agriculture & Livestock Research, Ministry of Agriculture, Fisheries and Water Resources of Oman for the financial support of the research

ACKNOWLEDGEMENTS

Authors are grateful to the Director General of Agriculture and Livestock Research and Director of Plant Production Research Center, for providing logistic support at the research centers. Authors also extend their acknowledgment and appreciation to the staff of Atyab FoodTech laboratory at Atyab FoodTech Trading and Services Company L.L.C for their cooperation and technical support in proximate analysis.

REFERENCES

- Jacobsen SE. The worldwide potential for quinoa (*Chenopodium quinoa* Willd.). In: Food Reviews International. 2003; pp. 167-77. [http://dx.doi.org/10.1081/FRI-120018883]
- Ruiz KB, Biondi S, Martínez EA, Orsini F, Antognoni F, Jacobsen SE. Quinoa – a model crop for understanding salt-tolerance mechanisms in halophytes. Plant Biosyst 2016; 150: 357-71. [http://dx.doi.org/10.1080/11263504.2015.1027317]
- Park JH, Lee YJ, Kim YH, Yoon KS. Antioxidant and antimicrobial activities of quinoa (*Chenopodium quinoa* Willd.) seeds cultivated in Korea. Prev Nutr Food Sci 2017; 22(3): 195-202. [http://dx.doi.org/10.3746/pnf.2017.22.3.195] [PMID: 29043217]
- Angeli V, Miguel Silva P, Crispim Massuela D, et al. Quinoa (*Chenopodium quinoa* Willd.): An overview of the potentials of the “golden grain” and socio-economic and environmental aspects of its cultivation and marketization. Foods 2020; 9(2): 216. [http://dx.doi.org/10.3390/foods9020216] [PMID: 32092899]
- Bazile D, Baudron F. “The dynamics of the global expansion of quinoa growing in view of its high biodiversity,” in State-of-the-Art Report on Quinoa Around the World in 2013. Roma: FAO & CIRAD 2015; pp. 42-55.
- Bazile D, Jacobsen SE, Verniau A. The global expansion of quinoa: Trends and limits. Front Plant Sci 2016; 7: 622. [http://dx.doi.org/10.3389/fpls.2016.00622] [PMID: 27242826]
- Maliro MFA, Guwela VF, Nyaiika J, Murphy KM. Preliminary studies of the performance of quinoa (*Chenopodium quinoa* Willd.) genotypes under irrigated and rainfed conditions of central Malawi. Front Plant

- Sci 2017; 8: 227.
[http://dx.doi.org/10.3389/fpls.2017.00227] [PMID: 28289421]
- [8] Jacobsen S-E, Monteros C, Corcuera LJ, Bravo LA, Christiansen JL, Mujica A. Frost resistance mechanisms in quinoa (*Chenopodium quinoa* Willd.). *Eur J Agron* 2007; 26: 471-5.
[http://dx.doi.org/10.1016/j.eja.2007.01.006]
- [9] Fuentes F, Bhargava A. Morphological analysis of quinoa germplasm grown under lowland desert conditions. *J Agron Crop Sci* 2011; 197: 124-34.
[http://dx.doi.org/10.1111/j.1439-037X.2010.00445.x]
- [10] Orsini F, Accorsi M, Gianquinto G, et al. Beyond the ionic and osmotic response to salinity in *Chenopodium quinoa*: functional elements of successful halophytism. *Funct Plant Biol* 2011; 38(10): 818-31.
[http://dx.doi.org/10.1071/FP11088] [PMID: 32480939]
- [11] Ruiz KB, Biondi S, Martínez E, Orsini F, Antognoni F, Jacobsen SE. Quinoa—a model crop for understanding salt-tolerance mechanisms in halophytes. *Plant Biosys Int J Deal Aspects Plant Bio* 2016; 3;150(2): 357-71.
[http://dx.doi.org/10.1080/11263504.2015.1027317]
- [12] Ruiz KB, Biondi S, Osés R, et al. Quinoa biodiversity and sustainability for food security under climate change. A review. *Agron Sustain Dev* 2015; 34: 349-59.
[http://dx.doi.org/10.1007/s13593-013-0195-0]
- [13] Rojas W, Pinto M, Flores J, Polar V. The contribution of community genebanks to in situ conservation of quinoa and cañahua: The experience of bolivia. On farm conservation of neglected and underutilized species: status, trends and novel approaches to cope with climate change: Proceedings of an International Conference. Frankfurt, Germany: Bioversity International 2011; p. 65. Available from: nuscommunity.org/fileadmin/templates/nuscommunity.org/upload/documents/Publications/2011-2014/2012_padulosi_bergamini_Lawrence_Bioversity_international.pdf#page=78
- [14] Quinoa FAO. An ancient crop to contribute to world food security. Regional Office for Latin America and Caribbean 2011; 63. Available from: <http://www.fao.org/3/aq287e/aq287e.pdf>
- [15] FAO. Dietary protein quality evaluation in human nutrition report of an FAO expert consultation 2011. Available from: <http://www.fao.org/3/a-i3124e.pdf>
- [16] Bhargava A, Shukla S, Rajan S, Ohri D. Genetic diversity for morphological and quality traits in quinoa (*Chenopodium quinoa* Willd.) germplasm. *Genet Resour Crop Evol* 2007; 54: 167-73.
[http://dx.doi.org/10.1007/s10722-005-3011-0]
- [17] Wu G, Peterson AJ, Morris CF, Murphy KM. Quinoa seed quality response to sodium chloride and sodium sulfate salinity. *Front Plant Sci* 2016; 7: 790.
[http://dx.doi.org/10.3389/fpls.2016.00790] [PMID: 27375648]
- [18] Jaikishun S, Li W, Yang Z, Song S. Quinoa: In perspective of global challenges. *Agronomy (Basel)* 2019; 2019
[http://dx.doi.org/10.3390/agronomy9040176]
- [19] FAO. Food outlook. Glob. Inf. early Warn. Syst food Agric 2015; 1-119.
[http://dx.doi.org/10.1044/leader.PPL.19102014.18]
- [20] Lutz M, Bascuñán-Godoy L. The revival of quinoa: A crop for health. In: Waisundara V, Shiomu M, Eds. Superfood and functional food- An overview and its utilization to processed foods Intechopen 2017; 37-54.
[http://dx.doi.org/10.5772/65451]
- [21] Becker VI, Goessling JW, Duarte B, et al. Combined effects of soil salinity and high temperature on photosynthesis and growth of quinoa plants (*Chenopodium quinoa*). *Funct Plant Biol* 2017; 44(7): 665-78.
[http://dx.doi.org/10.1071/FP16370] [PMID: 32480597]
- [22] Statista. Quinoa production worldwide from 2010 to 2019 (in metric tons). 2021. Available from: <https://www.statista.com/statistics/486442/global-quinoa-production>
- [23] Hariadi Y, Marandon K, Tian Y, Jacobsen SE, Shabala S. Ionic and osmotic relations in quinoa (*Chenopodium quinoa* Willd.) plants grown at various salinity levels. *J Exp Bot* 2011; 62(1): 185-93.
[http://dx.doi.org/10.1093/jxb/erq257] [PMID: 20732880]
- [24] Maughan PJ, Turner TB, Coleman CE, et al. Characterization of Salt Overly Sensitive 1 (SOS1) gene homologs in quinoa (*Chenopodium quinoa* Willd.). *Genome* 2009; 52(7): 647-57.
[http://dx.doi.org/10.1139/G09-041] [PMID: 19767895]
- [25] Choukr-Allah R, Rao NK, Hirich A, et al. Quinoa for marginal environments: towards future food and nutritional security in MENA and central Asia regions. *Front Plant Sci* 2016; 7: 346.
[http://dx.doi.org/10.3389/fpls.2016.00346] [PMID: 27066019]
- [26] Jacobsen SE, Mujica A. The genetic resources of Andean grain amaranths (*Amaranthus caudatus* L., *A. cruentus* and *A. Hypochondriacus* L.) in America. *Plant Genet Resour Newsl* 2003; 133: 41-4.
- [27] Jacobsen SE, Mujica A, Jensen CR. Resistance of quinoa (*Chenopodium quinoa* Willd.) to adverse abiotic factors. *Food Rev Int* 2003; 19(1-2): 99-109.
[http://dx.doi.org/10.1081/FRI-120018872]
- [28] MAF. Annual report of agriculture & livestock research 2018. In: Directorate General of Agriculture & Livestock Research, Ministry of Agriculture & Fisheries, Sultanate of Oman. 2018; p. 62.
- [29] Bertero D, King RW, Hall AJ. Photoperiod-sensitive development phases in Quinoa (*Chenopodium quinoa* Willd.). *Field Crops Res* 1999; 60: 231-43.
[http://dx.doi.org/10.1016/S0378-4290(98)00128-2]
- [30] Akhtar M, Nadaf SK. Scientific production of field crops in Oman. Ministry of Agriculture & Fisheries Sultanate of Oman 2001; 87.
- [31] Ling Q, Huang W, Jarvis P. Use of a SPAD-502 meter to measure leaf chlorophyll concentration in *Arabidopsis thaliana*. *Photosynth Res* 2011; 107(2): 209-14.
[http://dx.doi.org/10.1007/s11220-010-9606-0] [PMID: 21188527]
- [32] International Organization for Standardization (ISO). Occupational health and safety management systems—Requirements with guidance for use (ISO/DIS Standard No 45001) 2016. Available from: http://www.iso.org/iso/catalogue_detail?csnumber=63787
- [33] Gomez KA, Gomez AA. Statistical procedures for agricultural research. 2nd ed. New York: John Wiley and sons 1984.
- [34] International VSN. Genstat for Windows. 21st Edition. UK: VSN International, Hemel Hempstead 2020.
- [35] Spehar CR, Rocha JES. Exploiting genotypic variability from low-altitude Brazilian Savannah-adapted *Chenopodium quinoa*. *Euphytica* 2010; 175: 13-21.
[http://dx.doi.org/10.1007/s10681-010-0154-7]
- [36] Vasconcelos ES, Echer MM, Kliemann MA, Lang MJ. Selection and recommend of quinoa (*Chenopodium quinoa*) genotypes based on the yield genotypic adaptability and stability. *Rev Ceres* 2019; 66(2): 117-23.
[http://dx.doi.org/10.1590/0034-737x201966020006]
- [37] Risi J, Galwey NW. Genotype × environment interaction in the andean grain crop quinoa (*Chenopodium quinoa*) in temperate environments. *Plant Breed* 1991; 107: 141-7.
[http://dx.doi.org/10.1111/j.1439-0523.1991.tb00542.x]
- [38] Adams SR, Cockshull KE, Cave CRJ. Effect of temperature on the growth and development of tomato fruits. *Ann Bot* 2001; 88: 869-77.
[http://dx.doi.org/10.1006/anbo.2001.1524]
- [39] Rao N, Shahid M. Quinoa—a promising new crop for the arabian peninsula. *Am J Agric Environ Sci* 2002; 12: 1350-5.
[http://dx.doi.org/10.5829/idosi.ajaes.2012.12.1]
- [40] Belmonte C, Vasconcelos E, Tsutsumi C, et al. Agronomic and productivity performance for quinoa genotypes in an agroecological and conventional production system. *Am J Plant Sci* 2018; 09: 880-91.
[http://dx.doi.org/10.4236/ajps.2018.94067]
- [41] Hussain MI, Muscolo A, Ahmed M, Asghar MA, Al-Dakheel AJ. Agro-morphological, yield and quality traits and interrelationship with yield stability in quinoa (*Chenopodium quinoa* willd.) genotypes under saline marginal environment. *Plants* 2020; 9(12): 1-18.
[http://dx.doi.org/10.3390/plants9121763] [PMID: 33322139]
- [42] Shah SS, Shi L, Li Z, Ren G, Zhou B, Qin P. Yield, agronomic and forage quality traits of different quinoa (*Chenopodium quinoa* Willd.) genotypes in northeast china. *Agronomy (Basel)* 2020; 10(12): 1908.
[http://dx.doi.org/10.3390/agronomy10121908]
- [43] Hammam KA, Mansour SF. Effect of irrigation rates and organic fertilization on growth, yield and active constituents of quinoa (*Chenopodium quinoa* willd) plant. *Egypt J Agric Res* 2018; 96(4): 1473-89.
- [44] Ciftci G, Zulkadir G, Gokce MS, Karaburu E, Bozdag E, Idikut L. The effect of row distances on quinoa yield and yield components in late planting period. *Int J Res Pub and Rev* 2020; 1(4): 37-42.
- [45] Biswas BK, Tanni ZA. Quinoa (*Chenopodium quinoa* Willd.) – A potential new crop in Bangladesh: agronomic performance with sowing date. *Bangladesh Agron J* 2020; 23(1): 67-73.
[http://dx.doi.org/10.3329/baj.v23i1.50121]
- [46] Riccardi M, Mele G, Pulvento C, Lavini A, d'Andria R, Jacobsen SE. Non-destructive evaluation of chlorophyll content in quinoa and amaranth leaves by simple and multiple regression analysis of RGB image components. *Photosynth Res* 2014; 120(3): 263-72.
[http://dx.doi.org/10.1007/s11220-014-9970-2] [PMID: 24442792]

- [47] Algosaihi AM, El-Garawany MM, Badrani AL, Almadini AM. Effect of irrigation water salinity on the growth of quinoa plant seedlings. *J Agric Sci* 2015; 7(8): 205. [http://dx.doi.org/10.5539/jas.v7n8p205]
- [48] Qureshi A, Daba A. Differential analysis of five quinoa (*Chenopodium quinoa* W.) genotypes under different salt stresses in a controlled environment. *American-Eurasian J Sust Agr* 2020; 14: 14-21. [http://dx.doi.org/10.22587/aejsa.2020.14.1.2]
- [49] Valencia-Chamorro SA. Quinoa In: Caballero B, Ed. *Encyclopedia of food science and nutrition* 8: 4895-902. Available from: <http://www.sciencedirect.com/reference/239729>
- [50] Jancurová M, Minarovičová L, Dandár A. *Czech J Food Sci* 2009; 27: 71-9. [http://dx.doi.org/10.17221/32/2008-CJFS]
- [51] Spehar CR, De Barros Santos RL. Agronomic performance of quinoa selected in the Brazilian Savannah. *Pesqui Agropecu Bras* 2005; 40: 609-12. [http://dx.doi.org/10.1590/S0100-204X2005000600012]
- [52] Dost M. Field evaluation results across locations and identification of suitable quinoa varieties. Project (TcP/Rab/3403–Fao) 2015.
- [53] Chandra S, Dwivedi P, Baig MMV, Shinde LP. Importance of quinoa and amaranth in global food security in India. *J Agric Econ* 2018; 5: 26-37.
- [54] Castellión M, Matiacevich S, Buera P, Maldonado S. Protein deterioration and longevity of quinoa seeds during long-term storage. *Food Chem* 2010; 121: 952-8. [http://dx.doi.org/10.1016/j.foodchem.2010.01.025]
- [55] Matiacevich SB, Castellión ML, Maldonado SB, Buera MP. Water-dependent thermal transitions in quinoa embryos. *Thermochim Acta* 2006; 448: 117-22. [http://dx.doi.org/10.1016/j.tca.2006.06.016]
- [56] Pulvento C, Riccardi M, Lavini A, D'Andria R, Iafelice G, Marconi E. Field trial evaluation of two *Chenopodium quinoa* genotypes grown under rain-fed conditions in a typical mediterranean environment in south italy. *J Agron Crop Sci* 2010; 196: 407-11. [http://dx.doi.org/10.1111/j.1439-037X.2010.00431.x]
- [57] Ando H, Chen YC, Tang H, Shimizu M, Watanabe K, Mitsunaga T. Food components in fractions of quinoa seed. *Food Sci Technol Res* 2002; 8: 80-4. [http://dx.doi.org/10.3136/fstr.8.80]
- [58] Vidueiros SM, Curtí RN, Dyer LM, *et al.* Diversity and interrelationships in nutritional traits in cultivated quinoa (*Chenopodium quinoa* Willd.) from Northwest Argentina. *J Cereal Sci* 2015; 62: 87-93. [http://dx.doi.org/10.1016/j.jcs.2015.01.001]
- [59] FAO/INFOODS Databases. Food composition database for biodiversity version 4.0–biofood comp 4.0. Available online: <http://www.fao.org/3/a-i7364e.pdf>
- [60] Cotovanu I, Batariuc A, Mironeasa S. Characterization of quinoa seeds milling fractions and their effect on the rheological properties of wheat flour dough. *Appl Sci (Basel)* 2020; 10: 7225. [http://dx.doi.org/10.3390/app10207225]

© 2021 AIKhamisi *et al.*

This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International Public License (CC-BY 4.0), a copy of which is available at: <https://creativecommons.org/licenses/by/4.0/legalcode>. This license permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.