The Open Agriculture Journal
DOI: 10.2174/0118743315478526082406164 7 , 2026, 20, e187433154785
ISSN: 1874-3315
1

RESEARCH ARTICLE **OPEN ACCESS**

Screening of Drought-Tolerant, Silicate-Solubilizing Bacteria and Their Efficacy in Enhancing the Growth and Yield of Rice under Water Stress

Minh Hung Ho[1], Martini Mohammad Yusoff[1,*], Kee Zuan Ali Tan[2], Zulkarami Berahim[3] and Hawa ZE Jaafar[3]

[1]*Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia*
[2]*UDRP - Regenerative Agriculture for Sustainable Crop Productivity, d/a Department of Land Management, Faculty of Agriculture, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia*
[3]*Institute of Tropical Agriculture and Food Security, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia*

Abstract:

Introduction: The beneficial impacts of silicate-solubilizing bacteria (SSB) on rice growth and yield under water stress remain poorly understood. Thus, this study was conducted to screen, identify, and evaluate the efficacy of drought-tolerant silicate bacteria in improving rice growth and yield under water deficit conditions.

Methods: Ten isolates were screened for their silicate-solubilizing capacity and their drought tolerance. A two-factor pot experiment was subsequently conducted in a glasshouse to evaluate the effects of SSB on rice growth and yield under both flooded and water deficit conditions. The water deficit condition was imposed using an alternate wetting and drying technique with water applied every 10 days from 35 to 105 days after sowing. The experiment was carried out in a randomized complete block design with four replications. The pot experiment included four bacterial inoculations and two soil conditions.

Results: *In vitro* assays identified *Pseudomonas aeruginosa* and *Enterobacter sichuanensis* as the most effective strains, exhibiting the highest levels of silicate solubilization and drought tolerance. In the pot experiment, the water deficit condition effectively induced physiological stress in rice, as evidenced by a reduction in relative water content (RWC) compared with the flooded condition. Treatments with *E. sichuanensis* and co-inoculation significantly increased plant height, relative water content, root length, SPAD value, Si and lignin content in rice. Across both flooded and water-deficit conditions, the *E. sichuanensis* and co-inoculation treatments improved total grain yield by 9.25% and 12.88%, respectively, compared with the control.

Discussion: Notably, the *E. sichuanensis* and co-inoculation treatments enhanced Si uptake in rice regardless of water conditions, resulting in improved physiological attributes and yield components compared to the control.

Conclusion: These findings underscore the potential of *E. sichuanensis* and co-inoculation strategies to serve as biofertilizers to mitigate the impacts of water stress in rice and contribute to sustainable rice production. Further research is needed to validate their efficacy at the field scale and to optimize their formulation as biofertilizers.

Keywords: *Oryza sativa*, Water scarcity, Bioinoculants, PGPB, Silicon, Eco-friendly agriculture.

© 2026 The Author(s). Published by Bentham Open.
This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International Public License (CC-BY 4.0), a copy of which is available at: https://creativecommons.org/licenses/by/4.0/legalcode. This license permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

*Address correspondence to this author at the Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia; E-mail: martinimy@upm.edu.my

Cite as: Ho M, Yusoff M, Tan K, Berahim Z, Jaafar H. Screening of Drought-Tolerant, Silicate-Solubilizing Bacteria and Their Efficacy in Enhancing the Growth and Yield of Rice under Water Stress. Open Agric J, 2026; 20: e187433154785. http://dx.doi.org/10.2174/0118743315478526082406164

Received: April 23, 2026
Revised: July 01, 2026
Accepted: July 10, 2026
Published: November 04, 2026

Send Orders for Reprints to reprints@benthamscience.net

1. INTRODUCTION

Rice (*Oryza sativa* L.) is a staple food for more than half of the global population. It contributes over 75% of dietary energy intake in several developing countries [1]. Unlike other staple crops, rice is particularly susceptible to water scarcity because of its high water requirements. Flooded rice cultivation under conventional practices typically demands from 1200 to 2000 mm of water per cropping cycle [2], whereas producing 1 kg of rice grain may require up to 2500 L of water [3]. A study by Yang *et al*. highlighted that exposure to drought during the flowering stage significantly disrupts rice physiology, leading to substantial yield losses [4]. Furthermore, the increasing global population is predicted to drive high demand for rice consumption in the coming years [5], and this challenge is compounded by climate change, which is intensifying water deficiency in crop production [6], further threatening global food security. Therefore, developing a sustainable and efficient strategy for rice production under water stress is essential.

Drought is a persistent natural hazard that poses substantial risks to ecosystems, agriculture, and human society. According to Mishra and Singh, drought occurrence is strongly influenced by climatic factors, including temperature, wind speed, relative humidity, and rainfall characteristics [7]. In developing countries, where livelihoods are highly dependent on agriculture, drought events frequently result in severe socio-economic impacts, including food insecurity, displacement, and increased risks of conflict [8]. Climate change is predicted to intensify the frequency and severity of drought globally [9], thereby impairing crop production and exacerbating the potential for food crises [10].

Silicon (Si) is recognized as a beneficial element that enhances plant drought tolerance by improving morphological and physiological traits [11]. In soil, Si predominantly exists as oxides and silicates bound to other elements [12], resulting in limited soluble Si available for plant uptake [13]. To address this limitation, Si fertilizers are commonly applied to increase soil soluble Si concentration, thereby alleviating drought stress in rice plants [14]. However, Si fertilizers are often expensive and supply limited concentrations of soluble Si for plant uptake [15]. Moreover, the use of these Si-based fertilizers can have adverse effects on soil health and the ecological system [16]. Thus, the application of conventional Si-based fertilizers should be replaced with a more sustainable and environmentally friendly approach for crop production.

Plant growth-promoting bacteria (PGPB) are beneficial bacteria that enhance plant growth [17]. Several studies have shown that PGPB have the capacity to convert insoluble nutrients in the soil into available forms for plant absorption [18-21]. Among these, silicate-solubilizing bacteria (SSB) have strong potential to enhance Si availability and may serve as effective biofertilizers for crops under abiotic stress, owing to their cost-effectiveness and environmental friendliness [22]. Nevertheless, the efficacy of SSB on rice growth and yield under water stress conditions remains unclear. This study hypothesizes that inoculation with drought-resilient SSB can enhance rice performance under water stress by improving silicon uptake. Therefore, this study aimed to screen, identify, and evaluate the effects of selected SSBs on rice growth and yield under both water-deficit and flooded conditions in response to drought stress.

2. MATERIALS AND METHODS

2.1. Experimental Site and Bacterial Preparation

The *in vitro* screening experiment for PGPB was conducted at the Microbiology Laboratory in the Department of Land Management, Faculty of Agriculture, Universiti Putra Malaysia. Ten bacterial strains for this experiment were collected from the Institute of Bioscience, Universiti Putra Malaysia. These isolates are listed in Table **1**.

2.2. Silicate Solubilizing Capacity, Drought Tolerance, and Bacterial Identification on PGPB

The ability of PGPB to solubilize silicate was assessed through plate and liquid assays. The plate assay was conducted using magnesium trisilicate agar, including insoluble magnesium trisilicate (Combi-Blocks, cat. no. COMH97E3391C) 2.5 g, glucose 10 g, agar 20 g, and distilled water 1000 mL as described by Vasanthi *et al*. [23]. After 4-day incubation at 30°C ± 1, the solubilization index (SI) was calculated by using the formula (1):

SI = Colony diameter + Halo zone diameter/Colony diameter

Table 1. Ten PGPB utilized for the *in vitro* experiment.

Accession Number	Species	Source
UPMC 248	*Pseudomonas aeruginosa*	Rice plants
UPMC 382	*Bacillus* spp.	Aerobic rice
UPMC 383	*Bacillus* spp.	Aerobic rice
UPMC 385	*Bacillus* spp.	Aerobic rice
UPMC 386	*Bacillus* spp.	Aerobic rice
UPMC 1340	*Enterobacter* spp.	Soil
UPMC 1341	*Enterobacter* spp.	Soil
UPMC 1342	*Enterobacter* spp.	Soil
UPMC 1446	*Bacillus* spp.	Oil palm
UPMC 1449	*Bacillus* spp.	Rice roots

Liquid assays were carried out using a basal medium containing glucose (Millipore, cat. no. 49159, ≥ 99%) 10 g, $(NH_4)_2SO_4$ (Sigma-Aldrich, cat. no. A4915, ACS reagent, ≥ 99%) 1 g, $MgSO_4$ (Sigma-Aldrich, cat. no. 203726, ≥ 99% trace metals basis) 0.2 g, KCl (Sigma-Aldrich, cat. no. 204099, ≥ 99% trace metals basis) 0.2 g, K_2HPO_4 (Sigma-Aldrich, cat. no. P3786, ACS reagent, ≥ 98%) 0.1 g, calcium silicate (Sigma-Aldrich, cat. no. 372668, -200 mesh, ≥ 90%) 0.25 g, and distilled water 1000 mL [24]. After inoculating bacteria into the basal medium, Si concentration was recorded by Inductively Coupled Plasma Optical Emission Spectrometry (Optima 2000DV, Perkin Elmer Instruments, USA) at 5 and 10 days of incubation (DAI).

Drought-tolerant strains were determined by inoculating bacterial suspension into Tryptic Soy Broth (TSB; Millipore, cat. no. 22092) with different concentrations of polyethylene glycol 6000 (PEG-6000, Molecular Biology Grade, Sigma-Aldrich, cat. no. 528877) corresponding to osmotic potentials of 0, -0.25, -0.5, -0.75, and -1.25 MPa derived from these PEG concentrations [25]. Bacterial isolates exhibiting an increase in optical density (OD) after 48 hours of incubation were classified as drought-tolerant strains.

Selected strains were sent to Apical Scientific Sdn Bhd for bacterial identification. The bacterial full-length 16S gene (~1500 bp) was amplified using the universal primers 27F and 1492R. The total reaction volume of 25 µl contained gDNA purified using an in-house optimized protocol, 0.3 pmol of each primer, deoxynucleotide triphosphates (dNTPs, 400 µM each), 0.5 U of thermostable DNA polymerase, supplied PCR buffer, and water. The PCR was performed as follows: 1 cycle (94°C for 2 min) for initial denaturation; 25 cycles (98°C for 10 sec; 53°C for 30 sec; 68°C for 1 min) for denaturation, annealing, and extension. The PCR products were purified by the standard PCR clean-up method. The purified PCR products were subjected to bidirectional sequencing with universal primers 785F and 907R using BigDye® Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems).

2.3. Pot Experiment

Following the *in vitro* screening of PGPB, the two different strains with the best performance were selected to evaluate their efficacy on rice plants in a pot experiment conducted at Field 15, Faculty of Agriculture, Universiti Putra Malaysia. During this period, the actual measured temperature and relative humidity ranged from 29-33 °C and 79-81%, respectively. This experiment was arranged in a randomized complete block design with two factors and four replications. The first factor consisted of two water regimes: water deficit and flooded conditions. The second factor included four bacterial inoculation treatments: (i) control (no inoculation), (ii) inoculation with *Pseudomonas aeruginosa*, (iii) inoculation with *Enterobacter sichuanensis*, and (iv) co-inoculation.

The soil used in the pot experiment was collected from the Padi Transformation IR 4.0 Research Station (5.962913°N,102.287910°E) located in Kampung Padang Raja, Ketereh, Kelantan. This site is located in a major rice-producing region of Peninsular Malaysia, which is highly vulnerable to seasonal water scarcity. The region undergoes a pronounced dry period characterized by elevated temperatures [26]. Soil was collected from a paddy field at a depth of 0–30 cm in accordance with local agricultural guidelines. No formal permits were required for the collection. Subsequently, the soil was air-dried, cleaned of residues, sieved, and mixed. After that, a soil analysis was performed to determine soil properties, including pH, number of SSB, and available nutrients, including organic carbon, total nitrogen, available phosphorus, soluble Si, potassium, calcium, magnesium, and sodium. The soil properties were summarized in Table **2**.

Subsequently, 15 kg of soil was placed into individual pots (38.5 cm height x 34.5 cm diameter), which were then filled with water and left for 14 days prior to rice transplanting. The rice variety MR297 used in this study was obtained from the Department of Agriculture (DOA), Malaysia. Next, germinated seeds were sown into 200-cell trays filled with peat moss as the growth medium. Three rice seedlings were transplanted into each pot at a depth of approximately 1 cm.

Table 2. Results of soil analysis prior to planting.

Soil Properties	Analysis Results
pH	6.5
Organic carbon	0.38%
Total nitrogen	0.04%
Available phosphorus	42 ppm
Soluble Si	1.72 mg kg^{-1}
Potassium	0.05 meq/100g
Calcium	1.67 meq/100g
Magnesium	0.26 meq/100g
Sodium	0.16 meq/100g

For bacterial inoculation, cells were harvested by centrifugation at 8000 rpm, the supernatant removed, and the pellet resuspended in sterile distilled water. Seedlings were inoculated with inoculum at 15 days after sowing (DAS) by submerging their roots in bacterial solution (10^8 CFU mL^{-1}) for 30 minutes [27], while sterile distilled water was used for the control treatment. Then, the inoculated seedlings were transplanted into the pots. Re-inoculation was performed at 45 DAS by pouring the bacterial inoculum (10^8 CFU mL^{-1}) near the root zone [28].

Fertilizer application was carried out in accordance with standard agronomic guidelines established by the Malaysian Agricultural Research and Development Institute. Fertilizers were applied in splits at 7, 30, 50, and 70 days after transplanting (DAT). Fertilizers were applied at different growth stages according to the recommended application schedule. At 7 days after transplanting (DAT), 140 kg ha^{-1} NPK (17:20:10), 57 kg ha^{-1} triple superphosphate (TSP; 46% P_2O_5), and 42 kg ha^{-1} muriate of potash (MOP; 60% K_2O) were applied. At 30 DAT, 80 kg ha^{-1} urea (46% N) was applied. At 50 DAT, a combination of 12 kg ha^{-1} urea, 100 kg ha^{-1} NPK (17:20:20), and 100 kg ha^{-1} NPK (17:3:25 + 2MgO) was applied. At 70 DAT, 50 kg ha^{-1} NPK (17:3:25 + 2MgO) and 20 kg ha^{-1} urea (46% N) were applied. In addition, calcium silicate (Sigma-Aldrich, Cat. No. 372668, −200 mesh, ≥90%) was applied as a silicon source at a rate of 100 kg ha^{-1} during basal fertilizer application [22].

The water deficit condition was initiated at 35 DAS by withholding irrigation to establish an intermittent wetting-and-drying cycle [29]. Rice plants under water-deficit conditions were irrigated at 10-day intervals (35, 45, 55, 65, 75, 85, 95, and 105 DAS) with a 5 cm water level above the soil surface, allowing the soil to progressively dry between irrigation periods. The imposed water deficit condition was biologically validated by a marked reduction in RWC compared to plants maintained under continuous flooding. Rice plants under flooded conditions were continuously maintained at a 5 cm water level above the soil surface throughout the experiment.

2.4. Soil Measurement

A 1-gram soil sample was taken and transferred into a conical flask containing 99 mL of phosphate buffer, then shaken for 1 hour under laboratory conditions. Subsequently, 100 µL of 10^{-2} and 10^{-3} dilutions were inoculated onto magnesium trisilicate agar medium and incubated at 30°C ± 1 for 24–36 hours [30]. The number of SSB was determined by counting bacterial colonies with halo zones.

Si concentration in soil at 60, 90, and 120 DAS was measured using the method described by Pereira *et al.* [31]. A 10-gram soil sample was mixed with 50 mL of Na_2CO_3 (10 g/L; Sigma-Aldrich, cat. no. S7795, BioXtra, ≥ 99%) and 50 mL of NH_4NO_3 (16 g/L; Sigma-Aldrich, cat. no. 221244, ACS reagent, ≥ 98%). The mixture was shaken at 60 rpm for an hour and subsequently left to stand at room temperature for 5 days. Then, the Molybdenum Blue Colorimetric method was applied to determine the Si concentration in the sample solution [32]. A 1 mL soil solution aliquot was combined with 2.5 mL of 20% ammonium acetate (Sigma-Aldrich, cat. no. 238074, ACS reagent, ≥ 97%) and 1 mL of 0.3 M ammonium molybdate (Sigma-Aldrich, cat. no. A7302, ACS reagent, 81.0 – 83.0% MoO_3 basis) in a 50 mL microcentrifuge tube and vortexed. After standing for 5 min, 0.5 mL of 20% tartaric acid (Sigma-Aldrich, cat. no. 251380, ACS reagent grade, ≥ 99.5%), 2 mL of 20% acetic acid (Sigma-Aldrich, cat. no. 695092, glacial, ACS reagent, ≥ 99.7%), and 0.5 mL of reducing solution consisting of 2 g Na_2SO_3 (Sigma-Aldrich, cat. no. 239321, ACS reagent, ≥ 98.0%), 0.4 g $C_{10}H_9NO_4S$ (Sigma-Aldrich, cat. no. 398969, ACS reagent, ≥ 90%) 25 g $NaHSO_3$ (Sigma-Aldrich, cat. no. 243973, ACS reagent), and 250 mL H_2O were added. The solution was left at ambient temperature for 1 hour before spectrophotometric analysis at 815 nm (Thermo Scientific Multiskan GO, USA).

2.5. Growth Traits

At 90 DAS, plant height (cm) was measured from the base of the stem to the tip of the highest leaf blade. The flag leaf was subjected to the measurement of leaf area, with length (cm) recorded from tip to base at the panicle sheath and width (cm) measured at the midpoint. The RWC in the flag leaf was assessed according to the procedure of Slatyer and Barrs [33]. To measure root length, rice roots were spread in acrylic trays filled with distilled water to minimize overlap, scanned with an Epson V700, and analyzed for length using WinRHIZO root-scanning software (Regent Instruments Inc., Ottawa, ON, Canada).

2.6. Biochemical Traits

A MINOLTA™ SPAD-502 Plus (Konica Minolta Optics, Japan) was employed to measure SPAD values on the topmost fully expanded leaf at 60 and 90 DAS. In addition, lignin content in rice tissue was determined using the method described by Suzuki *et al*. [34].

The protocol of Wei-min *et al*. was used to quantify Si content in rice stems at the harvest stage [35]. Briefly, rice stems were oven-dried at 70°C to constant weight, then ground and sieved to a particle size of 0.5 mm. Next, a 0.1 g sample was added to a 50 mL Falcon tube containing 3 mL of 50% NaOH (Sigma-Aldrich, cat. no. 221465, ACS reagent, ≥ 97.0%) and autoclaved at 120°C for 20 minutes at 15 psi, after which 50 mL of distilled water was added. Subsequently, the Molybdenum Blue Colorimetric method was utilized to determine the Si concentration in the sample solution [32].

2.7. Yield Components

Yield traits included panicle numbers pot^{-1}, 1000-grain weight by weighing 1000 filled grains pot^{-1}, filled grains $panicle^{-1}$ by counting filled grains from 5 panicles pot^{-1}, filled grain ratio by dividing total filled grains by total grains, and total grain weight pot^{-1} by weighing all grains pot^{-1}.

2.8. Data Analysis

All data on rice plant development and biochemical characteristics were statistically analyzed using analysis of variance (ANOVA) in RStudio (version 2023.12.1+402; Posit, Boston, Massachusetts, USA). Treatment means were compared using the Least Significant Difference (LSD) test at a 0.05% significance level, and the tests were performed in four replications. Assumptions of normality and homogeneity of variance were verified for all datasets. For soil Si data, these assessments were conducted across all sampling stages (60, 90, and 120 DAS) to account for the inherent biological variability of rhizosphere-based experiments. Microsoft Excel was employed to generate graphical presentations.

3. RESULTS

3.1. Silicate Solubilization

No significant differences in SI were observed among UPMC 382, 383, 385, 386, 1446, and 1449. UPMC 248 (1.32) showed a higher SI than UPMC 382 and 386 (1.13-1.14) but did not differ from UPMC 383, 385, 1446, and 1449 (1.21-1.24). However, the highest SI values were recorded in UPMC 1340, 1341, and 1342 (1.80-1.86) (Table **3**). In the liquid assay at 5 DAI, UPMC 248, 386, 1340, 1341, 1342, 1446, and 1449 (71.39-75.29 mg L^{-1}) showed significantly higher silicon solubilization than the control (68.93 mg L-1). In contrast, all bacterial isolates (80.34-91.04 mg L^{-1}) had significantly greater Si concentration than the control (78.71 mg L^{-1}) at 10 DAI, except for UPMC 382 (80.28 mg L^{-1}).

3.2. *In vitro* Drought Evaluation

Addition of PEG-6000 to the TSB medium inhibited the growth and survival of bacterial strains (Fig. **1**). All bacteria grew well at -0.25 MPa, whereas growth was inhibited at -0.5 MPa, reducing the number of isolates to seven. At lower water potential (-0.75 and -1.25 MPa), only six isolates (UPMC 248, 386, 383, 1340, 1341, and 1342) remained viable.

3.3. Bacterial Identification based on Full-length 16S rDNA Gene Sequence

Phylogenetic analysis using sequences retrieved from the National Center for Biotechnology Information (NCBI) revealed that approximately 1473 bp were sequenced for UPMC 1340, UPMC 1341, and UPMC 1342 (Fig. **2**). The gene sequence results were searched on the Basic Local Alignment Search Tool, which demonstrated 99.73% sequence similarity between these isolates and *Enterobacter sichuanensis* (accession number NR_179946.1). To avoid potential overlap in effects, two distinct strains (UPMC 248 and 1341) were chosen for the pot experiment.

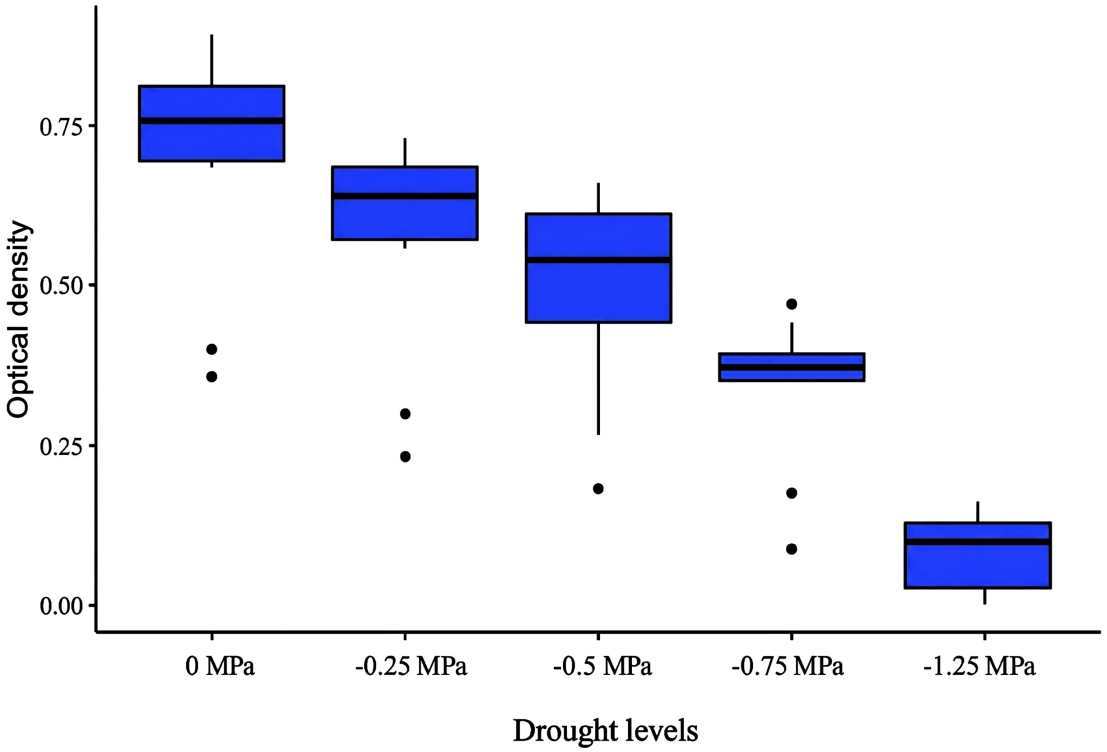

Fig. (1). Optical density of ten isolates on in-vitro drought treatment at five different water potentials. Box plot showing the optical density (OD600) of ten silicate-solubilizing bacteria after 48 h incubation under five drought stress levels such as 0, -0.25, -0.5, -0.75, and -1.25 MPa. The boxes indicate the interquartile range, the horizonal line denotes the median, whisk extend to the minimum and maximum values, and dots represent outliers.

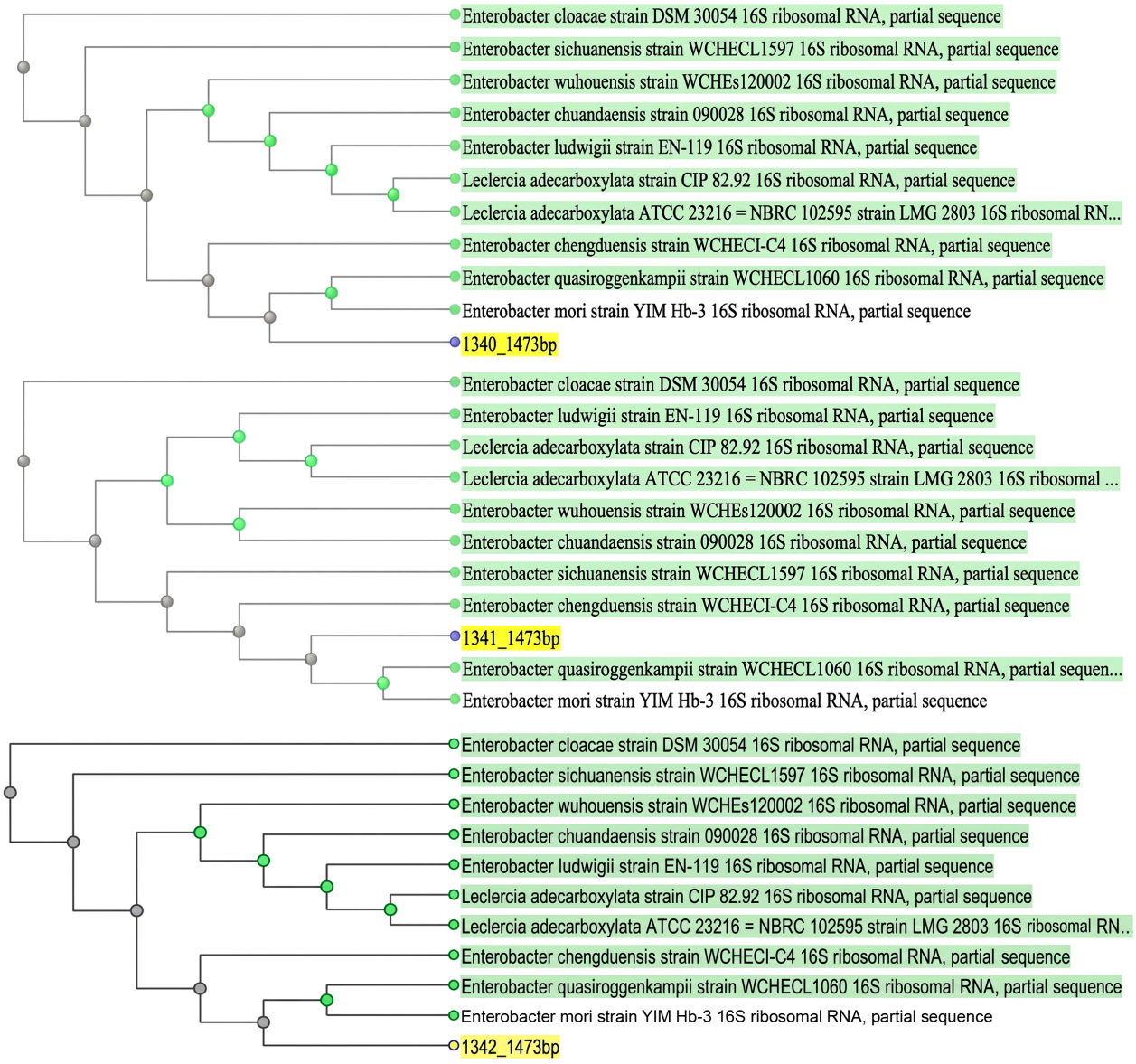

Fig. (2). Phylogenetic tree based on 16S rRNA gene sequences indicating the relationship of three isolates UPMC 1340, UPMC 1341, and UPMC 1342 to closely related isolates. Bar denotes the nucleotide divergence value.

Table 3. Solubilization index and Si concentration on plate and liquid assays.

Treatment	Solubilization Index (SI)	Si Concentration (mg L^{-1})	
		5 DAI	10 DAI
Control	-	68.93^{d} (67.68, 70.19)	78.71^{d} (77.58, 79.83)
UPMC 248	1.32^{b} (1.22, 1.42)	72.55^{b} (71.29, 73.80)	82.20^{b} (81.07, 83.32)
UPMC 382	1.13^{c} (1.03, 1.23)	69.02^{d} (67.76, 70.27)	80.28^{cd} (79.16, 81.41)

(Table 3) contd.....

Treatment	Solubilization Index (SI)	Si Concentration (mg L^{-1})	
		5 DAI	10 DAI
UPMC 383	1.22^{bc} (1.12, 1.32)	70.06^{cd} (68.81, 71.32)	81.42^{bc} (80.30, 82.55)
UPMC 385	1.21^{bc} (1.11, 1.31)	69.31^{d} (68.06, 70.57)	80.34^{c} (79.22, 81.46)
UPMC 386	1.14^{c} (1.04, 1.24)	71.78^{bc} (70.52, 73.03)	81.48^{bc} (80.35, 82.60)
UPMC 1340	1.80^{a} (1.70, 1.90)	72.93^{b} (71.68, 74. 19)	90.77^{a} (89.65, 91.90)
UPMC 1341	1.86^{a} (1.76, 1.96)	75.07^{a} (73.82, 76.33)	91.04^{a} (89.92, 92.17)
UPMC 1342	1.83^{a} (1.73, 1.93)	75.29^{a} (74.04, 76.55)	90.46^{a} (89.34, 91.58)
UPMC 1446	1.22^{bc} (1.12, 1.32)	71.39^{bc} (70.14, 72.65)	80.55^{c} (79.43, 81.67)
UPMC 1449	1.24^{bc} (1.14, 1.34)	71.60^{bc} (70.35, 72.86)	81.18^{bc} (80.06, 82.31)
F	40.35***	13.77***	77.92***
CV (%)	5.86	1.40	1.08

Note: Means within column followed by the same letter are not significantly different by least significant difference, $p \geq 0.05$; * = Significant at $p \leq 0.05$; ** = Significant at $p \leq 0.01$; *** = Significant at $p \leq 0.001$; ns = Not significant. Values in parentheses represent the 95% confidence intervals.

3.4. Soil Measurement

Table **4** showed no significant interaction in the number of SSB in soil between bacterial inoculations and water conditions at any of the examined stages in rice plants. The number of SSB in the *E. sichuanensis* and co-inoculation treatment was significantly higher than the control at 60 DAS, whereas only the co-inoculation showed a higher density of SSB than the control at 90 DAS. However, the number of SSB across all treatments did not differ significantly at 120 DAS. Compared with flooded conditions, the number of SSB under water-deficit conditions was significantly lower at 60 and 90 DAS. In contrast, water deficit conditions (3.93 $\log_{10}$CFU g^{-1} soil) showed a significantly higher number of SSB than flooded conditions (3.58 $\log_{10}$CFU g^{-1} soil) at 120 DAS.

Table 4. Number of silicate-solubilizing bacteria of rice plants treated with SSB under different water conditions.

Treatment	DF	Number of SSB in soil ($\log_{10}$CFU g^{-1} soil)			
		0 DAS	60 DAS	90 DAS	120 DAS
Types of bacteria inoculation (A)		-	-	-	-
Control	-	3.30	3.69^{c} (3.52, 3.87)	3.61^{b} (3.37, 3.85)	3.60^{a} (3.25, 3.94)
P. aeruginosa	-	3.30	3.93^{bc} (3.75, 4.11)	3.86^{ab} (3.62, 4.10)	3.74^{a} (3.39, 4.09)
E. sichuanensis	-	3.30	4.03^{b} (3.86, 4.21)	3.71^{b} (3.47, 4.00)	3.76^{a} (3.41, 4.11)
Co-inoculation	-	3.30	4.36^{a} (4.19, 4.54)	4.10^{a} (3.85, 4.34)	3.91^{a} (3.56, 4.26)
Water conditions (B)	-	-	-	-	-
Water deficit	-	3.30	3.78^{b} (3.66, 3.91)	3.69^{b} (3.51, 3.86)	3.93^{a} (3.68, 4.17)
Flooded	-	3.30	4.23^{a} (4.10, 4.36)	3.95^{a} (3.78, 4.13)	3.58^{b} (3.33, 3.82)
F (A)	3	ns	10.43***	3.28*	0.59^{ns}
F (B)	1	ns	26.93***	5.26*	4.41*
F (AxB)	3	ns	0.21^{ns}	1.22^{ns}	0.21^{ns}
CV (%)	-	0.00	6.12	8.68	12.51

Note: Means within column followed by the same letter are not significantly different by least significant difference, $p \geq 0.05$; * = Significant at $p \leq 0.05$; ** = Significant at $p \leq 0.01$; *** = Significant at $p \leq 0.001$; ns = Not significant. Values in parentheses represent the 95% confidence intervals.

Soluble Si in soil showed no interaction effects between bacterial inoculations and water conditions in any sampling period (Table 5). The co-inoculation treatment showed significantly higher soluble Si levels than the control at all examined stages. Compared to the control, the *P. aeruginosa* treatment significantly increased soluble Si at 60 and 120 DAS, but the *E. sichuanensis* treatment showed no significant effects at any sampling time. Regarding water conditions, soluble Si in the flooded condition (7.83 mg kg-1) was significantly higher than in the water-deficit condition (4.42 mg kg^{-1}) at 60 DAS. Nevertheless, no significant difference in soluble Si concentration was recorded in the water conditions at 90 and 120 DAS.

3.5. Growth and Biochemical Attributes

The interactions between two factors were not significant in plant height, leaf area, SPAD values, lignin, and Si content, except for RWC and root length (Tables 6 and 7). Compared to the control, the bacterial treatments improved plant height, leaf area, RWC, root length, SPAD values, lignin, and Si content. The plant height in the co-inoculation treatment was comparable to that of the *E. sichuanensis* treatment and greater than that of the *P. aeruginosa* treatment. For the leaf area, there were no differences among the bacterial treatments. However, the greatest RWC was observed in the co-inoculation treatment, followed by the *E. sichuanensis* and *P. aeruginosa* treatments. The root length in the co-inoculation treatment was significantly higher than that of the *P. aeruginosa* treatment, but not different from the *E. sichuanensis* treatment. Additionally, SPAD values in the co-inoculation treatment were equivalent to those in the single *E. sichuanensis* treatment and greater than those in the control. The SSB treatments increased lignin content by 3.5-9.03% and Si content by 22.82-42.72% relative to the control. Water-deficit conditions significantly reduced plant height, leaf area, RWC, root length, and SPAD values. On the contrary, lignin content in water-deficit conditions did not differ from that in flooded conditions, whereas water-deficit conditions ameliorated Si content compared to flooded conditions.

Table 5. Soluble Si concentration in the soil of rice plants treated with SSB under different water conditions.

Treatment	DF	Soluble Si concentration in soil (mg kg^{-1})			
		0 DAS	60 DAS	90 DAS	120 DAS
Types of bacteria inoculation (A)		-	-	-	-
Control		1.72	3.13^{b} (0.65, 5.62)	1.01^{b} (-0.14, 2.16)	1.75^{c} (-0.41, 3.92)
P. aeruginosa		1.72	7.28^{a} (4.79, 9.76)	2.29^{ab} (1.13, 3.44)	5.70^{ab} (3.54, 7.86)
E. sichuanensis		1.72	5.46^{ab} (2.97, 7.94)	2.00^{ab} (0.84, 3.15)	3.45^{bc} (1.29, 5.62)
Co-inoculation		1.72	8.64^{a} (6.15, 11.13)	3.50^{a} (2.35, 4.65)	6.62^{a} (4.46, 8.79)
Water conditions (B)		-	-	-	-
Water deficit		1.72	4.42^{b} (2.66, 6.18)	2.00^{a} (1.18, 2.81)	5.11^{a} (3.58, 6.64)
Flooded		1.72	7.83^{a} (6.07, 9.59)	2.40^{a} (1.58, 3.21)	3.66^{a} (2.13, 5.19)
F (A)	3	ns	3.97*	3.41*	4.48*
F (B)	1	ns	8.10**	0.53^{ns}	1.94^{ns}
F (AxB)	3	ns	0.53^{ns}	1.17^{ns}	0.34^{ns}
CV (%)		0.00	55.24	71.39	67.19

Note: Means within column followed by the same letter are not significantly different by least significant difference, $p \geq 0.05$; * = Significant at $p \leq 0.05$; ** = Significant at $p \leq 0.01$; *** = Significant at $p \leq 0.001$; ns = Not significant. Values in parentheses represent the 95% confidence intervals.

Table 6. Plant height, leaf area, RWC, and root length on rice plants treated with SSB under different water conditions.

Treatment	DF	Plant height (cm)	Leaf area (cm^2)	RWC (%)	Root length (m)
Types of bacteria inoculation (A)		-	-		
Control	-	38.27^{c} (37.53, 39.00)	28.81^{b} (26.90, 30.72)	41.16^{d} (38.57, 43.74)	113.35^{c} (103.51, 123.20)
P. aeruginosa	-	38.95^{bc} (38.22, 39.68)	30.30^{ab} (28.39, 32.21)	46.84^{c} (44.26, 49.42)	136.02^{b} (126.17, 145.86)
E. sichuanensis	-	39.35^{ab} (38.62, 40.08)	3 1.33^{ab} (29.42, 33.24)	55.24^{b} (52.66, 57.82)	147.34^{ab} (137.49, 157.18)
Co-inoculation	-	40.06^{a} (39.33, 40.80)	32.74^{a} (30.83, 34.65)	60.17^{a} (57.58, 62.75)	159.32^{a} (149.48, 169.16)
Water conditions (B)	-	-	-	-	-
Water deficit	-	36.06^{b} (35.54, 36.58)	27.66^{b} (26.31, 29.01)	45.82^{b} (43.99, 47.64)	111.22^{b} (104.26, 118.18)
Flooded	-	42.25^{a} (41.73, 42.77)	33.93^{a} (32.58, 35.28)	55.88^{a} (54.05, 57.70)	166.79^{a} (159.83, 173.75)
F (A)	3	4.53*	3.26*	46.78***	17.09***

(Table 6) contd.....

Treatment	DF	Plant height (cm)	Leaf area (cm^2)	RWC (%)	Root length (m)
F (B)	1	307.60***	46.71***	65.70***	137.81***
F (AxB)	3	0.51^{ns}	0.25^{ns}	3.50*	6.78**
CV (%)	-	2.55	8.43	6.90	9.63

Note: Means within column followed by the same letter are not significantly different by least significant difference, $p \geq 0.05$; * = Significant at $p \leq 0.05$; ** = Significant at $p \leq 0.01$; *** = Significant at $p \leq 0.001$; ns = Not significant. Values in parentheses represent the 95% confidence intervals.

Table 7. SPAD values, lignin content, and Si content on rice plants treated with SSB under different water conditions.

Treatment	DF	SPAD Values	Lignin content (mg g^{-1})	Si content (mg g^{-1})
Types of bacteria inoculation (A)	-	-	-	-
Control	-	46.15^{c} (45.37, 46.94)	17.83^{c} (17.41, 18.24)	2.06^{c} (1.87, 2.26)
P. aeruginosa	-	47.02^{bc} (46.24, 47.81)	18.47^{b} (18.05, 18.89)	2.53^{b} (2.33, 2.72)
E. sichuanensis	-	47.41^{ab} (46.62, 48.19)	19.05^{ab} (18.63, 19.46)	2.57^{b} (2.37, 2.76)
Co-inoculation	-	48.18^{a} (47.40, 48.97)	19.44^{a} (19.03, 19.86)	2.94^{a} (2.74, 3.13)
Water conditions (B)	-	-	-	-
Water deficit	-	46.38^{b} (45.82, 46.93)	18.85^{a} (18.56, 19.15)	2.88^{a} (2.74, 3.02)
Flooded	-	48.01^{a} (47.45, 48.56)	18.54^{a} (18.24, 18.83)	2.17^{b} (2.03, 2.31)
F (A)	3	5.00**	12.35***	14.65***
F (B)	1	18.74***	2.49^{ns}	57.32***
F (AxB)	3	0.05^{ns}	1.63^{ns}	0.75^{ns}
CV (%)	-	2.26	3.03	10.50

Note: Means within column followed by the same letter are not significantly different by least significant difference, $p \geq 0.05$; * = Significant at $p \leq 0.05$; ** = Significant at $p \leq 0.01$; *** = Significant at $p \leq 0.001$; ns = Not significant. Values in parentheses represent the 95% confidence intervals.

In addition, the interaction between the two factors significantly affected RWC and root length. Significant differences in RWC between flooded and water deficit conditions were recorded in the single inoculations, but the co-inoculation treatment increased that of water deficit conditions, resulting in non-significant differences between flooded and water deficit conditions (Fig. **3**). Rice plants under flooded conditions exhibited significantly longer roots compared to those under water deficit conditions across all SSB treatments (Fig. **4**).

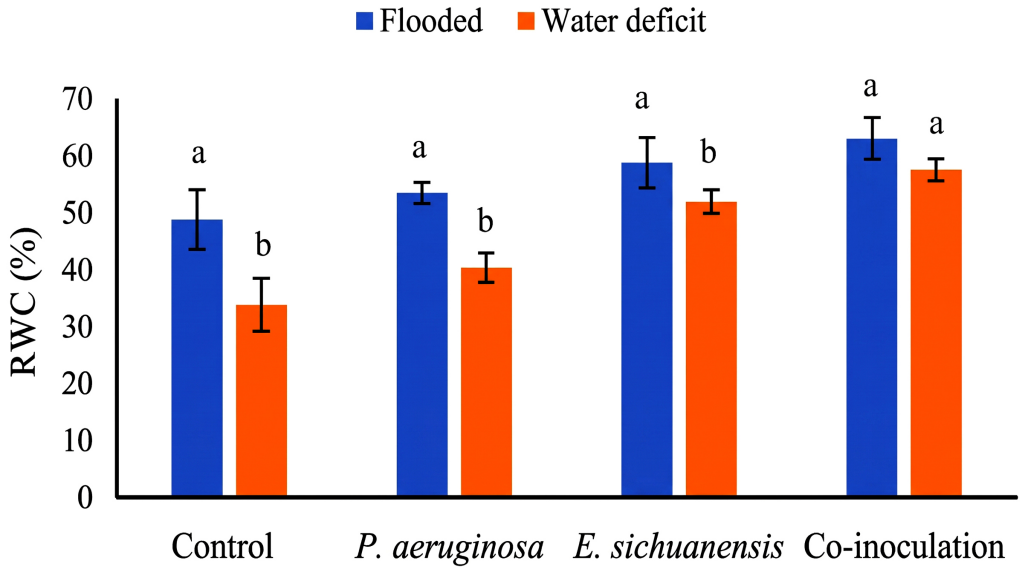

Fig. (3). Effect of bacterial inoculation on relative water content of rice plants. Relative water content (RWC) of rice plants under flooded and water deficit conditions with four bacterial inoculations (control, *P. aeruginosa*, *E. sichuanensis*, and co-inoculation). Error bars indicate the standard error of the mean (n=4). Columns marked with different letters differ significantly at the 5% probability level.

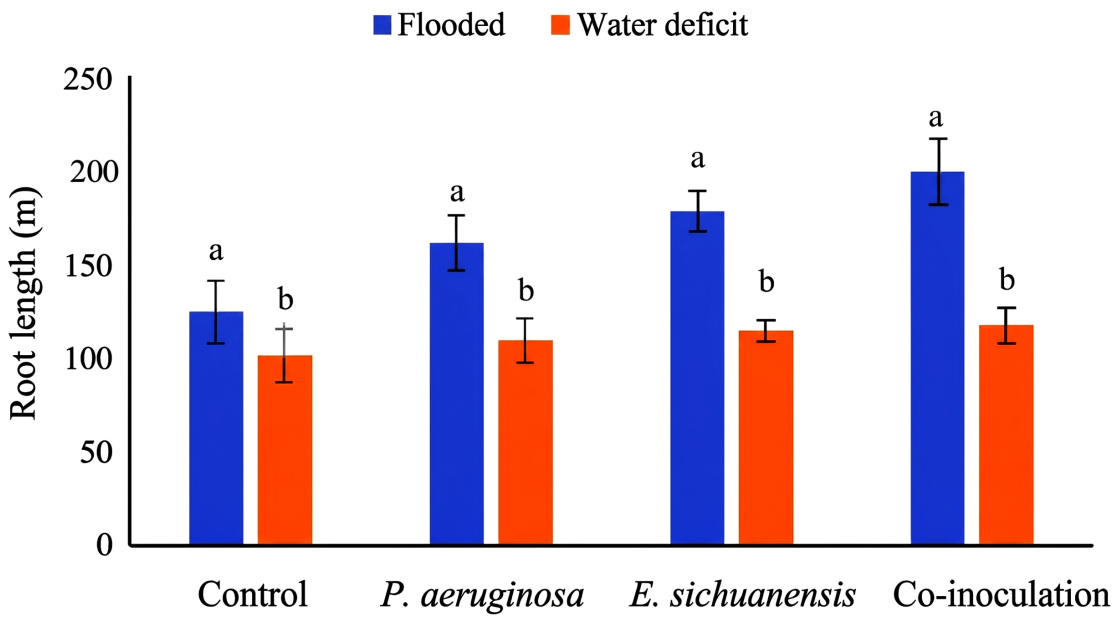

Fig. (4). Effect of bacterial inoculation on root length of rice plants. Root length of rice plants under flooded and water deficit conditions with four bacterial inoculations (control, *P. aeruginosa*, *E. sichuanesis*, and co-inoculation). Error bars indicate the standard error of the mean (n=4). Columns marked with different letters differ significantly at the 1% probability level.

3.6. Yield Components

There were no significant interactions between the two factors for all yield components (Table **8**). The SSB treatments did not significantly affect panicle number in pot^{-1} or 1000-grain weight compared to the control. Filled grain panicle^{-1} in the *E. sichuanensis* and co-inoculation treatments (116.58-117.34) was significantly higher than the control (104.36). Only the *E. sichuanensis* treatment showed a higher filled-grain percentage than the control. The total grain yield in the *P. aeruginosa* treatment was equivalent to that of both the control and other bacterial treatments. However, the *E. sichuanensis* and co-inoculation treatments increased the total grain yield by 9.25-12.88% compared to the control.

Table 8. Panicle number pot^{-1}, 1000-grain weight, filled grains panicle^{-1}, filled grain percentage, and total grain yield pot^{-1} on rice plants treated with SSB under different water conditions.

Treatment	DF	Panicle number pot^{-1}	1000-grain weight (g)	Filled grains panicle^{-1}	Filled grain percentage (%)	Total grain yield (g pot^{-1})
Types of bacteria inoculation (A)			-	-	-	-
Control		43.75^{a} (41.20, 46.30)	21.56^{a} (21.07, 22.05)	104.36^{c} (99.38, 109.34)	63.11^{b} (61.00, 65.21)	92.19^{b} (86.34, 98.04)
P. aeruginosa		47.25^{a} (44.70, 49.80)	21.69^{a} (21.20, 22.18)	109.57^{bc} (104.59, 114.55)	63.50^{b} (61.39, 65.60)	96.54^{ab} (90.69, 102.39)
E. sichuanensis		45.63^{a} (43.08, 48.17)	21.82^{a} (21.34, 22.31)	116.58^{ab} (111.60, 121.56)	66.67^{a} (64.57, 68.78)	100.72^{a} (94.87, 106.57)
Co-inoculation		47.13^{a} (44.58, 49.67)	22.23^{a} (21.75, 22.72)	117.34^{a} (112.36, 122.33)	66.05^{ab} (63.94, 68.15)	104.06^{a} (98.21, 109.91)
Water conditions (B)			-	-	-	-
Water deficit		49.50^{a} (47.70, 51.30)	20.91^{b} (20.57, 21.26)	75.62^{b} (72.10, 79.14)	52.56^{b} (51.07, 54.05)	68.38^{b} (64.25, 72.52)
Flooded		42.38^{b} (40.57, 44.18)	22.74^{a} (22.40, 23.09)	148.30^{a} (144.78, 151.83)	77.11^{a} (75.62, 78.60)	128.37^{a} (124.23, 132.51)
F (A)	3	1.78ns	1.55^{ns}	6.61**	3.13*	3.35*
F (B)	1	33.79***	60.92***	920.48***	587.19***	454.83***
F (AxB)	3	0.31ns	0.50^{ns}	1.08^{ns}	0.32^{ns}	0.28^{ns}
CV (%)		7.55	3.04	6.05	4.42	8.09

Note: Means within column followed by the same letter are not significantly different by least significant difference, $p \geq 0.05$; * = Significant at $p \leq 0.05$; ** = Significant at $p \leq 0.01$; *** = Significant at $p \leq 0.001$; ns = Not significant. Values in parentheses represent the 95% confidence intervals.

For water conditions, water deficit conditions significantly reduced 1000-grain weight, filled grains panicle^{-1}, filled grain percentage, and total grain yield, except for panicle number pot^{-1}. Compared to flooded conditions, water-deficit conditions reduced the number of panicles per pot^{-1} by 14.38%.

4. DISCUSSION

Among the ten bacterial isolates, UPMC 1340, 1341, and 1342 exhibited the highest solubilization activity, followed closely by UPMC 248 (Table **3**). In fact, solubilizing minerals by bacteria predominantly occurs *via* mechanisms involving the production of organic acids [36]. Reportedly, these solubilization bacteria may produce organic acids that promote a decrease in pH and facilitate ligand attack on cationic sites, thereby breaking Al-O-Si or Mg-, Ca-, or MgO-Si bonds, while Si-O-Si linkages in quartz or amorphous silica are mainly disrupted by proton or hydroxyl ion polarization [37-39]. This assumption is strongly supported by a study by Pastore *et al*., who reported a negative correlation between pH and dissolved silicate concentration, whereas the amount of organic acid produced by soil bacteria was positively related to the rate of silicate solubilization [39]. Although the value of pH was not measured in the present study, the superior performance of UPMC 248, 1340, 1341, and 1342, compared to other isolates, suggests that they might produce higher amounts of organic acids to facilitate the dissolution of silicate minerals, thereby exhibiting strong potential as inoculants for enhancing available Si in soil.

Drought conditions not only impair plant growth but also influence the composition and activity of the surrounding microbial community. Hence, it is understandable that bacterial diversity can be negatively impacted by water deficiency. As shown in (Fig. **1**), increasing drought stress led to a decline in the overall bacterial population. Despite this, drought tolerance was observed in six of the ten bacterial isolates tested, including UPMC 248, 386, 383, 1340, 1341, and 1342. According to Abdelaal *et al*., bacterial isolates subjected to drought stress tend to produce higher levels of exopolysaccharides that play a critical role in protecting bacterial cells and enhancing their survival in water-limited environments [40]. As described by Forni *et al*., this mechanism involves exopolysaccharides that form hydrophilic biofilms to retain moisture around bacterial cells, thereby protecting them from desiccation [41]. The drought tolerance of the six isolates suggests their potential to improve plant growth under water-stress conditions.

Identification based on 16S rDNA gene sequencing revealed that all three isolates showed the highest sequence similarity to *Enterobacter sichuanensis*, as determined through comparison with the NCBI database. Several studies have indicated that the genus *Enterobacter* is considered a PGPB, as it exhibits plant growth-promoting traits [42, 43]. Chhetri *et al*. found that *E. sichuanensis* can serve as a promising biofertilizer to promote plant growth [44]. In our study, the superior silicate-solubilizing performance of these *E. sichuanensis* strains may indicate a highly efficient metabolic capacity for the production of organic acids [36]. Through the organic acid-mediated solubilization mechanism discussed earlier, these strains have been reported to mobilize Si from recalcitrant silicate minerals by converting it into plant-available monosilicic acid [16], improving Si availability in the rhizosphere and supporting their potential as effective microbial inoculants.

SSB are recognized as beneficial isolates that possess the ability to convert insoluble silicates in the soil to accessible forms for plant absorption. It is well established that organic acid production by SSB releases H+ ions, which promote cation exchange at the mineral surface by replacing cations in the silicate structure [15], leading to the release of soluble Si in the soil. According to Raturi *et al*., increased Si availability in the rhizosphere enhances Si uptake in plant tissue [16]. The present study is consistent with previous studies, as rice plants treated with bacterial treatments showed higher Si uptake than the control (Table **7**). In contrast to the typical pattern in which elevated soil-soluble Si persists, inoculation with *E. sichuanensis* did not significantly increase soil-soluble Si concentrations at any examined stage relative to the control (Table **5**), while significantly increasing Si content in rice plants (Table **7**). This pattern may be explained by rapid, direct coupling between soil Si availability and plant uptake. Although *E. sichuanensis* continuously maintained silicate solubilization throughout the growth stages, rice is a strong silicon-accumulating crop [15], which may promote rapid root uptake of the released Si and limit the accumulation of soluble Si in the soil [45]. The Si accumulation observed in the control can be attributed to the baseline pool of native soluble Si and indigenous SSB naturally present in the soil. As an efficient silicon accumulator, rice plants absorb this available Si supply [46]. In addition, the deposition of Si following root absorption is a critical factor under water stress. Irfan *et al.* highlighted that Si can improve stem rigidity under water scarcity [47]. To evaluate the role of Si uptake on stem strength, lignin content, which is a vital contributor to plant rigidity [48], was measured in the present study. The SSB treatments that increased Si absorption in rice tissue led to higher lignin content than the control (Table **7**). Evidence indicated that the increased uptake of Si ameliorated stem strength through stimulating lignin accumulation in herbaceous peony [49]. The current finding advances the understanding of the role of SSB inoculation in enhancing rice cell wall rigidity under water stress by promoting lignin biosynthesis. The elevated lignin accumulation may be attributed to greater bioavailability of monosilicic acid (H_4SiO_4), which enhances the phenylpropanoid pathway by upregulating enzymes, such as phenylalanine ammonia-lyase (PAL), while silica deposition within the cell wall matrix reinforces rice stem structure, increasing mechanical strength under drought stress [50, 51]. This structural reinforcement underscores a crucial Si–drought

interaction, where enhanced lignification and cell wall silicification may help maintain stem rigidity, enabling inoculated rice plants to tolerate physical stress and prevent vascular collapse under water scarcity [52].

Previous studies have shown that Si plays an integral role in improving growth and physiological traits in rice [53, 54]. In this study, although all SSB treatments significantly increased Si uptake in rice compared to the control, the *E. sichuanensis* and co-inoculation treatments significantly improved growth and physiological parameters, such as plant height, RWC, root length, and SPAD values, compared to the control (Tables **6** and **7**). Notably, only the co-inoculation treatments significantly increased leaf area, and the *E. sichuanensis* treatment was equivalent to the control. This suggests that Si benefits depend on effective utilization rather than uptake alone, with co-inoculation likely stimulating root development, photosynthesis, and nutrient acquisition through synergistic interactions, whereas single inoculations, despite increasing Si uptake, were insufficient to promote leaf expansion. For yield components, rice plants in the *E. sichuanensis* and co-inoculation treatments exhibited significant improvements in filled grain panicle^{-1} and total grain yield compared to the control, regardless of water conditions (Table **8**). This could be because the increased SPAD values under the *E. sichuanensis* and co-inoculation treatments contributed to higher filled grain panicle-1 and total grain yield. According to Monteoliva *et al.*, chlorophyll content enhances photosynthesis, thereby improving crop yield [55].

Water insufficiency during the flowering stage reduces grain number and spikelet fertility, resulting in significant yield losses [4]. In this study, water deficit conditions hindered rice growth, resulting in lower yield characteristics than under flooded conditions (Table **8**). Jarin *et al.* reported that water stress decreased SPAD values, RWC, and flag leaf area in rice, thus decreasing grain yield [56]. This aligns with our findings, as rice plants exhibited reductions in plant height, leaf area, RWC, root length, and SPAD values, leading to reduced total grain yield.

The results indicate that bacterial inoculation interacted significantly with water conditions for RWC and root length (Table **6**). The co-inoculation treatment combined with the flooded conditions promoted higher RWC and root elongation in rice plants than those under water deficiency without bacterial inoculation (Figs. **3** and **4**). Si deposition in epidermal cells, the cuticle, guard cells, and endodermal tissues may stimulate the formation of a silica double layer, lowering cuticular transpiration by increasing leaf and epidermal thickness and minimizing water loss during water stress [47]. Furthermore, Si is well known for improving plant water uptake by regulating root anatomy. The reason for this is likely due to Si-mediated modulation of polyamine and 1-aminocyclopropane-1-carboxylic acid concentrations, which strengthen endodermal silicification and suberization, enhancing root growth and, consequently, improving water uptake under drought [57]. In addition, increased Si uptake facilitated by SSB may promote endodermal suberization and continuous root growth, thereby helping preserve cell expansion and hydraulic conductivity and optimizing root water uptake under soil-drying conditions [53]. In the present study, rice cultivated under flooded conditions with higher water status and root development is readily apparent (Table **5**). According to Vadez *et al.*, soil conductance is substantially higher than root hydraulic conductance under water-sufficient soils and thus regulates the water flow sustaining transpiration [58], while the reduction of soil hydraulic conductivity occurring in soil drying becomes a dominant constraint rather than the root conductance, which is then associated with root shrinkage combined with loss of soil contact, further limiting water acquisition.

5. STUDY LIMITATIONS

The experiment was restricted to one rice variety, one location, and one growing season, while the use of pots may not adequately simulate field soil hydrodynamics. Moreover, soil pH, changes in the soil microbiota, and the biosafety aspects of *Pseudomonas aeruginosa* as a biofertilizer candidate were beyond the scope of this study. Silicon content was quantified only in stems at harvest, and the proposed mechanisms of silicon solubilization were inferred rather than directly demonstrated.

CONCLUSION

The present findings reveal that the *E. sichuanensis* and co-inoculation treatments improved plant height, RWC, SPAD values, lignin content, and Si uptake, thus enhancing filled grains panicle^{-1} and total grain yield in rice under both flooded and water-stressed conditions. Notably, co-inoculation increased root length under water-deficit conditions to levels comparable to those observed under flooded conditions. These results suggest that the *E. sichuanensis* and co-inoculation applications can alleviate the adverse impacts of water stress in rice, and may contribute to food security under climate change and population expansion. Nevertheless, some plant and soil parameters were restricted by the limitations of the pot experiment. Thus, it is suggested that future studies explore the field-scale application and formulation of these strains as biofertilizers.

AUTHORS' CONTRIBUTIONS

All authors: conceptualization and writing-original draft, result interpretation and manuscript preparation; M.H.H.: Methodology and formal analysis; M.M.Y., K.Z.A.T., Z.B., and H.Z.J.: Investigation, validation and supervision; M.H.H.: Experimental design, sampling and data analysis; M.M.Y., K.ZA.T., Z.B, and H.Z.J: Review and editing; M.M.Y.: funding acquisition. All authors reviewed and agreed on this final manuscript for submission and publication.

LIST OF ABBREVIATIONS

SSB = Silicate-solubilizing bacteria

PGPB = Plant growth-promoting bacteria
DOA = Department of Agriculture
DAS = Days after sowing
SI = Solubilization index
DAI = Days of incubation
TSB = Tryptic Soy Broth
PEG = Polyethylene glycol
OD = Optical density
RWC = Relative water content
NCBI = National Center for Biotechnology Information

RESEARCH INVOLVING PLANTS

Not applicable.

CONSENT FOR PUBLICATION

Not applicable.

AVAILABILITY OF DATA AND MATERIALS

The data that support the findings of this study are available from the corresponding author, [M.M.Y.], on special request.

FUNDING

This research was funded by Universiti Putra Malaysia (UPM) under the Putra Grant GP-IPS (GP-IPS/2023/9745000) (Vot No: 9745000).

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

All authors would like to thank Universiti Putra Malaysia (UPM) for providing financial support through the Putra Grant GP-IPS (GP-IPS/2023/9745000) under the project entitled “Enhancement of Growth and Development of MR297 Rice Variety by Silicate Solubilizing Bacteria under Different Water Conditions”. The authors would also like to thank Southeast Asian Regional Center for Graduate Study and Research in Agriculture (SEARCA) and the German Academic Exchange (DAAD) for awarding a DAAD-SEARCA scholarship to Mr. Ho Minh Hung to pursue his Master’s studies at the Faculty of Agriculture, Universiti Putra Malaysia.

REFERENCES

[1] Bin Rahman ANMR, Zhang J. Flood and drought tolerance in rice: opposite but may coexist. Food Energy Secur 2016; 5(2): 76-88. http://dx.doi.org/10.1002/fes3.79

[2] Sellami MH, Abi Saab MT, Albrizio R. Toward water-smart cereal production: a narrative review of agronomic, genetic, and digital innovations for water productivity and climate resilience. Front Plant Sci 2026; 17: 1826562. http://dx.doi.org/10.3389/fpls.2026.1826562

[3] Bwire D, Saito H, Sidle RC, Nishiwaki J. Water management and hydrological characteristics of paddy-rice fields under alternate wetting and drying irrigation practice as climate smart practice: a review. Agronomy (Basel) 2024; 14(7): 1421. http://dx.doi.org/10.3390/agronomy14071421

[4] Yang X, Wang B, Chen L, Li P, Cao C. The different influences of drought stress at the flowering stage on rice physiological traits, grain yield, and quality. Sci Rep 2019; 9(1): 3742. http://dx.doi.org/10.1038/s41598-019-40161-0

[5] Bin Rahman AR, Zhang J. Trends in rice research: 2030 and beyond. Food and Energy Security 2022; 12(2): 1-17. http://dx.doi.org/10.1002/fes3.390

[6] Zhang J, Zhang S, Cheng M, *et al*. Effect of drought on agronomic traits of rice and wheat: A meta-analysis. Int J Environ Res Public Health 2018; 15(5): 839. http://dx.doi.org/10.3390/ijerph15050839

[7] Mishra AK, Singh VP. A review of drought concepts. J Hydrol (Amst) 2010; 391(1-2): 202-16. http://dx.doi.org/10.1016/j.jhydrol.2010.07.012

[8] Berg A, Sheffield J. Climate change and drought: the soil moisture perspective. Curr Clim Change Rep 2018; 4(2): 180-91. http://dx.doi.org/10.1007/s40641-018-0095-0

[9] Cook BI, Mankin JS, Anchukaitis KJ. Climate change and drought: From past to future. Curr Clim Change Rep 2018; 4(2): 164-79. http://dx.doi.org/10.1007/s40641-018-0093-2

[10] Kompas T, Che TN, Grafton RQ. Global impacts of heat and water stress on food production and severe food insecurity. Sci Rep 2024; 14(1): 14398. http://dx.doi.org/10.1038/s41598-024-65274-z

[11] Ramírez-Olvera SM, Trejo-Téllez LI, Gómez-Merino FC, Ruíz-Posadas LM, Alcántar-González EG, Saucedo-Veloz C. Silicon stimulates plant growth and metabolism in rice plants under conventional and osmotic stress conditions. Plants 2021; 10(4): 777. http://dx.doi.org/10.3390/plants10040777

[12] Bist V, Niranjan A, Ranjan M, Lehri A, Seem K, Srivastava S. Silicon-solubilizing media and its implication for characterization of bacteria to mitigate biotic stress. Front Plant Sci 2020; 11: 28. http://dx.doi.org/10.3389/fpls.2020.00028

[13] Souri Z, Khanna K, Karimi N, Ahmad P. Silicon and plants: current knowledge and future prospects. J Plant Growth Regul 2021; 40(3): 906-25. http://dx.doi.org/10.1007/s00344-020-10172-7

[14] Das D, Ullah H, Tisarum R, Cha-um S, Datta A. Morpho-physiological responses of tropical rice to potassium and silicon fertilization under water-deficit stress. J Soil Sci Plant Nutr 2023; 23(1): 220-37. http://dx.doi.org/10.1007/s42729-021-00712-9

[15] Etesami H, Schaller J. Improving phosphorus availability to rice through silicon management in paddy soils: A review of the role of silicate-solubilizing bacteria. Rhizosphere 2023; 27: 100749. http://dx.doi.org/10.1016/j.rhisph.2023.100749

[16] Raturi G, Sharma Y, Rana V, *et al*. Exploration of silicate solubilizing bacteria for sustainable agriculture and silicon biogeochemical cycle. Plant Physiol Biochem 2021; 166: 827-38. http://dx.doi.org/10.1016/j.plaphy.2021.06.039

[17] Glick BR. Introduction to plant growth-promoting bacteria. Beneficial plant-bacterial interactions. Cham: Springer International Publishing 2020; pp. 1-37. http://dx.doi.org/10.1007/978-3-030-44368-9_1

[18] Sashidhar B, Podile AR. Mineral phosphate solubilization by rhizosphere bacteria and scope for manipulation of the direct oxidation pathway involving glucose dehydrogenase. J Appl Microbiol 2010; 109(1): 1-12. http://dx.doi.org/10.1111/j.1365-2672.2009.04654.x

[19] Chandrakala C, Voleti SR, Bandeppa S, Sunil Kumar N, Latha PC. Silicate solubilization and plant growth promoting potential of Rhizobium sp. isolated from rice rhizosphere. Silicon 2019; 11(6): 2895-906. http://dx.doi.org/10.1007/s12633-019-0079-2

[20] Hashemnejad F, Barin M, Khezri M, Ghoosta Y, Hammer EC.

Isolation and identification of insoluble zinc-solubilising bacteria and evaluation of their ability to solubilise various zinc minerals. J Soil Sci Plant Nutr 2021; 21(3): 2501-9.
http://dx.doi.org/10.1007/s42729-021-00540-x

[21] Devi R, Kaur T, Kour D, *et al*. Minerals solubilizing and mobilizing microbiomes: A sustainable approach for managing minerals' deficiency in agricultural soil. J Appl Microbiol 2022; 133(3): 1245-72.
http://dx.doi.org/10.1111/jam.15627

[22] Nguyen NK, Tran DV. Silicate solubilizing bacteria enhanced salt tolerance capacity and improved rice yield cultivated in salt-affect soil. Asia-Pac J Sci Technol 2023; 28(4): 12.
http://dx.doi.org/10.14456/apst.2023.52

[23] Vasanthi N, Saleena LM, Raj SA. Silica solubilization potential of certain bacterial species in the presence of different silicate minerals. Silicon 2018; 10(2): 267-75.
http://dx.doi.org/10.1007/s12633-016-9438-4

[24] Shabbir I, Samad MYA, Othman R, *et al*. Silicate solubilizing bacteria UPMSSB7, a potential biocontrol agent against white root rot disease pathogen of rubber tree. J Rubber Res Kuala Lumpur Malaysia 2020; 23(3): 227-35.
http://dx.doi.org/10.1007/s42464-020-00052-w

[25] Sharath S, Triveni S, Nagaraju Y, Latha PC, Vidyasagar B. The role of phyllosphere bacteria in improving cotton growth and yield under drought conditions. Frontiers in Agronomy 2021; 3: 680466.
http://dx.doi.org/10.3389/fagro.2021.680466

[26] Fazlil Ilahi WF, Abu Hassan NH, Che'Ya NN, *et al*. Alternate Wetting and Drying (AWD) on Rice Irrigation. Pertanika, J Trop Agric Sci 2022; 45(3): 649-61.
http://dx.doi.org/10.47836/pjtas.45.3.07

[27] Khanam M, Solaiman ARM, Rahman GKMM, Haque MM, Alam MS. Effect of plant growth promoting bacteria on growth and nutrient content of rice. Asian J Adv Agricult Res 2022; 19(2): 20-9.
http://dx.doi.org/10.9734/ajaar/2022/v19i230242

[28] Ali-Tan KZ, Radziah O, Halimi MS, Rahim KA, Abdullah MZ, Shamsuddin ZH. Growth and yield responses of rice cv. MR219 to rhizobial and plant growth-promoting rhizobacterial inoculations under different fertilizer-N rates. Bangladesh J Bot 2017; 46(1): 481-8.

[29] Zakaria NI, Berahim Z, Md Hatta MA, *et al*. Physiological and yield performance of commercial rice varieties under cyclic water stress in Malaysia. Sains Malays 2024; 53(7): 1589-603.
http://dx.doi.org/10.17576/jsm-2024-5307-09

[30] Ng LC, Anuar SNA, Jong JW, Elham MSH. Phytobeneficial and plant growth-promotion properties of silicon-solubilising rhizobacteria on the growth and control of rice sheath blight disease. Asian J Plant Sci 2016; 15(3-4): 92-100.
http://dx.doi.org/10.3923/ajps.2016.92.100

[31] Pereira HS, Korndörfer GH, Moura WF, Correa GF. Extractors of available silicon in slags and fertilizers. Rev Bras Ciênc Solo 2003; 27: 265-74.
http://dx.doi.org/10.1590/S0100-06832003000200007

[32] Hallmark CT, Wilding LP, Smeck NE. Silicon. In: Page AL, Ed. Methods of Soil Analysis: Part 2 Chemical and Microbiological Properties. (2nd.). Madison, WI: American Society of Agronomy 1982; pp. 263-73.
http://dx.doi.org/10.2134/agronmonogr9.2.2ed.c15

[33] Slatyer RO, Barrs HD. Modifications to the relative turgidity technique with notes on its significance as an index of the internal water status of leaves. In: Eckardt FE, Ed. Methodology of Plant Eco-Physiology: Proceedings of the Montpellier Symposium. Paris: UNESCO 1965; pp. 331-42.

[34] Suzuki S, Suzuki Y, Yamamoto N, Hattori T, Sakamoto M, Umezawa T. High-throughput determination of thioglycolic acid lignin from rice. Plant Biotechnol (Tsukuba) 2009; 26(3): 337-40.
http://dx.doi.org/10.5511/plantbiotechnology.26.337

[35] Wei-min D, Ke-qin Z, Bin-wu DUAN, *et al*. Rapid determination of silicon content in rice. Rice Sci 2005; 12(2): 145-7.

[36] Etesami H, Jeong BR. Biodissolution of silica by rhizospheric silicate-solubilizing bacteria. Silicon and nano-silicon in environmental stress management and crop quality improvement. Academic Press 2022; pp. 265-76.
http://dx.doi.org/10.1016/B978-0-323-91225-9.00020-0

[37] Sheng XF, Zhao F, He LY, Qiu G, Chen L. Isolation and characterization of silicate mineral-solubilizing *Bacillus globisporus* Q12 from the surfaces of weathered feldspar. Can J Microbiol 2008; 54(12): 1064-8.
http://dx.doi.org/10.1139/W08-089

[38] Ehrlich H, Demadis KD, Pokrovsky OS, Koutsoukos PG. Modern views on desilicification: biosilica and abiotic silica dissolution in natural and artificial environments. Chem Rev 2010; 110(8): 4656-89.
http://dx.doi.org/10.1021/cr900334y

[39] Pastore G, Kernchen S, Spohn M. Microbial solubilization of silicon and phosphorus from bedrock in relation to abundance of phosphorus-solubilizing bacteria in temperate forest soils. Soil Biol Biochem 2020; 151: 108050.
http://dx.doi.org/10.1016/j.soilbio.2020.108050

[40] Abdelaal K, AlKahtani M, Attia K, Hafez Y, Király L, Künstler A. The role of plant growth-promoting bacteria in alleviating the adverse effects of drought on plants. Biology (Basel) 2021; 10(6): 520.
http://dx.doi.org/10.3390/biology10060520

[41] Forni C, Duca D, Glick BR. Mechanisms of plant response to salt and drought stress and their alteration by rhizobacteria. Plant Soil 2017; 410(1-2): 335-56.
http://dx.doi.org/10.1007/s11104-016-3007-x

[42] Ogbo F, Okonkwo J. Some characteristics of a plant growth promoting Enterobacter sp. isolated from the roots of maize. Adv Microbiol 2012; 2(3): 368-74.
http://dx.doi.org/10.4236/aim.2012.23046

[43] Khalifa AYZ, Alsyeeh AM, Almalki MA, Saleh FA. Characterization of the plant growth promoting bacterium, Enterobacter cloacae MSR1, isolated from roots of non-nodulating Medicago sativa. Saudi J Biol Sci 2016; 23(1): 79-86.
http://dx.doi.org/10.1016/j.sjbs.2015.06.008

[44] Chhetri S, Sherpa MT, Sharma L. Characterization of plant growth promoting bacteria isolated from rhizosphere of tomato cultivated in Sikkim Himalaya and their potential use as biofertilizer. Sci Rep 2025; 15(1): 15558.
http://dx.doi.org/10.1038/s41598-025-98953-6

[45] Kang SM, Waqas M, Shahzad R, *et al*. Isolation and characterization of a novel silicate-solubilizing bacterial strain *Burkholderia eburnea* CS4-2 that promotes growth of japonica rice (*Oryza sativa* L. cv. Dongjin). Soil Sci Plant Nutr 2017; 63(3): 233-41.
http://dx.doi.org/10.1080/00380768.2017.1314829

[46] Ashokan A, Anand S, Aparna B, Mini V. Silicon uptake mechanism and its multidimensional influences on stress mitigation in rice (Oryza sativa L.). Int J Plant Soil Sci 2023; 35(4): 16-24.
http://dx.doi.org/10.9734/ijpss/2023/v35i42793

[47] Irfan M, Maqsood MA, Rehman H, *et al*. Silicon nutrition in plants under water-deficit conditions: overview and prospects. Water 2023; 15(4): 739.
http://dx.doi.org/10.3390/w15040739

[48] Hussain S, Iqbal N, Pang T, Naeem Khan M, Liu W, Yang W. Weak stem under shade reveals the lignin reduction behavior. J Integr Agric 2019; 18(3): 496-505.
http://dx.doi.org/10.1016/S2095-3119(18)62111-2

[49] Zhao D, Xu C, Luan Y, Shi W, Tang Y, Tao J. Silicon enhances stem strength by promoting lignin accumulation in herbaceous peony (Paeonia lactiflora Pall.). Int J Biol Macromol 2021; 190: 769-79.
http://dx.doi.org/10.1016/j.ijbiomac.2021.09.016

[50] Debona D, Rodrigues FA, Datnoff LE. Silicon's role in abiotic and biotic plant stresses. Annu Rev Phytopathol 2017; 55(1): 85-107.
http://dx.doi.org/10.1146/annurev-phyto-080516-035312

[51] Maharjan E, Mahawar S, Chauhan S, *et al*. Bioprospecting of novel silica solubilizing bacteria as bioinoculants for sustainable

silica management. Front Microbiol 2025; 16: 1556406. http://dx.doi.org/10.3389/fmicb.2025.1556406

[52] Ogedengbe O, Hunt J, Pang A, Gupta D. Silicon-mediated drought resilience mechanisms in crops: from physiology to molecular insights. Front Plant Sci 2026; 17: 1788106. http://dx.doi.org/10.3389/fpls.2026.1788106

[53] Pati S, Pal B, Badole S, Hazra GC, Mandal B. Effect of silicon fertilization on growth, yield, and nutrient uptake of rice. Commun Soil Sci Plant Anal 2016; 47(3): 284-90. http://dx.doi.org/10.1080/00103624.2015.1122797

[54] Dorairaj D, Ismail MR, Sinniah UR, Kar Ban T. Influence of silicon on growth, yield, and lodging resistance of MR219, a lowland rice of Malaysia. J Plant Nutr 2017; 40(8): 1111-24. http://dx.doi.org/10.1080/01904167.2016.1264420

[55] Monteoliva MI, Guzzo MC, Posada GA. Breeding for drought tolerance by monitoring chlorophyll content. Gene Technol 2021; 10(165): 10-35248. http://dx.doi.org/10.35248/2329-6682.21.10.165

[56] Jarin AS, Islam MM, Rahat A, Ahmed S, Ghosh P, Murata Y. Drought stress tolerance in rice: physiological and biochemical insights. Int J Plant Biol 2024; 15(3): 692-718. http://dx.doi.org/10.3390/ijpb15030051

[57] Wang M, Wang R, Mur LAJ, Ruan J, Shen Q, Guo S. Functions of silicon in plant drought stress responses. Hortic Res 2021; 8(1): 254. http://dx.doi.org/10.1038/s41438-021-00681-1

[58] Vadez V, Grondin A, Chenu K, *et al*. Crop traits and production under drought. Nat Rev Earth Environ 2024; 5(3): 211-25. http://dx.doi.org/10.1038/s43017-023-00514-w