RESEARCH ARTICLE

ISSN: 1874-3315 1

OPEN ACCES

Near-infrared Spectroscopy for Direct Investigation of Quality Compounds in *Capsicum* spp (*Capsicum*



Kusumiyati Kusumiyati^{1,2,*}, Eizo Taira³ and Yusuf Eka Maulana²

annuum L. and Capsicum frutescens L.)

¹Department of Agronomy, Faculty of Agriculture, Universitas Padjadjaran, Sumedang 45363, Indonesia ²Laboratory of Horticulture, Faculty of Agriculture, Universitas Padjadjaran, Sumedang 45363, Indonesia ³Department of Bioscience and Biotechnology, Faculty of Agriculture, University of the Ryukyus, 1 Senbaru, Nishihara, Okinawa, 903-0213, Japan

Abstract:

Introduction: Capsicum spp., commonly known as peppers or chili peppers, are essential horticultural crops. The unique flavors, pungency levels, and nutritional profiles of these peppers are determined by a complex composition of quality compounds such as antioxidant activity, capsaicin, and dihydrocapsaicin. Assessing and quantifying these compounds is crucial for both agricultural and industrial purposes.

Methods: This research provides fresh insight into the use of near-infrared spectroscopy (NIRS) to investigate quality compounds in *Capsicum* spp., such as antioxidant activity, capsaicin, and dihydrocapsaicin. The NIRS utilized wavelengths ranging from 702 to 1065 nm with intervals of 3 nm. The total sample size was 400, consisting of 7 varieties of *Capsicum* spp., including cayenne pepper 'Domba', 'Manik', and 'Ratuni', as well as red chili peppers 'CB2', 'Tanjung 2', 'Lingga', and 'Tanjung'. The sample was divided into a calibration (n=300) and a prediction set (n=100), and then modeling was carried out using statistical software.

Results: Accuracy values for antioxidant activity, capsaicin, and dihydrocapsaicin obtained were $R_{cal} \le 0.86$, $R_{pred} \le 0.79$, RMSEC ≤ 28.16, RMSEP ≤ 34.58, and RPD ≤ 1.66.

Conclusion: These findings suggested that NIRS is a rapid and precise method for investigating quality compounds in *Capsicum* spp., such as antioxidant activity, capsaicin, and dihydrocapsaicin.

Keywords: A non-destructive method, Powdered sample, Quality compound, Spectrometer, Horticultural crops, Antioxidants.

© 2024 The Author(s). Published by Bentham Open.

This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International Public License (CC-BY 4.0), a copy of which is available at: https://creativecommons.org/licenses/by/4.0/legalcode. This license permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.



Cite as: Kusumiyati K, Taira E, Maulana Y. Near-infrared Spectroscopy for Direct Investigation of Quality Compounds in Capsicum spp (Capsicum annuum L. and Capsicum frutescens L.). Open Agric J, 2024; 18: e18743315289226. http://dx.doi.org/10.2174/0118743315289226240122063646



Received: October 25, 2023 Revised: January 06, 2024 Accepted: January 11, 2024 Published: March 27, 2024



Send Orders for Reprints to reprints@benthamscience.net

1. INTRODUCTION

Capsicum spp. is a group of horticultural crops that are in high demand worldwide due to their taste, appearance, and quality. This group consists of cayenne and red chili peppers that rot quickly after harvest, similar to other horticultural products. The process of spoilage in horticultural products is caused by their high water

content. Furthermore, cayenne and red chili peppers have a water content of 80-90% [1] but can be processed into a powder to improve durability. The goal is to reduce the water content, improve durability, and maintain quality. The average water content in chili powder was reported to be around 11% [2] and is used extensively as an additive for food products.

Capsicum spp. has various nutrients and is a source of antioxidants. These antioxidants are very beneficial to human health because they help prevent oxidative stress in the body. Antioxidants are compounds that inhibit oxidation and counteract free radicals by binding to free radical atoms or atoms with unpaired electrons. Cayenne pepper contains antioxidants, such as vitamin C, carotenoids, and capsaicin [3, 4], which can be determined using an antioxidant activity test, such as the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method.

Capsicum spp. has a unique nutritional content that includes capsaicinoid group compounds such as capsaicin and dihydrocapsaicin, which is about 77-98% in chili [5]. Furthermore, capsaicinoid content is determined by the genetic nature of the plant, maturity level, environment, and harvest period. The capsaicin, dihydrocapsaicin, and total capsaicinoid levels in cayenne pepper 'DARL 210' fruit increase from 480 mg/100g to 930 mg/100g, 416 mg/100g to 913 mg/100g, and 896 mg/100g to 1,843 mg/100g, respectively, 30 to 105 days after flowering (DAF) [6]. Another research reported that cayenne pepper 'Kanchanaburi 07' had capsaicin and dihydrocapsaicin levels ranging from 272 mg/100g to 485 mg/100g and 126 mg/100g to 190 mg/100g, respectively [7].

Laboratory analysis of the water content, capsaicin, and dihydrocapsaicin is relatively more complex because it requires time and expertise. This led to the development of quality measurements of optical-based and multivariate agricultural products [8-15]. Optical-based quality measurement includes the use of near-infrared spectroscopy (NIRS) combined with partial least squares regression (PLSR) [16-19]. Furthermore, the benefits of using NIRS include real-time measurements, no special skills required, and the ability to analyze qualitatively and quantitatively [20-22]. This method can also measure multiple desired product qualities using only one calibration model and has been successfully used to predict various agricultural products [23-32]. Research on the development of the NIRS in chili is still being conducted. Aprilia et al. [33] and Rohaeti et al. [34] succeeded in classifying chili at different levels of maturity and variety, while Haughey et al. [35] and Kusumiyati et al. [36, 37] predicted some of the quality content in chili using NIRS. Previous research, however, developed the NIRS model with a limited number of samples and varieties. Developing a model with a larger number of samples and varieties will increase the accuracy. Therefore, an NIRS model on Capsicum spp. with more numbers and varieties is required.

The purpose of this research was to evaluate potential NIRS for direct investigation of quality compounds in *Capsicum* spp., such as antioxidant activity, capsicum, and dihydrocapsaicin. The model development involves the wet analysis results in the laboratory and the NIRS spectra data. Furthermore, the application of NIRS as a direct investigation of quality compounds in *Capsicum* spp. enables quality monitoring, sorting, and product grading. The results are expected to contribute to the advancement of NIRS research, particularly in the investigation of *Capsicum* spp.

2. MATERIALS AND METHODS

2.1. Sample Collection

Powdered samples from various *Capsicum* spp. varieties, consisting of cayenne pepper 'Domba', 'Ratuni', and 'Manik' as well as red chili pepper 'CB2', 'Tanjung 2', 'Lingga', and 'Tanjung', were used in this research. Each sample was weighed and 30 g was placed in a Petri dish with a 10 cm diameter [36]. A total of 400 samples were used, divided into a calibration (300) and a prediction set (100). Sample analysis was carried out at the Laboratory of Horticulture, Faculty of Agriculture, Universitas Padjadjaran, Indonesia.

2.2. NIR Spectra Analysis

Spectra measurements of each sample were carried out at five different points on the Petri dish, including the top, middle, bottom, right, and left. The average value of the five points was used to obtain the value of each sample. Furthermore, the NirVana AG410 (Integrated Spectronics Pty, Ltd, Australia) NIR spectrometer with a wavelength range of 702 to 1065 nm and 3 nm intervals was used, and the results were in the form of absorbance spectra.

2.3. Sample Extraction

The extraction steps for measuring antioxidant activity, capsaicin, and dihydrocapsaicin were similar. Each powder sample was weighed, and 0.05 g was placed in a 10 ml vial and mixed with 10 ml of methanol before being placed in a sonicator (Ultrasonic Cleaner BK-2000) for 20 minutes at 65°C. The extract was then transferred to a 10 ml volumetric flask, and methanol was added to the limit mark. Centrifugation was then carried out for 10 minutes at 4000 Rpm, and the filtrate was placed in a 10 ml vial for further analysis of antioxidant activity, capsaicin, and dihydrocapsaicin [37].

2.3.1. Antioxidant Activity Analysis

Antioxidant activity was measured using the DPPH method. The extract and the DPPH solution were placed in a 5 ml flask and stored for 30 minutes in a dark room. A UV-Vis spectrophotometer (Shimadzu UV mini-1240, Tokyo, Japan) was used to measure the absorbance value at a wavelength of 517 nm. The inhibition value of DPPH free radicals was calculated based on the percentage inhibition. Furthermore, the 50% inhibition concentration (IC $_{50}$) value was obtained from the relationship between the percentage inhibition and the solution concentration of each sample. The antioxidant activity level of the sample was expressed in mg/100g.

2.3.2. Capsaicin and Dihydrocapsaicin Measurement

Capsaicin and dihydrocapsaicin measurements were carried out using HPLC (Shimadzu, LC 20AT Prominence, Japan) with a 5 μ m C-18 (150x4.6 mm) column. The mobile phase used was acetonitrile and 0.1% orthophosphoric acid in a 60:40 ratio equipped with a UV-Vis detector with a wavelength of 222 nm. The extract was filtered with a 0.45- μ m filter and the injection volume was 20 μ L, with a

column temperature of 30°C. The difference between these two parameters was the standard and the retention time. The capsaicin and dihydrocapsaicin standards were used in the capsaicin and dihydrocapsaicin measurements, respectively. Furthermore, the flow rate used was 1mL/minute, and the capsaicin and dihydrocapsaicin standards were prepared in concentrations of 2, 4, 8, 16, 32, 64, and 128 ppm, and 1, 2, 4, 8, 16, 32, and 64 ppm, respectively. The capsaicin and dihydrocapsaicin levels of the sample were calculated in mg/100g.

2.4. Data Analysis

The NIR spectroscopy data analysis included two main stages, namely preprocessing and multivariate analysis. Preprocessing was applied to reduce the residual noise in the spectra data. The preprocessing methods used were standard normal variate (SNV) and second derivative Savitzky-Golay (d2a). Furthermore, data processing was carried out using the PLSR method. The Unscrambler X 10.4 (Camo Software USA, Oslo, Norway) was used to analyze the data on the calibration and prediction set. Model accuracy was assessed based on the coefficient of correlation in the calibration (R_{cal}) and prediction set (R_{pred}), root mean square error of the calibration set (RMSEC), root mean square error of the prediction set (RMSEP), and the ratio of prediction to deviation (RPD).

3. RESULTS AND DISCUSSION

3.1. Quality Compound Results

Fig. (1) shows reference data on seven varieties of *Capsicum* spp. Based on these data, cayenne pepper 'Manik' has the highest antioxidant activity (2142.82 mg/100g), while red chili pepper 'CB2' has the lowest antioxidant activity (8916.71 mg/100g). The IC_{50} value is inversely proportional to the antioxidant activity, with a lower IC_{50} value indicating higher antioxidant activity. The cayenne pepper 'Manik' has the highest capsaicin and dihydrocapsaicin content, followed by 'Domba' and

'Ratuni'. Furthermore, the capsaicin content in this study is higher than the dihydrocapsaicin content. About 80-90% of the capsaicinoid group consists of capsaicin and dihydrocapsaicin, with the capsaicin content and the ratio between the two compounds being $\pm 69\%$ and 1:1 or 2:1, respectively [38, 39]. These compounds are also abundant in varieties with high antioxidant activity. This is consistent with previous research on the antioxidant activity of chili pepper and its relationship with capsaicin and dihydrocapsaicin content [40].

Table 1 shows that the antioxidant activity, capsaicin, and dihydrocapsaicin values range from 194.86~mg/100g to 13137.23~mg/100g, 12.46~mg/100g to 1378.28~mg/100g, and 7.15~mg/100g to 355.54~mg/100g, respectively. Data variation increases the reliability of a model, and the diversity of samples in a dataset is observed from the minimum and maximum values. Furthermore, the standard deviation can be used to describe the data variation. The standard deviation for antioxidant activity, capsaicin, and dihydrocapsaicin is 2965.15~mg/100g, 309.24~mg/100g, and 99.54~mg/100g, respectively.

3.2. Prediction Model of Quality Compounds

Calibration models of various Capsicum varieties allow faster direct investigation of quality compounds. The calibration models for multiple varieties will be more efficient than models for individual varieties. Table 2 shows that the antioxidant activity with original spectra data produced the best accuracy with R_{cal} , RMSEC, R_{pred} , and RMSEP of 0.91, 1214.34, 0.91, and 1176.48, respectively. In capsaicin, the calibration model with d2a spectra produced the best results with $R_{\mbox{\tiny cal}}$ and RMSEC of 0.86 and 151.22, respectively. Meanwhile, the prediction set for different or unknown samples produced an accuracy of R_{pred} and RMSEP with 0.79 and 190.76, respectively. The best model for dihydrocapsaicin is generated from the original spectra data with R_{cal}, RMSEC, R_{pred} , and RMSEP of 0.95, 28.16, 0.93, and 34.58, respectively.

Table 1. Summary results for quality compounds in Capsicum spp.

Quality Compounds	Min	Max	Mean	SD
Antioxidant activity (mg/100g)	194.86	13137.23	5769.37	2965.15
Capsaicin (mg/100g)	12.46	1378.28	262.36	309.24
Dihydrocapsaicin (mg/100g)	7.15	355.54	91.70	99.54

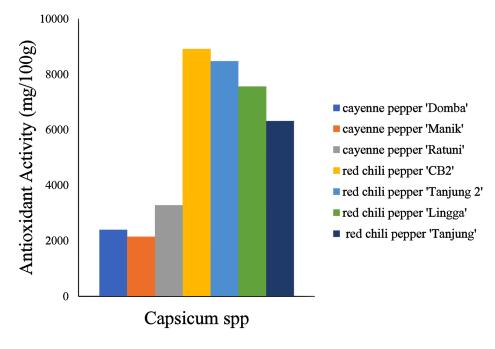
Table 2. Summary results for the accuracy of quality compounds in Capsicum spp. from NIRS.

Quality Compounds	Spectra Pre-treatment	$\mathbf{R}_{\mathrm{cal}}$	RMSEC	$\mathbf{R}_{ ext{pred}}$	RMSEP	RPD
Antioxidant activity	Original	0.91	1214.34	0.91	1176.48	2.53
	SNV	0.87	1422.36	0.90	1311.55	2.27
	d2a	0.84	1583.19	0.84	1573.16	1.89
Capsaicin	Original	0.87	145.87	0.78	195.93	1.61
	SNV	0.89	138.98	0.76	211.09	1.50
	d2a	0.86	151.22	0.79	190.76	1.66

(Table 2) contd.....

Quality Compounds	Spectra Pre-treatment	R _{cal}	RMSEC	R_{pred}	RMSEP	RPD
Dihydrocapsaicin	Original	0.95	28.16	0.93	34.58	2.92
	SNV	0.93	34.14	0.93	36.60	2.76
	d2a	0.93	34.52	0.891	45.70	2.21

Note: The best model is marked in bold.



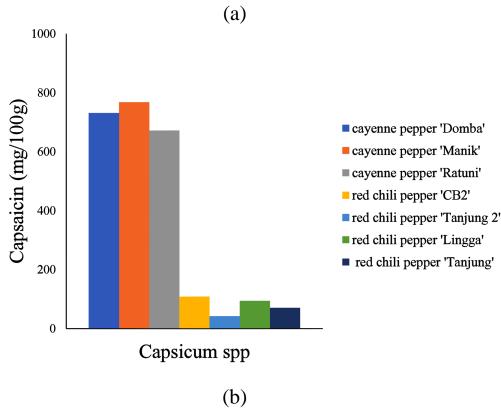


Fig. 1 contd.....

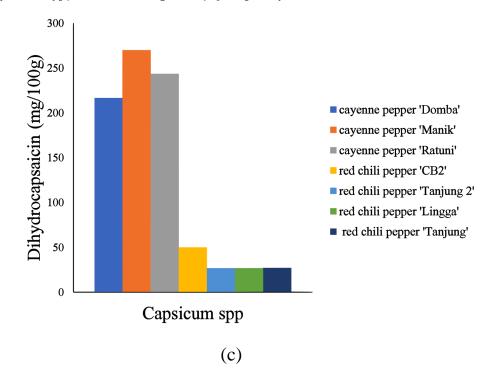


Fig. (1a-c). Mean of antioxidant activity, capsaicin, and dihydrocapsaicin for each Capsicum spp. variety.

Fig. (2) shows the correlation between reference data from quality compounds and NIRS predictions. The RPD values for antioxidant activity, capsaicin, and dihydrocapsaicin are 2.53, 1.66, and 2.92, respectively. The closer the data distribution is to the regression line, the more accurate the model is. Furthermore, $R_{\rm cal}/R_{\rm pred}$ approaching 1 and RMSEC/RMSEP approaching 0 indicates a strong correlation between spectroscopic

predictions and reference data [41, 42]. RPD values below 1.5 show poor prediction and discrepancy, while values between 1.5 and 2 suggest that the model can distinguish between low and high response variables. Additionally, an RPD value between 2 and 2.5 implies the possibility of making imprecise quantitative predictions, while a value between 2.5 and 3 or more indicates good to excellent prediction accuracy [32, 43].

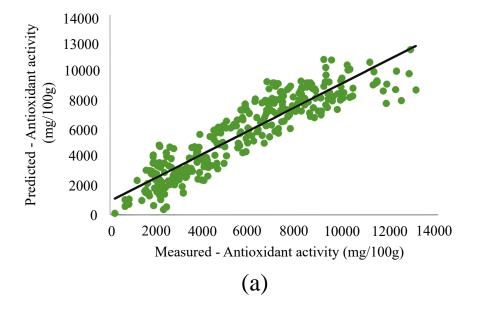


Fig. 4 contd.....

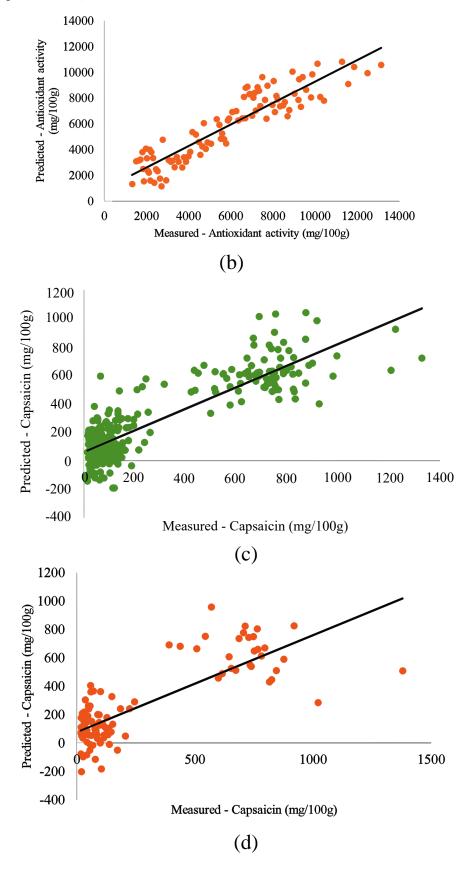


Fig. 4 contd.....

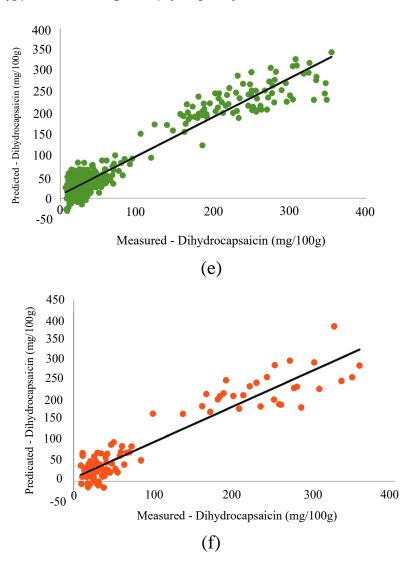


Fig. (2a-f). Scatter plots of the antioxidant activity, capsaicin, and dihydrocapsaicin for each variety of Capsicum spp, with green and orange indicating the calibration and prediction set, respectively.

CONCLUSION

The use of NIRS for a direct investigation of quality compounds, such as antioxidant activity, capsaicin, and dihydrocapsaicin in Capsicum spp. yielded satisfactory results. High $R_{\rm cal}$, $R_{\rm pred}$, and RPD values were obtained for each quality compound, including antioxidant activity (0.91, 0.91, and 2.53), capsaicin (0.86, 0.79, and 1.66), and dihydrocapsaicin (0.95, 0.93, and 2.92). Based on the modeling development results, the model was categorized as accurate and had a high potential to be used in quantitatively assessing antioxidant activity, capsaicin, and dihydrocapsaicin in Capsicum spp.

AUTHORS' CONTRIBUTIONS

All the authors participated in the study design, practical work, and writing the manuscript and approved the final manuscript.

CONSENT FOR PUBLICATION

Not applicable.

AVAILABILITY OF DATA AND MATERIALS

The data presented in this study are available on request from the corresponding author [K.K].

FUNDING

None.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

The authors are grateful to Ine Elisa Putri, and Yuda Hadiwijaya for their support, assistance, and chemical acquisition for the research.

REFERENCES

- [1] Anoraga SB, Sabarisman I, Ainuri M. Effect of different pretreatments on dried chilli (Capsicum annum L.) quality. IOP Conf Ser Earth Environ Sci 2018; 131: 012014. http://dx.doi.org/10.1088/1755-1315/131/1/012014
- [2] Toontom N, Meenune M, Posri W, et al. Effect of drying method on physical and chemical quality, hotness and volatile flavour characteristics of dried chilli. Int Food Res J 2012; 19: 1023-31.
- [3] Sun T, Xu Z, Wu CT, Janes M, Prinyawiwatkul W, No HK. Antioxidant activities of different colored sweet bell peppers (Capsicum annuum L.). J Food Sci 2007; 72(2): S98-S102. http://dx.doi.org/10.1111/j.1750-3841.2006.00245.x PMID: 17995862
- [4] Zimmer AR, Leonardi B, Miron D, Schapoval E, Oliveira JR, Gosmann G. Antioxidant and anti-inflammatory properties of Capsicum baccatum: From traditional use to scientific approach. J Ethnopharmacol 2012; 139(1): 228-33. http://dx.doi.org/10.1016/j.jep.2011.11.005 PMID: 22100562
- [5] Do T, Lacková Z, Adam V, et al. Determination of the content of capsaicin and dihydrocapsaicin in twelve varieties of chilli peppers using liquid chromatography with UV/VIS detection. MendelNet 2017; 2017: 861-6.
- [6] Pandey H, Deendayal , Pandey V, Pant T, Ahmed Z. Variation of capsaicinoids in chilli (*Capsicum frutescens* L.) cultivars with the maturity of fruits in middle hill conditions of western Himalayas. International Journal of Green Pharmacy 2010; 4(3): 178-82. http://dx.doi.org/10.4103/0973-8258.69177
- [7] Kraikruan W, Sukprakarn S, Mongkolporn O, et al. Capsaicin and dihydrocapsaicin contents of Thai chili cultivars. Witthayasan Kasetsat Witthayasat 2008; 42: 611-6.
- [8] Suhandy D, Yulia M. Chemometric quantification of peaberry coffee in blends using UV-visible spectroscopy and partial least squares regression. AIP Conference Proceedings. 2018; p. 060010.
- [9] Kusumiyati HY, Hadiwijaya Y, Elisa Putri I. Determination of water content of intact sapodilla using near infrared spectroscopy. IOP Conf Ser Earth Environ Sci 2018; 207: 012047. http://dx.doi.org/10.1088/1755-1315/207/1/012047
- [10] Schoot M, Kapper C, van Kollenburg GH, et al. Investigating the need for preprocessing of near-infrared spectroscopic data as a function of sample size. Chemom Intell Lab Syst 2020; 204: 104105.
 - http://dx.doi.org/10.1016/j.chemolab.2020.104105
- [11] Kasampalis DS, Tsouvaltzis P, Ntouros K. Nutritional composition changes in bell pepper as affected by the ripening stage of fruits at harvest or postharvest storage and assessed non-destructively. J Sci Food Agric 2022; 102(1): 445-54. PMID: 34143899
- [12] Suhandy D, Yulia M, Kusumiyati . The authentication of peaberry and civet ground roasted robusta coffee using UV-visible spectroscopy and PLS-DA method with two different particle sizes. IOP Conf Ser Earth Environ Sci 2019; 258: 012043. http://dx.doi.org/10.1088/1755-1315/258/1/012043
- [13] Kusumiyati , Munawar AA, Suhandy D. Fast and contactless assessment of intact mango fruit quality attributes using near infrared spectroscopy (NIRS). IOP Conference Series: Earth and Environmental Science. 21-22 September 2020; Banda Aceh, Indonesia. 2021.
- [14] Munawar AA, Devianti , Satriyo P, Bahari SA. Near infrared spectroscopy: Rapid and simultaneous approach to predict the fixed carbon, volatile matter and ash contents in biochar produced from agricultural residues. Acta Technol Agric 2022; 25(1): 1-6. http://dx.doi.org/10.2478/ata-2022-0001
- [15] Asikin Y, Kusumiyati , Shikanai T, Wada K. Volatile aroma components and MS-based electronic nose profiles of dogfruit (Pithecellobium jiringa) and stink bean (Parkia speciosa). J Adv Res 2018; 9: 79-85. http://dx.doi.org/10.1016/j.jare.2017.11.003 PMID: 30046489
- [16] Kusumiyati HY, Hadiwijaya Y, Putri IE, Munawar AA. Multi-

- product calibration model for soluble solids and water content quantification in Cucurbitaceae family, using visible/near-infrared spectroscopy. Heliyon 2021; 7(8): e07677.
- http://dx.doi.org/10.1016/j.heliyon.2021.e07677 PMID: 34401571
- [17] Kusumiyati MS, Mubarok S, Sutari W, et al. Non-destructive method for predicting sapodilla fruit quality using near infrared spectroscopy. IOP Conf Ser Earth Environ Sci 2019; 334(1): 012045. http://dx.doi.org/10.1088/1755-1315/334/1/012045
- [18] Kusumiyati MS, Mubarok S, Sutari W, Hadiwijaya Y. Application of spectra pre-treatments on firmness assessment of intact sapodilla using vis-nir spectroscopy. IOP Conf Ser Earth Environ Sci 2021; 644(1): 012001. http://dx.doi.org/10.1088/1755-1315/644/1/012001
- [19] Kusumiyati HY, Hadiwijaya Y, Putri IE, Mubarok S, Hamdani JS. Rapid and non-destructive prediction of total soluble solids of guava fruits at various storage periods using handheld nearinfrared instrument. IOP Conf Ser Earth Environ Sci 2020; 458(1): 012022.
- http://dx.doi.org/10.1088/1755-1315/458/1/012022

 [20] Kusumiyati HY, Hadiwijaya Y, Putri IE, Mubarok S. Water content prediction of 'crystal' guava using visible-near infrared spectroscopy and chemometrics approach. IOP Conf Ser Earth Environ Sci 2019; 393(1): 012099.

 http://dx.doi.org/10.1088/1755-1315/393/1/012099
- [21] Kusumiyati K, Hadiwijaya Y, Suhandy D, Munawar AA. Prediction of water content and soluble solids content of 'manalagi' apples using near infrared spectroscopy. IOP Conf Ser Earth Environ Sci 2021; 922(1): 012062. http://dx.doi.org/10.1088/1755-1315/922/1/012062
- [22] Legner R, Voigt M, Servatius C, Klein J, Hambitzer A, Jaeger M. A four-level maturity index for hot peppers (*Capsicum annum*) using non-invasive automated mobile raman spectroscopy for on-site testing. Appl Sci 2021; 11(4): 1614. http://dx.doi.org/10.3390/app11041614
- [23] Shao Y, He Y. Nondestructive measurement of acidity of strawberry using Vis/NIR spectroscopy. Int J Food Prop 2008; 11(1): 102-11. http://dx.doi.org/10.1080/10942910701257057
- [24] Li H, Zhang M, Shen M, et al. Effect of ambient temperature on the model stability of handheld devices for predicting the apple soluble solids content. Eur J Agron 2022; 133: 126430. http://dx.doi.org/10.1016/j.eja.2021.126430
- [25] Walsh KB, Blasco J, Zude-Sasse M, Sun X. Visible-NIR 'point' spectroscopy in postharvest fruit and vegetable assessment: The science behind three decades of commercial use. Postharvest Biol Technol 2020; 168: 111246. http://dx.doi.org/10.1016/j.postharvbio.2020.111246
- [26] Beghi R, Giovanelli G, Malegori C, Giovenzana V, Guidetti R. Testing of a VIS-NIR system for the monitoring of long-term apple storage. Food Bioprocess Technol 2014; 7(7): 2134-43. http://dx.doi.org/10.1007/s11947-014-1294-x
- [27] Kusumiyati K, Hadiwijaya Y, Sutari W, Arip Munawar A. Global model for in-field monitoring of sugar content and color of melon pulp with comparative regression approach. AIMS Agric Food 2022; 7(2): 312-25. http://dx.doi.org/10.3934/agrfood.2022020
- [28] Cozzolino D, Cynkar WU, Shah N, Smith P. Multivariate data analysis applied to spectroscopy: Potential application to juice and fruit quality. Food Res Int 2011; 44(7): 1888-96. http://dx.doi.org/10.1016/j.foodres.2011.01.041
- [29] Genis HE, Durna S, Boyaci IH. Determination of green pea and spinach adulteration in pistachio nuts using NIR spectroscopy. Lebensm Wiss Technol 2021; 136: 110008. http://dx.doi.org/10.1016/j.lwt.2020.110008
- [30] Zhang H, Duan Z, Li Y, et al. Vis/NIR reflectance spectroscopy for hybrid rice variety identification and chlorophyll content evaluation for different nitrogen fertilizer levels. R Soc Open Sci 2019; 6(10): 191132. http://dx.doi.org/10.1098/rsos.191132 PMID: 31824720

- [31] Fan S, Wang Q, Tian X, et al. Non-destructive evaluation of soluble solids content of apples using a developed portable Vis/NIR device. Biosyst Eng 2020; 193: 138-48. http://dx.doi.org/10.1016/j.biosystemseng.2020.02.017
- [32] Saeys W, Mouazen AM, Ramon H. Potential for onsite and online analysis of pig manure using visible and near infrared reflectance spectroscopy. Biosyst Eng 2005; 91(4): 393-402. http://dx.doi.org/10.1016/j.biosystemseng.2005.05.001
- [33] Aprilia YI, Khuriyati N, Sukartiko AC. Classification of chili powder (*Capsicum annuum* L.) antioxidant activity based on near infrared spectra. Food Res 2021; 5(S2): 51-6. http://dx.doi.org/10.26656/fr.2017.5(S2).008
- [34] Rohaeti E, Muzayanah K, Septaningsih DA, Rafi M. Fast analytical method for authentication of chili powder from synthetic dyes using uv-vis spectroscopy in combination with chemometrics. Indones J Chem 2019; 19(3): 668-74. http://dx.doi.org/10.22146/ijc.36297
- [35] Haughey SA, Galvin-King P, Ho YC, Bell SEJ, Elliott CT. The feasibility of using near infrared and Raman spectroscopic techniques to detect fraudulent adulteration of chili powders with Sudan dye. Food Control 2015; 48: 75-83. http://dx.doi.org/10.1016/j.foodcont.2014.03.047
- [36] Kusumiyati K, Putri IE, Munawar AA, et al. A data fusion model to merge the spectra data of intact and powdered cayenne pepper for the fast inspection of antioxidant properties. Sustainability 2022: 14: 1-11.
- [37] Kusumiyati K, Putri IE, Hamdani JS, Suhandy D. Real-time detection of the nutritional compounds in green 'Ratuni UNPAD'

- cayenne pepper. Horticulturae 2022; 8(6): 554. http://dx.doi.org/10.3390/horticulturae8060554
- [38] Hayman M, Kam PCA. Capsaicin: A review of its pharmacology and clinical applications. Curr Anaesth Crit Care 2008; 19(5-6): 338-43.
 - http://dx.doi.org/10.1016/j.cacc.2008.07.003
- [39] Rajput JC, Parulekar YR. Handbook of Vegetable Science and Technology: Production, Composition, Storage and Processing. New York: Marcel Dekker 1998.
- [40] Hamed M, Kalita D, Bartolo ME, Jayanty SS. Capsaicinoids, polyphenols and antioxidant activities of *Capsicum annuum*: Comparative study of the effect of ripening stage and cooking methods. Antioxidants 2019; 8(9): 364. http://dx.doi.org/10.3390/antiox8090364 PMID: 31480665
- [41] Awasthi S, Kumar R, Devanathan A, Acharya R, Rai AK. Multivariate methods for analysis of environmental reference materials using laser-induced breakdown spectroscopy. Anal Chem Res 2017; 12: 10-6. http://dx.doi.org/10.1016/j.ancr.2017.01.001
- [42] Kusumiyati K, Putri IE. Comparison of color spectrophotometer and Vis/NIR spectroscopy on assessing natural pigments of cucumber applied with different ethephon concentrations. Heliyon 2023; 9(12): e22564.
 - http://dx.doi.org/10.1016/j.heliyon.2023.e22564 PMID: 38125485

 3] Nicolai BM, Beullens K, Bobelyn E, et al. Nondestructive measurement of fruit and vegetable quality by means of NIR
- measurement of fruit and vegetable quality by means of NIR spectroscopy: A review. Postharvest Biol Technol 2007; 46(2): 99-118.
 - http://dx.doi.org/10.1016/j.postharvbio.2007.06.024